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***Erysiphe paracarpinicola*: A new species of *Erysiphe* sect. *Uncinula* on *Carpinus cordata* (Betulaceae)**

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ABSTRACT

A phylogeny of *Erysiphe* sect. *Uncinula* on *Carpinus* spp. was reconstructed using the 28S rDNA sequences and a combined alignment of the 28S, ITS, and IGS rDNA sequences. The analysis was supplemented with morphological data obtained from examination of voucher specimens. A sequence of *Erysiphe* sect. *Uncinula* on *C. cordata* formed a distinct lineage separated from sequences of other *Erysiphe* species on *Carpinus* spp., indicating a cryptic species, which is described as *E. paracarpinicola*. The new species is genetically as well as morphologically most similar to *E. carpinicola* s. str., but differs in having fewer asci per chasmothecium (mainly 3–5 vs 4–10) and shorter chasmothecial appendages. A key to species of *Erysiphe* sect. *Uncinula* on *Carpinus* spp. is provided.

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1. Introduction

Carpinus L. (Betulaceae) is the largest genus in the subfamily Coryloideae with about 35 species distributed in Eastern Asia, Europe and North America (Yoo and Wen 2007). Nine species of powdery mildews have been recorded parasitizing various species of *Carpinus* trees (Braun 1987; Braun et al. 2006; Braun and Cook 2012), namely, *Phyllactinia carpinii* (Rabenh.) Fuss, *Ph. carpinicola* U. Braun & S. Takam. *Erysiphe fimbriata* S. Takam. Masuya & Y. Nomura (sect. *Erysiphe*), *E. ellisii* (U. Braun) U. Braun & S. Takam. (sect. *Microsphaera*), and five species belonging to *Erysiphe* sect. *Uncinula*—*E. wuyiensis* (Z.X. Chen & R.X. Gao) U. Braun & S. Takam. *E. carpinii-cordatae* (Tanda & Y. Nomura) U. Braun, *E. arcuata* U. Braun, V.P. Heluta & S. Takam. *E. carpinicola* (Hara) U. Braun & S. Takam. *E. carpinii-laxiflorae* U. Braun, V.P. Heluta & S. Takam. Among them, species belonging to *E. sect. Uncinula* are mainly distributed in

East Asia (Japan, China, Korea, and Russian Far East), and only *E. arcuata* is distributed in Europe as well as in Asia (Braun and Cook 2012).

The morphological descriptions and phylogenetic affinities among *E. sect. Uncinula* parasitizing *Carpinus* spp. were previously described by Braun et al. (2006). The fungus on *C. cordata* Blume was described as *E. carpinii-cordatae*. However, a sequence from a specimen on *C. cordata* (MUMH207) differed from sequences of *E. carpinii-cordatae* (similarity = 66.5%). This sequence differed from all other *Uncinula* species on *Carpinus* spp. as well and belonged to a clade comprising *E. carpinicola* and *E. carpinii-laxiflorae* (Braun et al. 2006). However, the authors did not discuss the morphological characteristics of this fungus in detail, and maintained it as *Erysiphe* sp. The morphology of the herbarium specimen (MUMH207) collected in October 1996 has recently been re-examined, and nucleotide sequences of the intergenic spacer (IGS) region of rDNA

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for *Uncinula* species on *Carpinus* spp. have been determined to confirm the result of Braun et al. (2006). Our morphological examination and molecular phylogenetic analysis showed that the sequence is distinct from those of other closely related species of *E. sect. Uncinula* on *Carpinus* spp. Thus this fungus is described as a new species in this paper.

2. Materials and methods

2.1. Morphological examination

A voucher specimen of powdery mildew on *C. cordata* was obtained from Mie University Mycological Herbarium (MUMH 207), collected from Niigata Prefecture (Japan) in 1996, with the GenBank sequence code AB252464 derived from the same collection, which was also used by Braun et al. (2006). Additional specimens with the same location and collection date—MUMH 208, MUMH 297 and MUMH 183—were also examined. The method of morphological examination refers to Meeboon and Takamatsu (2012). Holo- and isotype material of the new species are deposited at the National Museum of Nature and Science (TNS), Japan and Mie University Mycological Herbarium (MUMH), Japan.

2.2. DNA sequencing and phylogenetic analysis

Whole-cell DNA was extracted from chasmothecia using the chelex method (Walsh et al. 1991) as described in Hirata and Takamatsu (1996). The IGS region was amplified by polymerase chain reaction (PCR) using the primer pairs IGS-12A/NS1R (Carbone and Kohn 1999). KOD FX Neo DNA polymerase (Toyobo, Japan) was used in the PCR reaction according to the manufacturer's protocol. The amplicons of IGS were sent to SolGent Co. Ltd. (Daejeon, South Korea) for sequencing using the primer pairs IGS-12A and NS1R. Representative sequences determined in this study were deposited in the DNA Data Bank of Japan (DDBJ) under the accession numbers of AB731687–AB731694 (Table 1).

