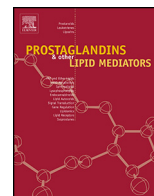




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Review

Vascular actions of 20-HETE

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ABSTRACT

20-hydroxyeicosatetraenoic acid (20-HETE) is a metabolite of arachidonic acid that exhibits a myriad of biological effects in the vascular system. This review discusses the current knowledge related to the effects of 20-HETE on vascular reactivity, activation, and remodeling, as well as its role in vascular inflammation and angiogenesis. The information explaining how 20-HETE and the renin-angiotensin system interact to promote hypertension, vasoconstriction, and vascular dysfunction is summarized in this article. 20-HETE enhances vascular inflammation and injury in models of diabetes, ischemia/reperfusion, and cerebrovascular oxidative stress. Recent studies also established a role for 20-HETE in normal and pathological angiogenesis conditions. This review will also discuss the molecular mechanisms through which 20-HETE induces these vascular actions. Potential additional studies are suggested to address shortcomings in the current knowledge of 20-HETE in the vascular system.

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1. Introduction

20-hydroxyeicosatetraenoic acid (20-HETE) is an eicosanoid that regulates a myriad of actions in the vascular system. It is synthesized through metabolism of arachidonic acid (AA) by cytochrome P450 (CYP) ω -hydroxylases. Several isoforms of CYP ω -hydroxylases, which are the main producers of 20-HETE, are

expressed in humans, mice, and rats. A number of research groups have established a role for 20-HETE in the vascular system through the use of cell, animal, and human models.

The main focus of this review is to discuss the effects of 20-HETE in the vascular system and the mechanisms involved in these processes. 20-HETE has an integral interaction with the renin-angiotensin system leading to a feed-forward mechanism that perpetuates vascular dysfunction and hypertension. The mechanisms involving 20-HETE in vascular reactivity, activation, and remodeling have been extensively studied. Changes in 20-HETE production also regulate vascular inflammation in diabetes, ischemia/reperfusion and cerebrovascular oxidative stress injury. Several studies established that 20-HETE enhances angiogenesis in

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Table 1
Summary of CYP450 ω -hydroxylases that produce 20-HETE in humans, rats, and mice.

Species	Cytochrome P450 ω -hydroxylases responsible for 20-HETE production
Human	CYP4A11 (CYP4A11); CYP4A22 (CYP4A22); CYP4F2 (CYP4F2); CYP4F3 (CYP4F3)
Rat	Cyp4a1 (CYP4A1); Cyp4a2 (CYP4A2); Cyp4a3 (CYP4A3); Cyp4a8 (CYP4A8)
Mouse	Cyp4a10 (CYP4A10); Cyp4a12 (CYP4A12)

gene; (protein).

normal and pathological conditions. Here we summarize the literature related to these vascular actions of 20-HETE and what is known regarding the mechanisms through which 20-HETE regulates these processes. We also address currently unanswered questions that are of interest to further advance the understanding of the vascular actions of 20-HETE.

2. 20-HETE biosynthesis

20-HETE is derived from metabolism of AA by CYP ω -hydroxylases of the CYP4A and CYP4F subfamilies. Arachidonic acid is a polyunsaturated fatty acid that is a major component of membrane phospholipids. AA is liberated from the plasma membrane by phospholipase A2. To produce 20-HETE, the CYP ω -hydroxylases insert a hydroxyl group at the terminal sp^3 carbon group of AA [1]. There are several isoforms of CYP4A/CYP4F responsible for the production of 20-HETE, which are summarized in Table 1. In humans, these isoforms are CYP4A11, CYP4A22, CYP4F2, and CYP4F3 [2–4]. The predominant 20-HETE synthase in humans is CYP4F2 followed by CYP4A11. CYP4F2 exhibits high activity in leukocytes and kidneys [4,5]. In mice, the 20-HETE producing enzymes include CYP4A10 and CYP4A12 [6]; CYP4A12 is the primary 20-HETE synthase [6,7]. In rats the 20-HETE producing enzymes include CYP4A1, CYP4A2, CYP4A3, and CYP4A8 [8–10].

Various studies have revealed functional variants in both human CYP4F2 and CYP4A11. Population differences have been observed in the CYP4A11 loss-of-function variant 8590T>C with higher frequency being observed in African-American and some Japanese populations [11]. *In vitro* experiments have demonstrated that several human CYP4F2 variants result in reduced production of 20-HETE [12]. In contrast to these *in vitro* results, an *in vivo* study revealed that the CYP4F2 V433M polymorphism was associated with increased urinary excretion of 20-HETE [13]. These discrepancies could be due to different factors regulating 20-HETE production in humans as compared to isolated *in vitro* systems. Caution should be taken when comparing *in vitro* results to human populations.

