

Available online at www.sciencedirect.com**MYCOSCIENCE**

ISSN 1340-3540 (print), 1618-2545 (online)

journal homepage: www.elsevier.com/locate/myc**Full paper**

***Pseudoidium javanicum*, a new species of powdery mildew on *Acalypha* spp. from Indonesia**

Jamjan Meeboon^a, Iman Hidayat^b, Susumu Takamatsu^{a,*}

^a Department of Bioresources, Graduate School, Mie University, 1577 Kurima-Machiya, Tsu 514-8507, Japan

^b Microbiology Division, Research Center for Biology, Indonesian Institute of Sciences-LIPI Jl, Raya Jakarta-Bogor KM 46, Cibinong, West Java 16911, Indonesia

ARTICLE INFO**Article history:**

Received 18 July 2012

Accepted 24 August 2012

Available online 31 December 2012

Keywords:

Erysiphaceae

Erysiphales

Euphorbiaceae

rDNA sequence

Taxonomy

ABSTRACT

Pseudoidium javanicum is proposed as a new species based on analyses of 28S, ITS and IGS rDNA sequences, and morphological data. This new species was found on *Acalypha wilkesiana* var. *marginata*, *A. argentea*, and *A. cristata* collected from Cibodas Botanical Garden, Bogor (West Java Province, Indonesia). Our analyses showed that all these specimens have identical rDNA sequences and similar morphological characteristics. They form a distinct clade separated from other species of Erysiphaceae. *Pseudoidium javanicum* differs from *Erysiphe acalypae* by having shorter conidiophores and foot cells 1–3 times as long as the 0–2 following cells. The conidial size of *Ps. javanicum* is also smaller than that of *E. jatrophae*.

© 2012 The Mycological Society of Japan. Published by Elsevier B.V. All rights reserved.

1. Introduction

Acalypha L. is one of the largest genera in the plant family Euphorbiaceae comprising about 462 species (Qin et al. 2006). The genus consists of herbs, shrubs, and trees and it is mainly distributed in tropical and subtropical regions, and a few species are found in temperate areas (Atha 2008). Five powdery mildew species have been recorded on hosts belonging to the genus *Acalypha*, namely, *Erysiphe acalypae* (F.L. Tai) R.Y. Zheng & G.Q. Chen, *E. jatrophae* Doidge, *Podosphaera euphorbiae-hirtae* (U. Braun & Somani) U. Braun & S. Takam., *Golovinomyces sparsus* (U. Braun) V.P. Heluta, and *Fibroidium acalypae* (Chidd.) U. Braun & R.T.A. Cook (Amano 1986; Braun 1987; Braun and Cook 2012).

During visits in the Cibodas Botanical Garden (West Java, Indonesia) in March 2011 and 2012, three species of *Acalypha* – *A. wilkesiana* var. *marginata* E. Morren, *A. argentea* Hort., and *A. cristata* Radcl.-Sm. – were found to be infected by powdery mildews. Morphological examination confirmed that the causal agents belong to the genus *Pseudoidium* [anamorph of *Erysiphe* R. Hedw. ex DC. emend. U. Braun & S. Takam.], but all specimens are distinct from *E. acalypae* (Tai 1946) by having shorter conidiophores and foot cells 1–3 times as long as the 0–2 following cells. All nucleotide sequences from internal transcribed spacer (ITS), intergenic spacer (IGS) and 28S regions of the ribosomal DNA showed that all specimens on *Acalypha* spp. collected in Indonesia form an independent lineage separated from other *Erysiphe*

* Corresponding author. Tel.: +81 59 231 9497; fax: +81 59 231 9637.

E-mail address: takamatu@bio.mie-u.ac.jp (S. Takamatsu).

1340-3540/\$ – see front matter © 2012 The Mycological Society of Japan. Published by Elsevier B.V. All rights reserved.

<http://dx.doi.org/10.1016/j.myc.2012.08.006>

species. Therefore, the fungus concerned has to be considered a new species.

2. Materials and methods

2.1. Morphological examination

Specimens were collected at Cibodas Botanical Garden, Bogor (West Java Province, Indonesia) in March 2011 and 2012. Details of host names, collection dates, localities, and collectors were recorded. Herbarium samples were rehydrated before examination by boiling a small piece of infected leaf with the fungal mycelium downwards in a drop of lactic acid on a slide as described by Shin and La (1993). After boiling, the rehydrated mycelium was scraped off and mounted in lactic acid using a light microscope with phase contrast. Thirty conidia, conidiophores, foot cells and mother cells were measured per sample. Specimens were deposited at the National Museum of Nature and Science (TNS) and Mie University Mycological Herbarium (MUMH) [Japan] and Herbarium Bogoriense (BO) [Indonesia].

