

#### Contents lists available at ScienceDirect

## Nitric Oxide

journal homepage: www.elsevier.com/locate/yniox



#### Review

Sub-proteome S-nitrosylation analysis in *Brassica juncea* hints at the regulation of Brassicaceae specific as well as other vital metabolic pathway(s) by nitric oxide and suggests post-translational modifications cross-talk



Ankita Sehrawat, Renu Deswal \*

Molecular Plant Physiology and Proteomics Laboratory, Department of Botany, University of Delhi, Delhi 110007, India

#### ARTICLE INFO

#### Article history: Received 1 April 2014 Revised 21 August 2014 Available online 28 August 2014

Keywords: Nitric oxide Thiol pool S-nitrosylation Cold stress Brassica juncea

#### ABSTRACT

Abiotic stress affects the normal physiology of the plants and results in crop loss. Brassica juncea is an oil yielding crop affected by abiotic stress. In future, over 30% yield loss by abiotic stress is predicted in India. Understanding the mechanism of plant response to stress would help in developing stress tolerant crops. Nitric oxide (NO) is now viewed as a remarkably important signaling molecule, involved in regulating stress responses. S-Nitrosylation is a NO based post-translational modification (PTM), linked with the regulation of many physiologically relevant targets. In the last decade, over 700 functionally varied S-nitrosylated proteins were identified, which suggested broad-spectrum regulation. To understand the physiological significance of S-nitrosylation, it was analyzed in cold stress, Functional categorization and validation of some of the B. juncea S-nitrosylated targets, suggested that NO produced during stress regulates cellular detoxification by modulating enzymes of ascorbate glutathione cycle, superoxide dismutase, glutathione S-transferase and glyoxalase I by S-nitrosylation in crude, ribulose-1,5bisphosphate carboxylase/oxygenase (RuBisCO) depleted and apoplastic fractions. Interestingly, S-nitrosylation of enzymes associated with glucosinolate hydrolysis pathway, suggests a novel regulation of this Brassicaceae specific pathway by NO. Moreover, identification of enzymes of Glycolysis and Calvin cycle in crude and RuBisCO depleted fractions showed the regulation of metabolic as well as photosynthetic pathways by S-nitrosylation. S-Nitrosylation of cell wall modifying and proteolytic enzymes in the apoplast suggested differential and spatial regulation by S-nitrosylation. To have an overview of physiological role(s) of NO, collective information on NO based signaling (mainly by S-nitrosylation) is presented in this review.

© 2014 Elsevier Inc. All rights reserved.

#### Contents

1.	Introd	luction	. 98
2.	Stress	induced nitric oxide production and thiol pool modulation	. 98
3.	S-Nitr	osoglutathione (GSNO) responsive S-nitrosylation	100
4.	Endog	genous and stress responsive S-nitrosylation	101
5.	Nitric	oxide regulates defense, stress, redox, metabolic and photosynthetic pathways	105
	5.1.	Nitrosylation mediated signaling of Brassicaceae specific glucosinolate hydrolysis pathway	105
	5.2.	Nitric oxide is involved in cellular detoxification by regulating enzymes of ascorbate-glutathione cycle,	
		by enhancing superoxide dismutase and glutathione S-transferase activity and by methylglyoxal detoxification	105
		5.2.1. Heavy metal stress	
		5.2.2. Salinity and cold stress	105
	5.3.	Role of nitric oxide in regulating protein synthesis, folding and proteolysis	106
	5.4.	Role of nitric oxide in regulating cell wall modifying enzymes	106
	5.5.	Nitric oxide regulates Glycolysis, ethylene and methionine biosynthesis pathways by S-nitrosylation	106

E-mail naddress: rdeswal@botany.du.ac.in (R. Deswal).

Abbreviations: Nitric oxide, NO; S-Nitrosothiols, SNOs; S-Nitrosoglutathione, GSNO.

<sup>\*</sup> Corresponding author. Fax: +91 011 27662273.

