

Genotype and Tumor Locus Determine Expression Profile of Pseudohypoxic Pheochromocytomas and Paragangliomas^{1,2}

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Abbreviations: AT, abdominal/thoracic; CSS, Carney-Stratakis Syndrome; HIF2 α , hypoxia-inducible factor 2 α ; HN, head/neck; IPA, Ingenuity Pathway Analysis; NF1/NF1, neurofibromatosis 1 syndrome/gene; OXPHOS, oxidative phosphorylation; PAMR, prediction analysis for microarray; PGL, paraganglioma; PGL1, 2, 3, 4, familial PGL types 1, 2, 3, 4; PHD, prolyl hydroxylase; PHD2/EGLN1, prolyl hydroxylase 2; PHEO, pheochromocytoma; qRT-PCR, quantitative real-time polymerase chain reaction; SAM, significance analysis of microarray; SDH, succinate dehydrogenase; SDHA, SDH subunit A; SDHAF2, SDH complex assembly factor 2; SDHB, SDH subunit B; SDHC, SDH subunit C; SDHD, SDH subunit D; TMEM127, transmembrane protein 127; VHL/VHL, von Hippel-Lindau syndrome/gene

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Abstract

Pheochromocytomas (PHEOs) and paragangliomas (PGLs) related to mutations in the mitochondrial succinate dehydrogenase (SDH) subunits A, B, C, and D, SDH complex assembly factor 2, and the von Hippel-Lindau (VHL) genes share a pseudohypoxic expression profile. However, genotype-specific differences in expression have been emerging. Development of effective new therapies for distinctive manifestations, e.g., a high rate of malignancy in SDHB- or predisposition to multifocal PGLs in SDHD patients, mandates improved stratification. To identify mutation/location-related characteristics among pseudohypoxic PHEOs/PGLs, we used comprehensive microarray profiling (SDHB: $n = 18$, SDHD-abdominal/thoracic (AT): $n = 6$, SDHD-head/neck (HN): $n = 8$, VHL: $n = 13$). To avoid location-specific bias, typical adrenal medulla genes were derived from matched normal medullas and cortices ($n = 8$) for data normalization. Unsupervised analysis identified two dominant clusters, separating SDHB and SDHD-AT PHEOs/PGLs (cluster A) from VHL PHEOs and SDHD-HN PGLs (cluster B). Supervised analysis yielded 6937 highly predictive genes (misclassification error rate of 0.175). Enrichment analysis revealed that energy metabolism and inflammation/fibrosis-related genes were most pronouncedly changed in clusters A and B, respectively. A minimum subset of 40 classifiers was validated by quantitative real-time polymerase chain reaction (quantitative real-time polymerase chain reaction vs. microarray: $r = 0.87$). Expression of several individual classifiers was identified as characteristic for VHL and SDHD-HN PHEOs and PGLs. In the present study, we show for the first time that SDHD-HN PGLs share more features with VHL PHEOs than with SDHD-AT PGLs. The presented data suggest novel subclassification of pseudohypoxic PHEOs/PGLs and implies cluster-specific pathogenic mechanisms and treatment strategies.

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Introduction

Predispositions to certain tumors have been linked to an ever-increasing number of mutations. To date, mutations in 11 different genes have been associated with development of paragangliomas (PGLs), which are catecholamine-producing, chromaffin cell tumors, including adrenal pheochromocytomas (PHEOs). Initially, discovery of mutations was guided by syndromic presentation and family history; however, more recently discovered mutations can present in seemingly sporadic fashion. Known PHEO/PGL susceptibility genes are *von Hippel-Lindau* (VHL) and *neurofibromatosis 1* (NF1) in the homonymous syndromes (VHL and NF1, respectively), *RET proto-oncogene* in multiple endocrine neoplasia type 2, *succinate dehydrogenase D* (SDHD) in familial PGL type 1 (PGL1) and Carney-Stratakis Syndrome (CSS), *SDHC* in PGL3 and CSS, *SDHB* in PGL4 and CSS, *SDH complex assembly factor 2* (SDHAF2) in PGL2, *prolyl hydroxylase 2* (PHD2/EGLN1), *transmembrane protein 127* (TMEM127), *kinesin family member 1B*, *SDHA*, and *MYC-associated factor X* (reviewed in [1]). Most recently, activating mutations in hypoxia-inducible factor 2 α (HIF2 α) have been associated with PGL and polycythemia [2].

Notwithstanding the multitude of susceptibility genes, mutation-derived PHEOs/PGLs separate into merely two main clusters, one containing *SDHA*, *SDHB*, *SDHC*, *SDHD*, *SDHAF2*, and *VHL* and the other consisting of *NF1*, *RET proto-oncogene*, *TMEM127*, *kinesin family member 1B*, and *MYC-associated factor X* mutation-derived PHEOs/PGLs (reviewed in [1,3]). The PHEOs/PGLs of the first mentioned cluster are characterized by a pseudohypoxic phenotype, i.e., inappropriate stabilization of HIF α subunits under normoxia (reviewed in [1,3]).

Under normoxia, hydroxylation of HIF α by PHDs (PHD1/EGLN2, PHD2/EGLN1, and PHD3/EGLN3) designates them for VHL-dependent ubiquitylation and subsequent degradation [4]. Accordingly, *VHL* mutations can promote HIF α stabilization. Similarly, SDH dysfunction causes HIF stabilization by succinate or reactive oxygen species accumulation-mediated PHD inhibition [5,6].

Despite increasing evidence for differences within the pseudohypoxic cluster [7–9], the molecular basis for distinct clinical behaviors including the preferential site of tumor development, biochemical phenotype, or metastatic potential remains largely unknown. Mutations in *SDHB* have been associated with extra-adrenal PGLs and high risk of malignancy [1,3]. *SDHD* mutations, however, predispose to multifocal PHEOs/PGLs, primarily from the head and neck (HN) region, with low metastatic risk [3]. However, HN PGL can be inoperable, or surgery can lead to severe side effects due to close vicinity to major blood vessels and nerves. *VHL* mutation-derived PHEOs/PGLs are almost always adrenal, non-metastatic, but frequently bilateral and/or recurrent, thus adrenal sparing treatment options are of high importance [3]. *SDHA*, *SDHC*, and *SDHAF2* mutation-derived PHEOs/PGLs are rare and have not yet been characterized in detail. Large cohort studies including PHEO/PGL patients revealed that *SDHA*, *SDHC*, and *SDHAF2* are extremely rare (0.1–0.5%, 0.4–2.2%, and non-detectable, respectively) [9–13]. In a Dutch population of HN PGL patients, *SDHAF2* mutations were found in 4% and *SDHC* mutations in 0.4% of cases (*SDHA* not tested) [14].

To date, genetic testing presents an important diagnostic tool for risk assessment of PHEO/PGL patients and their families. However, genetic testing is cost intensive and targeted therapeutic options for

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