Vascular synthesis and release of 20-HETE occurs primarily in vascular smooth muscle cells [14–20]. These cells are not the sole source of 20-HETE; it can arise from myeloid cells in the peripheral blood and bone marrow [21–23]. 20-HETE is also produced in human neutrophils and platelets [24]. Neutrophil and platelet 20-HETE production is increased by Ang II and endothelin-1 treatment [24]. Androgen is also a potent inducer of 20-HETE synthesis [25]. Interestingly, endothelial progenitor cells, which are involved in postnatal neovascularization, produce 20-HETE [26]. In contrast, vascular endothelial cells in most circulatory beds are devoid of 20-HETE synthase activity [27].

3. 20-HETE and the renin-angiotensin system (RAS)

The renin-angiotensin system (RAS) serves a critical role in the regulation of blood pressure. The RAS is comprised of several components including renin, angiotensin-converting enzyme (ACE), and angiotensin II type 1 receptor (AT1R). Formation of the

vasoactive octapeptide angiotensin II (Ang II) occurs through step-wise degradation of angiotensinogen. Angiotensinogen, which is primarily produced by the liver, is first converted to the decapeptide angiotensin I (Ang I) via the enzyme renin. Ang I is further cleaved by ACE to its vasoactive Ang II form. The vasomotor actions of Ang II are primarily via activation of the AT1R within the vasculature resulting in vasoconstriction and a variety of other vascular, renal, and fluid balance effects [28,29].

Several studies document the complex interactions between the RAS and 20-HETE in hypertension. The release and synthesis of 20-HETE is induced by several autacoids including endothelin-1 [30–32] and Ang II [33,34]. Ang II stimulates the synthesis and release of 20-HETE from isolated rat preglomerular microvessels to enhance the pressor effects of Ang II [35–37]. 20-HETE mediates the mitogenic [15,33,38–40] and vasoconstrictor effects of Ang II by mediating hypertrophy and hypertension through activation of the Ras/MAP kinase pathway [41]. Thus, inhibition of 20-HETE synthesis attenuates the renal pressure response to Ang II as well as inhibits the development of Ang II-dependent hypertension [42,43]. Interestingly, Ang II's actions on vascular cells parallel the biological actions of 20-HETE: stimulation of superoxide/ROS, NF- κ B activation, and induction of inflammatory adhesion molecules (ICAM/VCAM) [44–52]. Conversely, recent studies identified 20-HETE as a potent inducer and transcriptional activator of endothelial ACE expression in microvascular endothelial cells [53,54].

Animal models of hypertension that demonstrate increased vascular 20-HETE production are also RAS-mediated and-dependent. These models include the spontaneous hypertensive rat (SHR) [55,56] and androgen-induced hypertension [17,57,58]. Androgen influences renal 20-HETE synthesis in spontaneously hypertensive rats [59]. Sprague-Dawley rats overexpressing CYP4A2 in the vascular endothelium exhibit increased 20-HETE production and hypertension [6,17,60]. The increase in blood pressure coincides with increased expression of vascular ACE and is normalized by ACE inhibition or AT1R blockade [53,60]. These observations suggest a feed forward mechanism by which the 20-HETE axis and the RAS work in concert to promote vascular dysfunction and hypertension [61]. The interaction between the RAS and 20-HETE is depicted in Fig. 1.

4. Vascular reactivity, endothelial dysfunction and endothelial activation in response to 20-HETE

20-HETE sensitizes vascular smooth muscle cells to a variety of constrictor stimuli, including Ang II, phenylephrine and endothelin [18,62,63] through several mechanisms. 20-HETE sensitizes smooth muscle through inhibition of the large conductance Ca^{2+} -activated K^+ (BKCa) channels. Inhibition of BKCa channels depolarizes plasma membranes, increases Ca^{2+} entry through L-type Ca^{2+} channels, and elevates cytosolic $[Ca^{2+}]$ to potentiate vasoconstriction [14,15,64]. 20-HETE can also increase conductance of the L-type Ca^{2+} channels through PKC activation. In small coronary arteries, 20-HETE activates Rho kinase resulting in phosphorylation of myosin light chain to increase sensitivity of the vessel to Ca^{2+} [41]. 20-HETE not only induces vasoconstriction but also reduces vascular relaxation. For example, 20-HETE attenuates the relaxing response to acetylcholine in renal interlobar arteries pre-constricted with phenylephrine [65–67]. These vascular effects depend on the complex relationship between RAS and 20-HETE as concurrent pretreatment of vessels with 20-HETE and an ACE inhibitor (Lisinopril) or AT1R blocker (Losartan) attenuate 20-HETE's effects [54].

Nitric oxide (NO) produced by the vascular endothelium is an important mediator in the defense against vascular injury and

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