	5.6. S-nitrosylation of photosynthetic proteins inhibits their activity	106
	5.7. Nitric oxide promotes seed germination, causes dormancy release and regulates seed storage proteins	107
6.	Nitric oxide (NO) based post-translational modifications cross-talk	107
7.	Conclusions and future prospects	107
	Acknowledgments	110
	Appendix: Supplementary material	110
	References	

#### 1. Introduction

Brassica juncea (Indian mustard) is an oil yielding crop of Brassicaceae family in the order Capparales. B. juncea faces yield loss due to abiotic stress (cold, salinity and heavy metal [1-3]). Although, India is the second largest cultivator of Brassica sp., but due to crop loss by these stress, it is not able to meet its annual edible oil requirement and 40% of its requirements are met by import [1]. Moreover, to further aggravate the situation, over 30% yield loss by abiotic stress is predicted in India alone. Therefore, it is important to understand its stress signaling mechanism, which would help in generating stress tolerant crops. In the last two decades, nitric oxide (NO) has emerged as an important signaling molecule, which regulates plant growth and development [4–6]. Its role in regulating large number of physiological processes and providing stress tolerance has been extensively reviewed [7-14]. Most of the work on NO signaling has been done on A. thaliana (a model member of Brassicaceae). However, the current status of NO signaling in crops including B. juncea has not been reviewed, therefore, this review presents evidences for NO signaling in B. juncea (Fig. 1). Additionally, effect of stress on the thiol pool is also described. As NO regulates diverse biological processes by regulating proteins through NO based posttranslational modifications (PTMs) [15,16], current status of these PTMs in *B. juncea* is also presented. An attempt has also been made to understand the effect of NO in regulating both Brassicaceae specific (glucosinolate-hydrolysis pathway) as well as other vital metabolic and regulatory pathways. Moreover, this review highlights the areas where further work is required to understand NO signaling in the Indian mustard particularly.

# 2. Stress induced nitric oxide production and thiol pool modulation

Eight major pathways, classified as oxidative and reductive pathways produce NO in plants [17–19]. Oxidative pathways include oxidation of L-arginine [by nitric oxide synthase (NOS)-like enzyme], polyamines and hydroxylamine, while reductive pathways include reduction of nitrite by enzymatic- [by nitrate reductase (NR), nitrite: NO reductase (Ni-NOR), xanthine oxidoreductase (XOR), cytochrome c oxidase and/or reductase] and non-enzymatic pathways (in an acidic environment). NOS-like enzyme catalyzes the oxidation of L-arginine to citrulline and NO is released as a by-product of the reaction. NOS-like enzyme activity showed 78.1% and 139.5% increase at 4 and 0 °C stress respectively in Chorispora bungeana suspension cultures [20]. Additionally, treatment with L-NNA (N (omega)-nitro-l-arginine, an inhibitor of NOS) inhibited (70%) NO production in cold (30 days) treated Solanum lycopersicum fruits, therefore, indicating NOS-like enzyme dependent NO production in cold [21]. NR is the major NO producing enzymes and produce NO by reducing nitrite in the presence of NAD(P)H. Cold

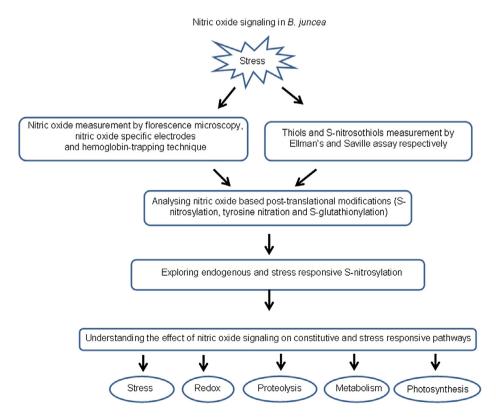


Fig. 1. Flow diagram showing steps of nitric oxide signaling analysis in B. juncea.

### Download English Version:

# https://daneshyari.com/en/article/2000512

Download Persian Version:

https://daneshyari.com/article/2000512

Daneshyari.com