



Cells in focus

The renal connecting tubule: Resolved and unresolved issues in Ca^{2+} transport

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ABSTRACT

The renal connecting tubule (CNT) localizes to the distal part of the nephron between the distal convoluted tubule and the collecting duct, and consists of two different cell types: segment-specific and intercalated cells. The former reabsorb water (H_2O), sodium (Na^+) and calcium (Ca^{2+}) ions to the blood compartment, while secreting potassium ions (K^+) into the pro-urine. The latter cells contribute to the renal control of the acid-base balance. Several factors and hormones tightly regulate these transport processes. Although the CNT reabsorbs only $\sim 15\%$ of filtered Ca^{2+} load, this segment is finally decisive for the amount of Ca^{2+} that appears in the urine. Impaired Ca^{2+} transport across CNT can provoke severe urinary Ca^{2+} excretion, called hypercalciuria. This review mainly focuses on the activity, abundance and expression of the epithelial Ca^{2+} channel named Transient Receptor Potential Vanilloid 5 (TRPV5) that is the gatekeeper of active Ca^{2+} reabsorption in the CNT.

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Cell facts

- The renal connecting tubule is located in the distal part of the nephron and consists of segment-specific and intercalated cells.
- The segment-specific cells finely regulate the reabsorption of H_2O , Na^+ , K^+ and Ca^{2+} .
- The segment-specific cells express TRPV5 which facilitates the rate-limiting step in active transcellular Ca^{2+} transport.

1. Introduction

The kidneys control the maintenance of body ion balance and arterial blood pressure. Blood is filtrated by the glomeruli and the electrolytes are reabsorbed from the pro-urine along the various segments constituting the nephron. The distal part of the nephron that comprises the distal convoluted tubule (DCT), the connecting duct (CNT) and the collecting duct (CD) achieves the final adjustment of renal ion excretion which is regulated by various hormones.

The CNT displays different morphologic characteristics. While the CNT is short and directly linked to the CD in the superficial cortex, it is longer and merges with other CNT segments in the

juxtamedullary region to form the “arcades” before flowing into the cortical collecting duct (Fig. 1A). The epithelium of CNT is composed of two different cell types: intercalated and non-intercalated cells that are intermingled with one another at a ratio of approximately 1:2. The nomenclature of these cells is not yet unambiguously defined. The non-intercalated cells have been named CNT cells (Seldin and Giebisch, 1992). However, the term “CNT cells” is confusing since intercalated cells are also CNT cells, i.e. cells originating from the CNT. Alternatively, authors (Verrey, 2001; Ronzaud et al., 2007; Wang et al., 2010) proposed the term “principal cells” to characterize the non-intercalated cells of the aldosterone-sensitive distal part of the nephron, composed of DCT, CNT and CD. However, in a strict sense, the term “principal cells” refers only to the non-intercalated cells of the CD (Seldin and Giebisch, 1992). Thus, in this review, we prefer to use the term “segment-specific cells” to describe the non-intercalated cells of the CNT (Loffing and Korbmayer, 2009).

Segment-specific cells of the CNT, which have a polygonal shape, express many transporters involved in transcellular ion transport. They actively transport H_2O , Na^+ , K^+ and Ca^{2+} (Fig. 1B). Intercalated cells that have a rounded shape regulate the urinary acidification. Both cell types contain a high density of mitochondria (Fig. 1C and D).

2. CNT origin

The embryological origin of CNT is not entirely clear. It has been proposed that it originates from the nephrogenic blastema (Neiss and Klehn, 1981) or from the branching ureteric bud (Howie et al., 1993). However, others suggest that the CNT develops by mutual induction processes initiated from adjoining segments. This last