Two kinds of phylogenetic analyses, viz. 28S rDNA, and combined analysis of 28S, internal transcribed spacer (ITS), and IGS rDNA sequences, were performed in this study. In the 28S rDNA analysis, 41 sequences of the 28S rDNA sequences including the sequences of *Erysiphe* sect. *Uncinula* on *Carpinus* spp. retrieved from GenBank were aligned using MUSCLE (Edgar 2004) implemented in MEGA 5 (Tamura et al. 2011). This

alignment was edited manually by eye. A sequence of *E. australiana* (McAlpine) U. Braun & S. Takam. was used as an out-group. Maximum likelihood (ML) and neighbor joining (NJ) analyses were performed by MEGA 5 and maximum parsimony (MP) analysis was performed by PAUP* 4.0b10 (Swofford 2002). In the ML and NJ analyses, the best-fit evolution model for the alignment was chosen from the 24 alternative models by the Bayesian information criterion using MEGA 5. The Kimura 2-parameter (Kimura 1980) + G + I model was selected as the best evolution model to construct trees of the 28S rDNA. Partial deletion was set as gap/missing data treatment with site coverage cutoff was set at 95%. Nearest-Neighbor-Interchange (NNI) was selected for ML heuristic method and initial tree for ML was set automatically. MP analysis was done with the heuristic search option using the 'tree-bisection-reconstruction' (TBR) algorithm. All sites were treated as unordered and unweighted, with gaps treated as missing data. The strength of the internal branches of the resulting tree was tested with bootstrap analysis using 1000 replications (Felsenstein 1985) in all ML, NJ and MP analyses.

In the combined analysis, the partition homogeneity test (Farris et al. 1995) was conducted using PAUP* 4.0b10 (Swofford 2002) to determine whether the 28S, ITS, and IGS data sets were in conflict, with 100 replicates. Only ML analysis was performed for this data set by the same conditions described above using MEGA 5. Tamura 3-parameter (Tamura 1992) + G + I model was selected as the best evolution model for this data set and the tree was rooted with mid-point rooting method. The alignments used in this study were deposited in TreeBASE (<http://www.treebase.org/>) under the accession number S12900.

Shimodaira–Hasegawa test (Shimodaira and Hasegawa 1999) of the 28S rDNA tree was conducted to test the hypothesis that *Erysiphe* sect. *Uncinula* on *Carpinus* spp. are monophyletic. The constraint tree was constructed in MacClade version 4 (Maddison and Maddison 2000) and executed in PAUP* 4.0b10 (Swofford 2002).

3. Results

3.1. Taxonomy

Erysiphe paracarpinicola Meeboon & S. Takam., sp. nov.
Figs. 3a–i, 4a–c.
MycoBank no.: MB800802

Table 1 – Sources of fungal material used for molecular analyses and DNA database accession numbers.

Host	Specimen no.	Location and year	Fungal species	Voucher no.	Accession no.	
					ITS+28S	IGS
<i>Carpinus cordata</i>	MUMH207	Niigata, Japan; 1996	<i>Erysiphe paracarpinicola</i>	KW30170	AB252464	AB731687
<i>C. betulus</i>	MUMH3197	Halle, Germany; 2004	<i>E. arcuata</i>	GLM53866, HAL1012F	AB252460	AB731688
<i>C. betulus</i>	MUMH3237	Saxony, Germany; 2004	<i>E. arcuata</i>	GLM53866, HAL1012F	AB252461	AB731689
<i>C. cordata</i>	MUMH3408	Sapporo, Japan; 2004	<i>E. carpini-cordatae</i>		AB252466	AB731690
<i>C. japonica</i>	MUMH243	Shiga, Japan; 1996	<i>E. carpinicola</i>	HAL1902F	AB252467	AB731691
<i>C. japonica</i>	MUMH3547	Gifu, Japan; 2004	<i>E. carpinicola</i>	KW30173	AB252468	AB731692
<i>C. laxiflora</i>	MUMH3503	Gifu, Japan; 2004	<i>E. carpini-laxiflorae</i>	KW30176	AB252470	AB731693
<i>C. laxiflora</i>	MUMH3640	Shiga, Japan; 2004	<i>E. carpini-axiflorae</i>	KW30179	AB252471	AB731694

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