



Luteolysis in *Bos indicus* cows on Days 5 and 7 of estrous cycle with varying doses of PGF_{2α}

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ABSTRACT

The aim of this study was to evaluate luteolysis using three doses of PGF_{2α} on Day 5 or Day 7 of the estrous cycle in nonlactating Nellore (*Bos indicus*) cows. Cows (n = 323) were assigned within date of estrus (Day 0 of estrous cycle) to receive 12.5, 25.0, or 50.0 mg of PGF_{2α} on either Day 5 or Day 7 of the estrous cycle in a 3 × 2 factorial arrangement. Blood samples for progesterone (P₄) concentrations were collected at 0, 24, 48, and 72 hours after PGF_{2α} to assess luteolysis (L). Luteolysis was defined on the basis of P₄ concentrations at 72 hours using either less than 0.5 ng/mL (L0.5) or less than 1.0 ng/mL (L1.0) as the cut off. Luteolysis was considered “partial” when P₄ concentration declined within 24 hours after PGF_{2α} but failed to decline further or, in some cases, increased. Incidence of luteolysis was less (P < 0.01) on Day 5 than Day 7 of the estrous cycle (17.3 vs. 47.6% and 30.4 vs. 77.2%; for L0.5 and L1.0, respectively). Dose of PGF_{2α} increased (P < 0.01) L1.0 (12.5 mg = 38.9%; 25.0 mg = 52.3%; and 50.0 mg = 70.4%). Incidence of partial luteolysis for cows on Day 5 (57.1%) was greater (P < 0.01) than that on Day 7 (19.1%) of the estrous cycle and was more prevalent (P < 0.01) with lower doses of PGF_{2α} (12.5 mg = 49.1%; 25.0 mg = 37.4%; and 50.0 mg = 27.8%). In conclusion, both days of the estrous cycle and doses of PGF_{2α} influenced the incidence of complete and partial luteolysis in Nellore cows and should be an important consideration when devising estrus synchronization programs in this species.

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1. Introduction

Prostaglandin F_{2α} is a hormone that triggers luteolysis in cows, inducing the end of luteal phase [1]. However, despite the presence of similar concentrations of high-affinity PGF_{2α}

receptors in the early CL [2,3], luteolysis from exogenous PGF_{2α} does not usually occur until 5 to 6 days after estrus [4–9]. An important aspect of synchronization programs that use GnRH to synchronize emergence of a new follicular wave through induction of ovulation of the dominant follicle is to ensure lysis of the CL 5 to 7 days later in the Ovsynch- and/or CO-Synch-based protocols [10,11]. The mechanisms of action of PGF_{2α} in newly formed CL are not completely understood [6,7,9].

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Studies reported that luteolysis occurred in more than 90% in cows that received a single (25 mg) dose of PGF_{2α} after Day 8 of the estrous cycle [11]. In contrast, a single (25 mg) dose of PGF_{2α} in the first 4 days of the cycle did not induce luteolysis [6,7]. However, the incidence of luteolysis induced by a double dose of PGF_{2α} (50 mg) around Day 5 of the estrous cycle is not well established [4,5,7]. In addition, studies have demonstrated that a transient decrease in circulating progesterone (P₄) concentration in cows does not always indicate complete luteolysis when PGF_{2α} is administered around Day 5 of the estrous cycle [5,6]. This response is referred to as partial luteolysis and is characterized as a decrease in P₄ concentration during the first 24 hours after PGF_{2α}, but after that, no further decline or, in some cases, a rebound in systemic concentrations occur [5,6,12].

Ensuring complete luteolysis and total decrease in circulating P₄ at the time of artificial insemination (AI) is essential for optimizing results in fixed timed AI (FTAI) protocols [13]. To induce complete luteolysis in recently formed CL, several strategies have been studied, including the use of a greater dose of PGF_{2α} and/or multiple doses of PGF_{2α} given at varying intervals [6,7].

In recent years, incidence of luteolysis during the first days of the estrous cycle has been investigated in multiple experiments [6,7]. All these studies have used *Bos taurus* cows as the animal model. The objective of the present study was to investigate the efficacy of differing PGF_{2α} doses to induce luteolysis on Day 5 or Day 7 of the estrous cycle in Nelore cows.

2. Materials and methods

The experiment was performed in the Experimental Station Agrozootécnica Hildegard Georgina Von Pritzelwiltz (23°34'25" S; 50°58'17" W), in Londrina, Paraná, Brazil, which belongs to the Fundação de Estudos Agrários Luiz de Queiroz.

Nonlactating, cyclic Nelore cows (n = 323) with no uterine or ovarian abnormalities (based on a transrectal scan of the reproductive tract by ultrasonography) were used. Groups of approximately 80 cows at random stages of their estrous cycle received 25.0 mg of PGF_{2α} (dinoprost tromethamine–Lutalyse, Zoetis, São Paulo, SP, Brazil). Estrus detection was performed twice daily for 5 days; aided by vasectomized bulls fitted with chin–ball markers.

