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^{13}C-based metabolic flux analysis of *Saccharomyces cerevisiae* with a reduced Crabtree effect

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***Saccharomyces cerevisiae* shows a Crabtree effect that produces ethanol in a high glucose concentration even under fully aerobic condition. For efficient production of cake yeast or compressed yeast for baking, ethanol by-production is not desired since glucose limited chemostat or fed-batch cultivations are performed to suppress the Crabtree effect. In this study, the ^{13}C-based metabolic flux analysis (^{13}C-MFA) was performed for the S288C derived *S. cerevisiae* strain to characterize a metabolic state under the reduced Crabtree effect. *S. cerevisiae* cells were cultured at a low dilution rate (0.1 h^{-1}) under the glucose-limited chemostat condition. The estimated metabolic flux distribution showed that the acetyl-CoA in mitochondria was mainly produced from pyruvate by pyruvate dehydrogenase (PDH) reaction and that the level of the metabolic flux through the pentose phosphate pathway was much higher than that of the Embden–Meyerhof–Parnas pathway, which contributes to high biomass yield at low dilution rate by supplying NADPH required for cell growth.**

[**Key words:** ^{13}C-based metabolic flux analysis; Crabtree effect; Central carbon metabolism; *Saccharomyces cerevisiae*; Redox balance]

Saccharomyces cerevisiae is an industrial microorganism with stable ethanol fermentation performance even under aerobic and high glucose concentration conditions (1). The Crabtree effect is, however, not preferable to efficiently produce cake yeast or compressed yeast for baking because glucose-limited chemostat or fed-batch cultivation is required to suppress ethanol by-production. While baker's yeasts are industrially produced at a specific growth rate of 0.25 h^{-1} for faster cell growth, it has been reported that no ethanol production by the Crabtree effect occurs in a glucose-limited culture with dilution rates of approximately 0.1–0.15 h^{-1} (2). Previous studies suggested that the limited respiratory capacity of *S. cerevisiae* mitochondria is countered by the Crabtree effect to maintain the redox balance in the cytosol (3,4).

To further characterize the metabolic state related to the Crabtree effect, ^{13}C-based metabolic flux analysis (^{13}C-MFA) was conducted using *S. cerevisiae* ATCC 32167 strain (2). The glucose limited chemostat cultivations showed that a Crabtree effect was observed at dilution rates of 0.30 h^{-1} and 0.40 h^{-1} that had ethanol yields of 0.16 and 0.36 C-mol C-mol-glucose^{-1}, respectively. The metabolic flux analysis at a dilution rate of 0.40 h^{-1} revealed that the flux level of the Embden–Meyerhof–Parnas (EMP) pathway (77.9 when the glucose uptake rate was set to 100) was larger than that of the pentose phosphate (PP) pathway (15.1 relative to the glucose uptake rate). In the case of a dilution rate of 0.15 h^{-1}, no ethanol production by the Crabtree effect was observed, and the biomass yield from the glucose was increased to 0.57 C-mol C-mol-glucose^{-1}, which is twice as high as the yield with a dilution rate of 0.40 h^{-1} (0.29 C-mol C-mol-glucose^{-1}) (2). The metabolic flux level of the EMP pathway was reduced to 25.2 and that of PP pathway was increased to 54.7. This result suggests that the activation of the PP pathway should relate to the reduced Crabtree effect in the ATCC 32167 strain.

The ^{13}C-MFA study also suggested a metabolic route for mitochondrial acetyl-CoA supply in the presence of a reduced Crabtree effect. The metabolic flux analysis revealed that mitochondrial acetyl-CoA was dominantly supplied from mitochondrial pyruvate by pyruvate dehydrogenase (PDH) in the fermentative growth condition (μ = 0.40 h^{-1}). The metabolic flux of the PDH reaction was determined to be 23.7 relative to the glucose uptake rate when the glucose uptake rate was set to 100. In the oxidative condition (μ = 0.15 h^{-1}), intercompartmental acetyl-CoA transport was the major route for mitochondrial acetyl-CoA supply (53.4). In this pathway, pyruvate was first decarboxylated to acetaldehyde by pyruvate decarboxylase (PDC), which was converted to acetate and acetyl-CoA in the cytosol, and then transported into the

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mitochondria (6,7). Similar results were reported in a ^{13}C-MFA study of the CEN.PK113-7D strain cultivated in a glucose-limited chemostat culture at a dilution rate of 0.10 h^{-1} (8). However, the enzyme kinetic data suggested that the PDH pathway rather than the PDC pathway should be mainly responsible for the mitochondrial acetyl-CoA supply under oxidative conditions (5–7).

