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Lab Resource: Stem Cell Line

Generation of human control iPS cell line CHOPWT10 from healthy adult peripheral blood mononuclear cells



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ABSTRACT

The CHOPWT10 iPS cell line was generated to be used as a control for applications such as in differentiation analyses to the three germ layers and derivative tissues. Peripheral blood mononuclear cells (PBMCs) obtained from a healthy adult male were reprogrammed using the non-integrating Sendai virus expressing Oct3/4, Sox2, c-Myc, and Klf4.

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Resource table: CHOPWT10

Information in public databases

CHOPWT10 Name of stem cell construct

Institution The Children's Hospital of Philadelphia (CHOP)

Person who created resource Jean Ann Maguire

Contact person and email Deborah L. French, frenchd@email.chop.edu

Date archived/stock date 2014 Origin Human PBMCs

Type of resource Biological reagent: iPS cell from human PBMCs

Subtype Cell line

Oct4, Sox2, Klf4, c-Myc Key transcription factors

Identity and purity of cell line confirmed Authentication

N/A

(Figs. 1 and 2) Link to related literature N/A

Resource details

To generate CHOPWT10 cells, four Yamanaka reprogramming factors (Oct3/4, Sox2, c-Myc, and Klf4) were delivered into mononuclear cells through Sendai viral infection. The Sendai virus is non-integrating and the absence of the vectors in the reprogrammed cells was confirmed by PCR analysis (Fig. 1A).

DNA fingerprinting confirmed the genetic relation of the derivative line to the parental somatic line (Fig. 1B). The cells exhibited normal karyotype (46, XY) upon G-band analysis (Fig. 1C).

Pluripotency was verified by gene expression of pluripotent stem cell markers Oct4, Sox2, Nanog, Rex1, DMNT3B and ABCG2 by qPCR (Fig. 1D). In addition, pluripotent surface marker expression at single cell resolution of SSEA3, SSEA4, Tra-1-60, and Tra-1-81 were confirmed by flow cytometry (Fig. 1E). Differentiation capacity into three germ layers was confirmed by PluriTest™ and quantitative PCR (Fig. 2).

Materials and methods

Blood sample collection and processing

Blood samples were collected from a healthy adult male into BD Vacutainer cell preparation tubes. The PBMCs were isolated by centrifugation at 1800 rcf for 30 min, after which mononuclear cells were transferred to a fresh tube, washed with PBS, and pelleted at 300 rcf for 10 min. Pellets were resuspended in freezing media (90% FBS, 10% DMSO) and stored in liquid nitrogen. Blood collection was approved by the CHOP Institutional Review Board committee with written informed consent.

iPS cell reprogramming

The PBMCs were expanded and reprogrammed as previously described (Yang et al., 2012). Briefly, PBMCs were expanded in QBSF-60 medium containing EPO (2 U/mL), IGF-1 (40 ng/mL), SCF (50 ng/mL), IL-3 (10 ng/mL), dexamethasone (1.5 μM), ascorbic

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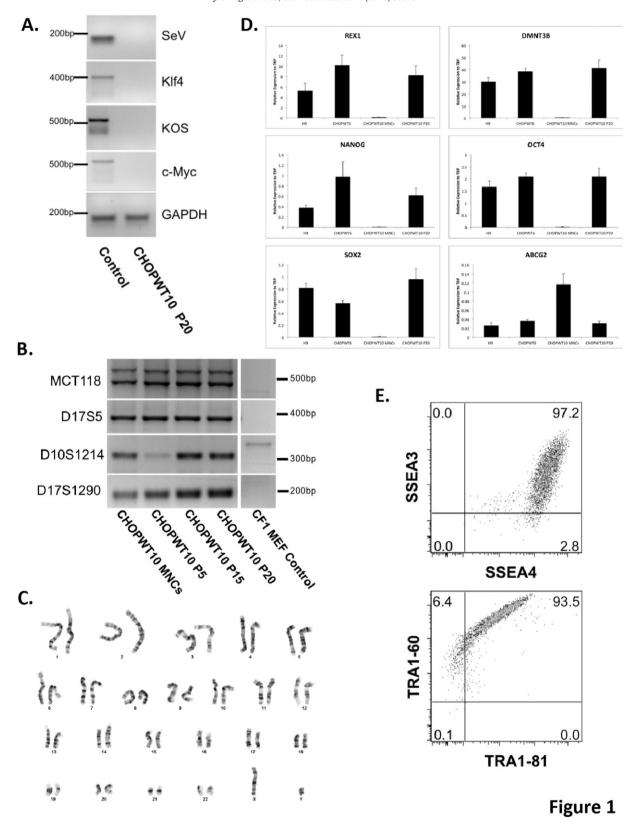


Fig. 1. Generation and characterization of CHOPWT10. (A) PCR analysis of reprogramming genes after 20 passages of CHOPWT10 using GAPDH as a loading control. Cells harvested immediately post-Sendai infection were used as a positive control. (B) CHOPWT10 iPS cells compared to parental cells by DNA fingerprinting. (C) Karyotype analysis of CHOPWT10. (D) Quantitative PCR of relative gene expression of pluripotency genes compared to embryonic stem cells (H9) and a control iPS cell line (CHOPWT6). (E) Flow cytometry profiles of surface markers on undifferentiated cells: SSEA3, SSEA4, TRA-1-60, and TRA-1-81.

acid (50 ng/mL), glutamine (1%), and penicillin/streptomycin (1%) for 7 days. Cells were transduced with Sendai virus expressing human Oct3/4, Sox2, Klf4, and c-Myc. (Life Technologies) (moi

according to manufacturer's instructions). Transduced cells were plated on culture dishes containing irradiated mouse embryonic fibroblasts (MEFs), and maintained in human embryonic stem

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