Cows detected in estrus were assigned, on the day of estrus, to receive one of three doses of PGF_{2α} on either Day 5 or Day 7 of the ensuing estrous cycle. Doses were 12.5, 25.0, or 50.0 mg of PGF_{2α} given on Day 5 or Day 7 of the estrous cycle. The 3 × 2 factorial arrangement resulted in six treatments: Day 5/12.5PGF_{2α} (n = 54), Day 5/25.0PGF_{2α} (n = 54), Day 5/50.0PGF_{2α} (n = 53), Day 7/12.5PGF_{2α} (n = 54), Day 7/25.0PGF_{2α} (n = 53), and Day 7/50.0PGF_{2α} (n = 55).

Ovulation was confirmed by detection of CL by ultrasonography (US) on Day 5 or Day 7, and CL diameter was recorded when PGF_{2α} was given (0 hours) and at 96 hours to assess morphologic luteolysis. Estrus detection was performed twice daily, aided by the use of vasectomized bulls fitted with chin–ball markers for a period of 120 hours.

Blood samples (10 mL) were collected via a coccygeal vessel at 0, 24, 48, and 72 hours after PGF_{2α} administration.

Blood samples were centrifuged for 15 minutes at 1800 × g. Serum was frozen at –20 °C for further analysis. Progesterone concentrations were measured by chemiluminescent assay using commercial IMMULITE 1000 kits (Siemens Healthcare Diagnostics, Deerfield, IL, USA). All the analyses were performed in the Nutrition and Animal Reproduction Laboratory–LZT/ESALQ/USP. The coefficients of variation were 2.7% and 3.6% for low and high adjuster, respectively.

Luteolysis was determined to have occurred when P₄ concentrations were less than 1.0 ng/mL (L1.0) or less than 0.5 ng/mL (L0.5) 72 hours after PGF_{2α}. When P₄ concentrations declined during the first 24 hours after treatment but did not decline further, or in some cases increased, this was classified as partial luteolysis. If P₄ concentration did not decrease during the 24 hours after PGF_{2α}, this was classified as failure of luteolysis.

A randomized experimental design in a 3 × 2 factorial arrangement (day of estrous cycle × PGF_{2α} dose) was used. The variables were analyzed after running normality (Shapiro–Wilk) and homogeneity of variance (Welch) tests. Variables with non-normal distribution were submitted to either logarithmic (log₁₀ [x + 1]) or quadratic (SQRT [x + 1/2]) transformation before analysis. Variables such as CL diameter at 0 and 96 hours after PGF_{2α} administration and the interval to estrus were analyzed using the MIXED procedure. The same procedure with repeated measures over time analyses was used to evaluate P₄ concentrations at 0, 24, 48, and 72 hours after PGF_{2α} in each day of the estrous cycle (Day 5 and Day 7), separately, according to PGF_{2α} dose. Three analyses of P₄ concentrations in each day of the estrous cycle (Day 5 and Day 7) were performed. The first analysis considered all cows, the second, only cows with complete luteolysis, and the third considering only cows with partial luteolysis. The GLIMMIX procedure was used for variables with binomial distribution, such as L0.5, L1.0, partial luteolysis rate, no luteolysis, and incidence of estrus.

The effect of relative drop in circulating P₄ concentration (100 – [P₄ concentration at 24 hours after PGF_{2α} × 100]/P₄ concentration at 0 hour at PGF_{2α}) on estrous behavior was analyzed by logistic regression (LIFETEST procedure) comparing PGF_{2α} doses or days of the estrous cycle when PGF_{2α} treatments were employed. The oddsratio statement was used to compare difference among doses of PGF_{2α}. Logistic regression curves were done using the coefficient generated by Interactive Data Analysis of SAS (logit = intercept + slop × [drop in P₄]). Probability curve was made by Y = (EXP [logit]/1 + EXP [logit]) × 100. Significant differences were considered when P < 0.05. All procedures used were from SAS 9.3 statistical software.

3. Results

There was no interaction between PGF_{2α} doses and day of the estrous cycle in any of the analyzed variables. Dose of PGF_{2α} did not alter the time of estrus after PGF_{2α} (70.2 ± 1.76 hours), and 50.4% of the cows that exhibited estrus showed estrus within 72 hours after PGF_{2α} (Fig. 1). However, the mean time of estrus was later (P < 0.01) in cows treated on Day 5 than those on Day 7 (76.0 ± 3.12 vs. 66.0 ± 2.29 hours, respectively). Cows treated with PGF_{2α} on Day 7 were more likely to exhibit estrus (P < 0.01) than

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