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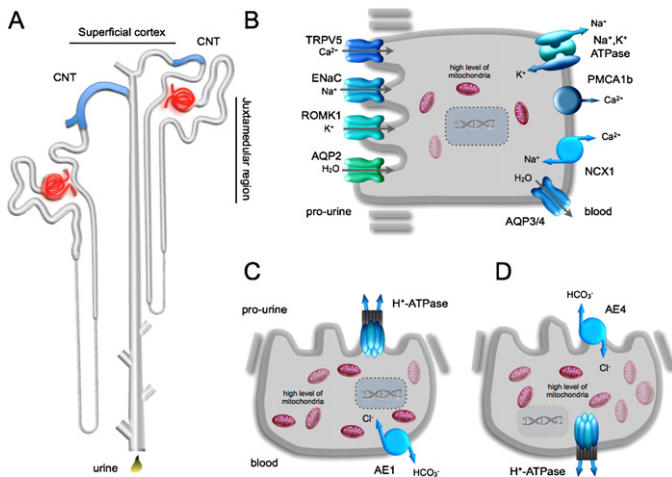


Fig. 1. The morphologic characteristics and cell types of the CNT. (A) The CNT (in blue) is located to the distal part of the nephron and consists of segment-specific and intercalated cells. While the CNT is short in the superficial cortex, it is longer in the juxtamedullary region. (B) Segment-specific cells of CNT are characterized by different transporters: (1) in the apical side: TRPV5 mediates the influx of Ca²⁺, ENaC mediates the influx of Na⁺, ROMK1 mediates the efflux of K⁺, AQP2 permits H₂O transport inside cell; (2) in the basolateral side: Ca²⁺ is extruded by the concerted action of the PMCA1b pump and the NCX1 exchanger, Na⁺ is extruded by the Na⁺/K⁺-ATPase which also permits the entry of K⁺, H₂O is extruded in the blood compartment by the AQP3 or AQP4. (C) Intercalated type A cells apically secrete H⁺ in the pro-urine compartment by a H⁺-ATPase and basolaterally extrudes HCO₃⁻ in the blood compartment by an Cl⁻/HCO₃⁻ exchanger (AE1). (D) Intercalated type B cells apically secrete HCO₃⁻ in the pro-urine compartment by an Cl⁻/HCO₃⁻ exchanger (AE4) and basolaterally extrudes H⁺ in the blood compartment by a H⁺-ATPase.

hypothesis is consistent with the fact that the CNT shares characteristics of both the DCT and CD. Like the DCT, the CNT expresses TRPV5 and like the CD, the CNT has numerous intercalated cells (Schmitt et al., 1999).

3. Function

3.1. Segment-specific cells of CNT

3.1.1. H₂O transport

Vasopressin increases the H₂O reabsorption in the CNT by binding to the basolateral V₂ receptor that stimulates a cyclic adenosine monophosphate and protein kinase A (cAMP-PKA)-dependent signaling pathway. This ultimately leads to the apical translocation and activation of the water channel aquaporin 2 (AQP2). H₂O is finally transported at the basolateral side by aquaporin 3 (AQP3) and 4 (AQP4) channels (Coleman et al., 2000).

3.1.2. Na⁺ transport

Active Na⁺ transport in the CNT is a two-step process that involves Na⁺ uptake by the apical ENaC and Na⁺ extrusion by the basolateral Na⁺ pump Na⁺/K⁺-ATPase. It is stimulated by several hormones including vasopressin (Bugaj et al., 2009), insulin (Markadieu et al., 2009) and aldosterone (Verrey et al., 2008). Aldosterone binds to the mineralocorticoid receptor and increases activity and expression of Na⁺ transporters and thus stimulates the transepithelial Na⁺ reabsorption. The existence of genetic disorders associated with a loss-of-function or hyperactivity of ENaC, leading to renal salt wasting syndromes and arterial hypertension, respectively, emphasizes that ENaC is the rate-limiting step of active Na⁺ transport (Loffing and Korbacher, 2009).

3.1.3. K⁺ transport

Segment-specific cells are also involved in the regulation of K⁺ secretion. K⁺ entry into the CNT cell is facilitated by