In this study, to investigate the role of the PP pathway and the major route for the mitochondrial acetyl-CoA supply in the presence of a reduced Crabtree effect, ^{13}C-MFA was performed for another experimental strain of *S. cerevisiae* S288C. The results revealed that acetyl-CoA was mainly produced via the PDH pathway in mitochondria in the presence of a reduced Crabtree effect in *S. cerevisiae* S288C. The results also showed that elevated PP pathway flux should be a common adaptation mechanism against oxidative conditions among *S. cerevisiae* strains.

MATERIALS AND METHODS

Strain, media, and culture conditions *S. cerevisiae* YM1 (*MATα ura3Δ0*) was constructed from the BY4739 (*MATα leu2Δ0 lys2Δ0 ura3Δ0*) strain by restoring leucine and lysine autotrophy. The BY4739 strain was derived from S288C (9). The complete open reading frames of the *LEU2* and *LYS2* promoters together with the 335 bp and 150 bp neighboring regions were amplified using polymerase chain reaction (PCR) applied to the genomic DNA of the *S. cerevisiae* S288C strain. The PCR was performed using the following primers: for *LEU2* forward: 5′-TCATATGGATTCCTAATCCTCGAGGAGAATTTATAATATAGTCACATAACGAGAACACACAG GGGCG-3′, *LEU2* reverse: 5′-TTACCTGTATTCCTTTACATCCTCC-3′, *LYS2* forward: 5′-AGTTGCTTTCTCCTATGGGAAGAGC-3′ and *LYS2* reverse: 5′-TACAATAAACCAAGA TGAAGCTGCC-3′, respectively. PCR was performed using the Z-taq polymerase (Takara Bio Inc., Shiga, Japan). Each amplified fragment was cloned into pGEM-T easy (Promega Co., Madison, WI, U.S.A.) and was then digested by *Not*I. The fragments were transformed into the *S. cerevisiae* BY4739 strain using the lithium acetate method (10) to obtain leucine and lysine autotroph strains.

The preculture and main culture were performed in synthetic dextrose (SD) medium [5 g L^{-1} glucose, 6.7 g L^{-1} yeast nitrogen base without amino acids (Difco Laboratories, Detroit, MI, USA)] containing 0.076 g L^{-1} uracil. Precultivation was performed at 30°C for 20 h in 5 mL of SD medium. The main culture was performed in a 1 L BNJ-P type bioreactor (ABLE, Japan) jar fermenter with a working volume of 500 mL at 30°C. The pH was maintained at 6.0 using 1*N* NaOH. The aeration rate and the agitation speed were set to 1.0 L min^{-1} and 350 rpm, respectively. For the continuous culture, the dilution rate was set to 0.1 h^{-1}. After the continuous culture reached a steady state, the feeding medium that contained the natural glucose, was replaced with 4.93 g L^{-1} glucose that contained labeled glucose composed of 41.0% [U–^{13}C] glucose, 30.7% [1–^{13}C] glucose and 28.3% natural glucose.

The cell growth and the glucose concentration were monitored using a spectrophotometer (OD_{660}, UVmini-1240; Shimadzu Corp., Kyoto, Japan) and a glucose sensor (BF-5; Oji Scientific Instruments, Japan), respectively. The cell dry mass was determined after centrifugation and drying at 60°C until a constant weight was achieved. The ethanol concentration was determined using a 7890A gas chromatography system (Agilent Technologies, Palo Alto, CA, USA) equipped with a capillary column (Restek 10,657 Stabilwax; 60 m × 0.32 mm ID × 1 μm; Shimadzu GLC, Japan) as previously described (11). The concentrations of glycerol, succinate, and acetate were determined using an HPLC system (HPLC Prominence, Shimadzu, Japan) equipped with an Aminex HPX-87H column (Bio-Rad, Hercules, CA, USA), a UV/VIS detector (SPD-20A), and a refractive index detector (RID-10A). The column was operated at 65°C using 1.5 mM H_2SO_4 as the mobile phase and with a flow rate set to 0.5 mL min^{-1}. The flow cell temperature of the refractive index detector was set to 35°C.

^{13}C-based metabolic flux analysis After feeding with the labeled glucose, the *S. cerevisiae* cells were sampled at 199, 201, and 203 h. The culture medium (10 ml) was centrifuged for 10 min at 10,000 rpm and 4°C. After washing with a 0.9% NaCl solution, the biomass was hydrolyzed using 6 *N* HCl at 105°C for 18 h. The hydrolyzed sample was filtered to remove cell debris and 10 μL of 600 μM cycloleucine was added as an internal standard. The sample was then dried under vacuum using an evaporator (Integrated Speedvac; Thermo Fisher scientific).The dried pellet was derivatized by adding 50 μL of acetonitrile and 50 μL of *N*-*tert*-butyldimethylsilyl-*N*-methyltrifluoroacetamide (MTBSTFA) followed by heating at 105°C for 1 h. The sample (1 μL) was analyzed by GC–MS (7890A GC and 5975C GC/MSD, Agilent Technologies) following a previously described method (12).

