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#### ARTICLE INFO

### ABSTRACT

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Key words: Simple sequence repeat Insertion and deletion Genome evolution Molecular mechanism Phage divergence Human gene mutations Phages are the most abundant biological entities and influence prokaryotic communities on Earth. Comparing closely related genomes sheds light on molecular events shaping phage evolution. Simple sequence repeat (SSR) variations impart over half of the genomic changes between T7M and T3, indicating an important role of SSRs in accelerating phage genetic divergence. Differences in coding and noncoding regions of phages infecting different hosts, coliphages T7M and T3, Yersinia phage  $\phi$ YeO3-12, and Salmonella phage  $\phi$ SG-JL2, frequently arise from SSR variations. Such variations modify noncoding and coding regions; the latter efficiently changes multiple amino acids, thereby hastening protein evolution. Four classes of events are found to drive SSR variations: insertion/deletion of SSR units, expansion/contraction of SSRs without alteration of genome length, changes of repeat motifs, and generation/loss of repeats. The categorization demonstrates the ways SSRs mutate in genomes during phage evolution. Indels are common constituents of genome variations and human diseases, yet, how they occur without preexisting repeat sequence is less understood. Non-repeat-unit-based misalignment-elongation (NRUBME) is proposed to be one mechanism for indels without adjacent repeats. NRUBME or consecutive NRUBME may also change repeat motifs or generate new repeats. NRUBME invoking a non-Watson-Crick base pair explains insertions that initiate mononucleotide repeats. Furthermore, NRUBME successfully interprets many inexplicable human di- to tetranucleotide repeat generations. This study provides the first evidence of SSR variations expediting phage divergence, and enables insights into the events and mechanisms of genome evolution. NRUBME allows us to emulate natural evolution to design indels for various applications.

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#### 1. Introduction

Phages have recently been utilized in targeted gene delivery, biocontrol of foodborne pathogens, and therapeutics for animal and human infections [1-3]. They are the most abundant entities in the biosphere with vastly diverse sequences [4]. The mechanisms by which the genetic diversity arises have been reviewed [4,5]. Genomic mosaicism is a pervasive feature of phage genomes [6]. Genetic exchanges by different recombination processes contribute to the genomic mosaicism [7–11]. In addition, duplication

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http://dx.doi.org/10.1016/j.mrfmmm.2016.04.001 0027-5107/© 2016 Elsevier B.V. All rights reserved. of genes, inversion of genomic segments, insertion of mobile elements, and point mutations also cause phage evolution [5,12–14]. Whether there are other mechanisms important to phage evolution awaits ascertainment.

Eukaryotic genomes harbor widespread simple sequence repeats (SSRs) of 1–6 base repeat units [15]. Prokaryotes have high genomic gene density and significantly less SSR repeat numbers and length relative to eukaryotes [16,17]. SSRs have been discovered in different eukaryotic viruses and utilized for virus genotyping [18–20]. For prokaryotic viruses, repeat sequences in the lysozyme gene of T4 phage have been employed for studying indel mutations [21]. Because bacteriophages have compact genomes with many functional constraints, the occurrence, variety, significance, and potential applications of SSRs in whole genomes need in-depth investigation.

Nucleotide indels are common in genomes and may affect human diseases [22]. Indels of repeat sequences have been proposed to occur by Streisinger slippage [21,23]. For deletions of nonrepeat nucleotides, mechanisms of misincorporation misalignment [24], dNTP stabilized misalignment [25], and transient

Abbreviations: SSR, simple sequence repeat; NRUBME, non-repeat-unit-based misalignment-elongation; T7M, T7 Meselson; DTRs, direct terminal repeats; RA, relative abundance; RD, relative density; RCRs, recombinant regions; NRCRs, non-recombinant regions; nWC, non-Watson-Crick; PDH, pyruvate dehydrogenase; XP, xeroderma pigmentosum.

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	A		
	T7M	23	GTACCCCCCC-ATAGCCCCTCTT
	тЗ	23	<b>GTACCCCCC</b> CATAGCCCCTCTT
	<b>∮YeO3-12</b>	23	GTACCCCCCATAGCCC-TCTT
	¢SG-JL2	23	GTACCCCCCATAGCCC-TCCT
			*****
F	3		
	т7 <b>М</b>	25492	TACGAGGGGGGGGGGGTTA
	т3	25490	TACGAAGGGGGGGGGGGGGGGGGTTA
	φYeO3-12	26256	TACGAGGGGTTA
	¢SG-JL2	25895	TACGAGGGGTTA
			****

**Fig. 1.** Indels in the noncoding regions of phages. (A) Indels of the C repeats in the terminal repeats. (B) Indels of the G repeats and the A between  $\phi$ 13 and gene 13. The sequences of four phages are aligned. The numbers shown on the left side are the nucleotide numbers in the genome of each phage. The differences between T7M and T3 are shown in gray shade.

F	ł								
	т7м	9603	GCGCGA	GGTGTACGC	AAGGI	CGGG	GCT		
	тЗ	9604	GCGA	GGTGTACGC	AAGGI	CGGG	-СТ		
	<b>∮YeO3-12</b>	10335	GCGCGA	GGTGTACGC	AAGGI	CGGGG	GCT		
	¢SG-JL2	10549	GCGCGA	GGTGTACGC	AAGGI	CGGGG	GCT		
			*** *	******	* * * * *	****	**		
B									
-	Т7М	9963	CCAGTG	GCGTGGC-T	САААС	GAA			
	тЗ	9961	CCAGTG	GC-TGGCGT	САААС	GAA			
	<b>∳YeO3-12</b>	10695 CCGGTGGAGTGGC-TCAAAGAA 10909 CCGGTGGAGTGGC-TCAAAGAA							
	¢SG-JL2								
			** ***	* **** *	****	***			
C			1	<b>`</b>					
	т7м	7 ARGV	RKVGA	т7м	127	PVAW	LKE		
	тЗ	7 AR-C	TQGRA	тЗ	126	PVAG	VKE		
	¢¥e03-12	7 ARGV	RKVGA	<b>∮YeO3-12</b>	127	PVEW	LKE		
	¢SG-JL2	7 ARGV	RKVGA	¢SG-JL2	127	PVEW	LKE		
		**	: *			**	: * *		

**Fig. 2.** Sequence changes in gene 3 of T7M and T3. (A) Indels in the CG and G repeats. (B) Indels of the nonrepeat nucleotide G. (C) Amino acid sequence change caused by (A). (D) Amino acid sequence change caused by (B). Sequences of gene 3 of  $\phi$ YeO3-12 and  $\phi$ SG-JL2 are also aligned for comparison. Positions of interest are shaded in grey for T7M and T3. The numbers shown on the left side are the genomic nucleotide numbers or gp3 residue numbers of each phage.

dislocation [26] have been proposed. The mechanism of nucleotide insertion at a position that does not manifest adjacent repeat sequence is poorly understood [27]. Furthermore, how a nonrepeat sequence generates its first repeated copy is unclear.

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