2.2. Phylogenetic analysis

DNA extraction of the powdery mildew specimens was conducted according to the chelex method (Walsh et al. 1991) as described in Hirata and Takamatsu (1996). The 5'-end of the 28S rDNA (including the domains D1 and D2), ITS region including the 5.8S rDNA, and IGS region were amplified by polymerase chain reaction (PCR) using the respective primer pairs: PM3/TW14 (28S), ITS5/PM6 (ITS fragment 1) and PM7/ITS4 (ITS fragment 2), and IGS-12A/NS1R (IGS; Carbone and Kohn 1999). KOD FX Neo DNA polymerase (Toyobo, Japan) was used in the PCR reaction according to the manufacturer's protocol. The amplicons of 28S rDNA, ITS and IGS were sent to SolGent Co. Ltd. (Daejeon, South Korea) for sequencing using primers NL1 and NLP2 (28S), ITS1 and ITS4 (ITS), and IGS-12A and NS1R (IGS). Representative sequences determined in this study were deposited in the DNA DataBase of Japan (DDBJ) under the accession numbers of AB733586 – AB733597 (Table 1). Sequences generated from the 28S rDNA region were aligned with other sequences of Erysiphaceae retrieved from DNA databases (DDBJ, EMBL, NCBI) using MEGA 5 (Tamura et al. 2011). The alignment was deposited in TreeBASE

(<http://www.treebase.org/>) under the accession number of S12969. Phylogenetic trees were calculated from the dataset using neighbor joining (NJ) (Saitou and Nei 1987), maximum parsimony (MP), and maximum likelihood (ML) methods in PAUP* 4.0b10 (Swofford 2002). For ML and NJ analyses, the most appropriate evolutionary model was determined for a given dataset using PAUP* and Modeltest 3.06 (Posada and Crandall 1998). A starting tree was obtained with the NJ method. With this tree, likelihood scores were calculated with PAUP* for 56 alternative models of evolution. The output file was then imported into Modeltest to compare the models using Akaike's (1974) information criterion (AIC). Once a model of evolution was chosen, it was used to construct phylogenetic trees with the NJ and ML method using PAUP*. The GTR+G+I model (Tavaré 1986) was selected as the best evolution model to construct trees. For MP analyses, the heuristic search option using the 'tree-bisection-reconstruction' (TBR) algorithm, the MULPARS option was in effect, and zero-length branches were collapsed. All sites were treated as unordered and unweighted, with gaps treated as missing data. The strength of the internal branches of the resulting tree was tested with bootstrap (BS) analysis using 1000 replications (Felsenstein 1985) for all analyses.

3. Results

3.1. Taxonomy

Pseudoidium javanicum Meeboon & S. Takam. sp. nov. Fig. 1.

Mycobank no.: MB 800883.

Differs from *E. acalyphae* by having shorter conidiophores [(46–)48–69(–83) × (5–)5.5–7(–8) μm] and foot cells 1–3 times as long as the 0–2 following cells. The conidial size [(19–)20–25(–27) × (8–)9–12(–13) μm] is smaller than in *E. jatrophae*.

Type: on *A. wilkesiana* var. *marginata* E. Morren (Euphorbiaceae), Indonesia, West Java province, Cibodas Botanical Garden, Bogor, 7 March 2012 (Holotypus, TNS-F-46915; Isotypus, MUMH 5559).

Ribosomal DNA sequence ex holotype: AB733593 (ITS), AB733597 (28S), AB733586 (IGS).

Etymology: the new species is named after the place (Java Island) where the fungus was collected.

Mycelium amphigenous, mostly epiphyllous, almost persistent, effuse; hyphal appressoria solitary, rarely in

Table 1 – Sources of *Pseudoidium javanicum* material used for molecular analyses and DNA database accession numbers.

| Host | Specimen no. | Location and year | Accession no. | | |
|--|--------------|-------------------------------------|---------------|----------|----------|
| | | | ITS | 28S | IGS |
| <i>Acalypha wilkesiana</i> var. <i>marginata</i> | MUMH5559 | Indonesia: West Java province; 2012 | AB733593 | AB733597 | AB733586 |
| <i>A. cristata</i> | MUMH5560 | Indonesia: West Java province; 2012 | – | – | AB733587 |
| <i>A. argentea</i> | MUMH5561 | Indonesia: West Java province; 2012 | – | – | AB733588 |
| <i>A. wilkesiana</i> var. <i>marginata</i> | MUMH5149 | Indonesia: West Java province; 2011 | AB733594 | AB733589 | – |
| <i>A. wilkesiana</i> var. <i>marginata</i> | MUMH5150 | Indonesia: West Java province; 2011 | AB733595 | AB733590 | – |
| <i>A. cristata</i> | MUMH5151 | Indonesia: West Java province; 2011 | AB733591 | – | – |
| <i>A. argentea</i> | MUMH5152 | Indonesia: West Java province; 2011 | AB733596 | AB733592 | – |

Download English Version:

<https://daneshyari.com/en/article/2060642>

Download Persian Version:

<https://daneshyari.com/article/2060642>

[Daneshyari.com](https://daneshyari.com)