The metabolic network of *S. cerevisiae* was constructed using slight modifications of previously published models (2). The metabolic model comprises the major pathways of central carbon metabolism (glycolysis, PP pathway, anaplerosis, and tricarboxylic acid cycle) and the transport reaction between cytosol and mitochondria. Pyruvate, oxaloacetate, and acetyl-CoA (AcCOA) in the cytosol and mitochondria are considered to be separate metabolite pools. Alanine is synthesized from both cytosolic and mitochondrial pathways by alanine amino transferase (13). For glycine synthesis, threonine aldolase (14) accounted for by synthesis from serine. The rates of biomass synthesis were calculated from published data (15).

The computational procedure was performed by OpenMebius (16) using MATLAB (MathWorks, Natick, MA, USA). A metabolic flux distribution was estimated by a nonlinear fitting of the metabolic model using the isotopic labeling patterns of proteinogenic amino acids determined by GC–MS. The nonlinear optimization was performed by using "fmincon" function in MATLAB optimization toolbox (MathWorks, Natick, MA, USA).

The 90% confidence intervals were determined using the grid search method at the range of fixed metabolic flux whose sum of squared residuals was less than the threshold level (17). The threshold level was determined by

$$\Phi_{res,sr} \leq \Phi_{res} + \frac{\Phi_{res}}{n-p} F_{\alpha}(1, n-p) \quad (1)$$

where $\Phi_{res,\,sr}$ is the minimized squared sum of residuals with one fixed flux, Φ_{res} is the original minimized squared sum of residuals, n is the number of independent data points used in the fitting, p is the degrees of freedom in the original flux fit, F is the F-distribution, and α is the confidence level (18).

Mass balance analysis ATP production through the EMP pathway and TCA cycle, together with ATP consumption by glucose uptake and phosphofructokinase, was calculated using the estimated metabolic flux distribution. The excess NADH and NADPH were assumed to be converted to ATP via oxidative phosphorylation with a P/O ratio of 1.0 as previously described (19). Several ATP consumption rates (per gram of dry cell weight, $gDCW^{-1}$) were calculated using the genome-scale model iFF708 considering growth-associated ATP maintenance, synthesis of biomass building blocks, polymerization cost, and nongrowth-associated ATP requirements (15). The amount of NADPH required for biomass formation has been estimated to be 9.31 mmol $gDCW^{-1}$ (20).

RESULTS

^{13}C-based metabolic flux analysis In this study, ^{13}C-MFA was performed using the YM1 strain (*MATα ura3Δ0*) constructed from the BY4739 strain that is congenic to S288C. Although the S288C strain shows slightly different metabolic phenotypes from other commonly used strains such as CEN.PK (21), the strain was employed in this study for comparison to the ATCC 32167 strain (2). The main reason for this comparison is that this is the most characterized *S. cerevisiae* strain.

A glucose-limited chemostat culture was conducted at dilution rate 0.1 h^{-1} to repress the Crabtree effect. The cultivation profile (Fig. 1) showed that the culture reached a metabolic steady state at 50 h after the initiation of cultivation. The Crabtree effect was successfully suppressed because *S. cerevisiae* cells only produced a small amount of acetate (0.023 mmol $gDCW^{-1}$ h^{-1}) rather than ethanol and glycerol. Based on the optical density (OD_{660} = 11.0) and the cell dry mass (0.21 gDCW OD_{660}^{-1} h^{-1}), the mean glucose

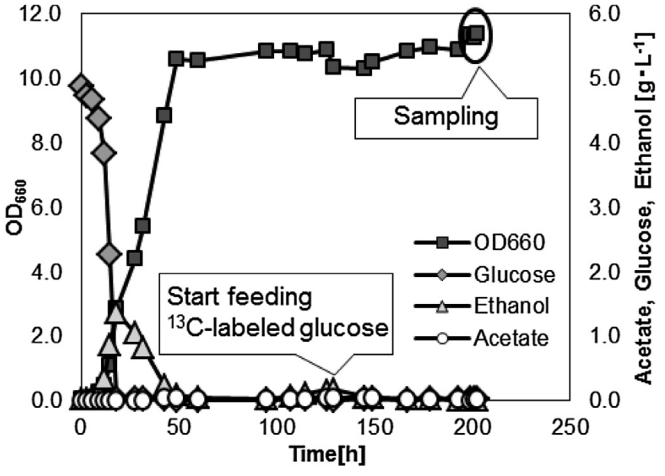

FIG. 1. Cultivation profiles. A glucose-limited chemostat culture was conducted at a dilution rate of 0.1 h^{-1}. The glucose-limited chemostat was performed 18 h after the culture was started. The feeding medium that contained natural glucose was replaced with labeled glucose at 129 h.