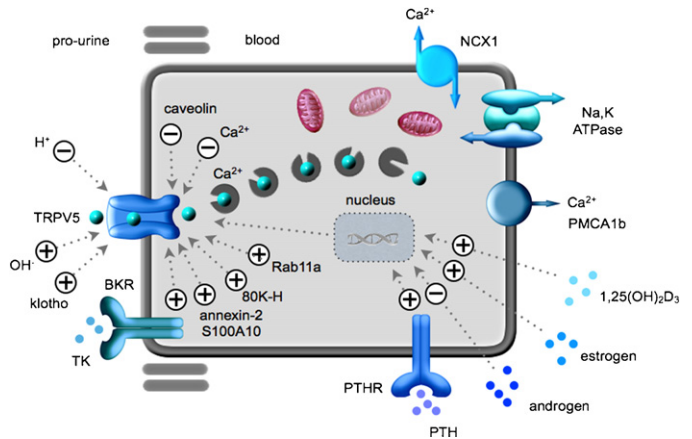


Fig. 2. Ca²⁺ transport across segment-specific cells of CNT. TRPV5 mediates the apical entry of Ca²⁺ inside the cell. In the urinary compartment, acidic conditions (H⁺) decrease the activity of TRPV5. Klotho directly interacts with TRPV5 and tissue kallikrein binds to the bradykinin receptor (BKR) located to the apical membrane. Both klotho and tissue kallikrein (TK) increase the apical abundance of TRPV5. Intracellularly, Ca²⁺ is buffered by calbindin-D_{28K} and is shuttled to the basolateral membrane that contains the NCX1 exchanger and the PMCA1b pump. Free cytosolic Ca²⁺ decreases the open probability of TRPV5 while 80K-H increases its activity. Rab11a and the annexin-2-S100A complex increase the apical insertion of TRPV5 while caveolin decrease its plasma membrane abundance. In the blood compartment, PTH, 1,25(OH)₂D₃ and estrogen increase the transcription and protein expression of TRPV5 while androgen inhibits its level of expression.

the Na⁺-K⁺/ATPase located in the basolateral membrane. Subsequently, accumulated K⁺ ions are released in the pro-urine via the renal outer medulla potassium channel ROMK1. The main factors regulating K⁺ secretion are the dietary K⁺ intake and aldosterone (Frindt et al., 2009).

3.1.4. Ca²⁺ transport

Ca²⁺ plays a fundamental role in many cellular processes and its extracellular concentration is controlled by the kidney, intestine and bone in a concerted fashion. In kidney, the late part of DCT and the CNT are the site of active Ca²⁺ transport and precisely regulate Ca²⁺ reabsorption. Although these distal segment reabsorb only ~15% of the Ca²⁺ filtered, they are responsible for the final adjustment of urinary Ca²⁺ excretion and are tightly regulated by several hormones. Ca²⁺ enters the CNT cell through TRPV5 (Hoenderop et al., 1999) and is extruded into the blood compartment by the concerted action of the plasma membrane Ca²⁺ pump (PMCA1b) and the Na⁺/Ca²⁺ exchanger (NCX1). TRPV5 knockout (TRPV5^{-/-}) mice exhibit severe urinary Ca²⁺ wasting, providing evidence for a gatekeeper role of TRPV5 in active renal Ca²⁺ transport (Hoenderop et al., 2003). *In vivo* micropuncture experiments confirmed that Ca²⁺ reabsorption is indeed impaired in the CNT. Moreover, TRPV5^{-/-} mice displayed phosphaturia, compensatory intestinal Ca²⁺ hyperabsorption, reduced bone thickness, polyuria and urinary acidification (Hoenderop et al., 2003). Intracellular Ca²⁺ concentration is kept low with respect to the extracellular fluid. Calbindin-D_{28K}, a Ca²⁺-binding protein, appears to shuttle Ca²⁺ from the TRPV5 channel to the basolateral Ca²⁺ transporters. Simultaneously, calbindin-D_{28K} buffers Ca²⁺ during high Ca²⁺ influx providing protection against toxic high cytosolic Ca²⁺ concentrations (Lambers et al., 2006).

The mechanisms by which TRPV5 is regulated can be classified in the following categories: (i) increase in TRPV5 protein expression; (ii) plasma membrane insertion of TRPV5; (iii) direct TRPV5 channel activation; and (iv) plasma membrane TRPV5 retrieval (Fig. 2 and Table 1). Calcitropic hormones, such as vitamin D₃ and parathyroid hormone (PTH), increase the protein expression of TRPV5 (Boros et al., 2009). The active form of vitamin D₃, 1,25-

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