

Dopamine transporter binding in rat striatum: a comparison of [O-methyl-¹¹C]β-CFT and [N-methyl-¹¹C]β-CFT

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Received 11 June 2008; received in revised form 13 August 2008; accepted 15 October 2008

Abstract

Introduction: Positron emission tomography scanning with radiolabeled phenyltropane cocaine analogs is important for quantifying the in vivo density of monoamine transporters, including the dopamine transporter (DAT). [¹¹C]β-CFT is useful for studying DAT as a marker of dopaminergic innervation in animal models of psychiatric and neurological disorders. [¹¹C]β-CFT is commonly labeled at the *N*-methyl position. However, labeling of [¹¹C]β-CFT at the *O*-methyl position is a simpler procedure and results in a shorter synthesis time [desirable in small-animal studies, where specific activity (SA) is crucial]. In this study, we sought to validate that the *O*-methylated form of [¹¹C]β-CFT provides equivalent quantitative results to that of the more commonly reported *N*-methyl form.

Methods: Four female Sprague–Dawley rats were scanned twice on the IndyPET II small-animal scanner, once with [*N*-methyl-¹¹C]β-CFT and once with [*O*-methyl-¹¹C]β-CFT. DAT binding potentials ($BP = B'_{avail}/K_d$) were estimated for right and left striata with a nonlinear least-squares algorithm, using a reference region (cerebellum) as the input function.

Results: [*N*-Methyl-¹¹C]β-CFT and [*O*-methyl-¹¹C]β-CFT were synthesized with 40–50% radiochemical yields (HPLC purification). Radiochemical purity was >99%. SA at end of bombardment was 258±30 GBq/μmol. Average BP values for right and left striata with [*N*-methyl-¹¹C]β-CFT were 1.16±0.08 and 1.23±0.14, respectively. BP values for [*O*-methyl-¹¹C]β-CFT were 1.18±0.08 (right) and 1.22±0.16 (left). Paired *t* tests demonstrated that labeling position did not affect striatal DAT BP.

Conclusions: These results suggest that [*O*-methyl-¹¹C]β-CFT is quantitatively equivalent to [*N*-methyl-¹¹C]β-CFT in the rat striatum.

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Keywords: Positron emission tomography; Dopamine transporter; [¹¹C]β-CFT; Striatum; Dopamine; Animal models

1. Introduction

Positron emission tomography (PET) scanning with radiolabeled phenyltropane cocaine analogs allows quantification of in vivo neuronal monoamine uptake sites. The most commonly employed radioligands, β-CIT [2β-carbomethoxy-3β-(4'-iodophenyl)tropane, RTI-55] [1] and β-CFT [2β-carbomethoxy-3β-(4'-fluorophenyl)tropane, WIN 35,428 (3)] [2], have a high affinity for the dopamine transporter (DAT) and are used as markers for striatal central dopaminergic innervation [3–10]. [¹¹C]β-CFT is widely

used to study DAT in animal models of psychiatric and neurological disorders and in human diseases such as Parkinson's disease [11–13]. The *N*-methylated form of [¹¹C]β-CFT is more commonly seen in the literature, as the precursor used for radiolabeling, nor-β-CFT [2β-carbomethoxy-3β-(4'-fluorophenyl)nortropane (5)], is commercially available. However, it is more cost-effective to synthesize precursor *de novo*. We pursued the use of [*O*-methyl-¹¹C]β-CFT [13–15] for several reasons: (a) the synthesis of β-CFT-acid [2β-carboxylic acid-3β-(4'-fluorophenyl)tropane (4)] for the production of [*O*-methyl-¹¹C]β-CFT is simpler than that of nor-β-CFT; (b) synthesis of β-CFT-acid results in higher chemical yields than the synthesis of nor-β-CFT; and (c) labeling of [¹¹C]β-CFT at the *O*-methyl position results in a slightly shorter synthesis

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time, which is advantageous in small-animal PET studies where specific activity (SA) is crucial.

Our group and others have successfully labeled the cocaine congener β -CIT at both nitrogen and oxygen positions [16,17]. Lundkvist et al. [17] reported that the late-time striatum:cerebellum ratios were similar for [*N*-methyl- ^{11}C] β -CIT and [*O*-methyl- ^{11}C] β -CIT. As β -CIT is chemically similar to β -CFT, the semiquantitative observation of Lundkvist et al. suggests that [*N*-methyl- ^{11}C] β -CFT and [*O*-methyl- ^{11}C] β -CFT may produce similar estimates of DAT binding. The purpose of this study was to determine if estimates of DAT binding in rat striata with [^{11}C] β -CFT labeled at the *O*-methyl position were equivalent to those from [*N*-methyl- ^{11}C] β -CFT.

2. Materials and methods

2.1. General

All commercial reagents and solvents were used without further purification. [^{11}C]Methyl triflate ([^{11}C]CH₃OTf) was prepared according to a literature procedure [18]. ^1H NMR spectra were recorded on a Bruker QE 300 NMR spectrometer using tetramethylsilane (TMS) as an internal standard. Chemical shift data for the proton resonances were reported in parts per million (δ scale) relative to internal standard TMS (δ 0.0), and coupling constants (*J*) were reported in hertz. All moisture- and/or air-sensitive reactions were performed under a positive pressure of nitrogen maintained by a direct line from a nitrogen source. Analytical HPLC was performed using a Prodigy (Phenomenex) 5- μm C-18 column, 4.6 \times 250 mm; 3:1:3 CH₃CN/MeOH/20 mM, pH 6.7, phosphate (buffer solution) mobile phase; 1.5 ml/min flow rate; and UV (254 nm) and gamma ray (NaI) flow detectors. Semipreparative HPLC was performed using a YMC-Pack ODS-A, S-5 μm , 12 nm, 10 \times 250 mm i.d. (Waters) C-18 column; 20% EtOH/80% 20 mM H₃PO₄ mobile phase, 5.0 ml/min flow rate, UV (254 nm) and gamma ray (NaI) flow detectors. Sterile Millex-GS 0.22 μm vented filter unit was obtained from Millipore Corporation (Bedford, MA).

The overall scheme of synthesis is depicted in Fig. 1.

2.1.1. Precursors and β -CFT standard syntheses

Hydrolysis of cocaine under acid conditions gave 2 β -carboxylic acid-3 β -hydroxytropane (**1**), which was used in the next step reaction without further purification. Compound **1** was reacted with POCl₃ and MeOH to provide anhydroecgonine methyl ester (**2**) in 86% yield [16,19]. ^1H NMR (CDCl₃): δ 6.81 (t, 1H, *J*=3.0 Hz), 3.78 (d, 1H, *J*=5.6 Hz), 3.74 (s, 3H, OCH₃), 3.24 (t, 1H, *J*=5.4 Hz), 2.69–2.62 (d, br, 1H, *J*=19.8 Hz), 2.35 (s, 3H, NCH₃), 2.20–2.07 (m, 2H), 1.88–1.81 (dd, 1H, *J*=19.8 Hz, 4.0 Hz), 1.80–1.26 (m, 2H).

The conjugate addition of Grignard reagent 4-fluorophenylmagnesium bromide to Compound **2** after acid workup [19] formed a mixture of two phenyltropane isomers: the biologically potent β -CFT (Compound **3**) and the less

biologically-active 2 α -carbomethoxy-3 β -(4'-fluorophenyl) tropane (α -CFT) [20] (45% and 17% yields, respectively). In addition to being an intermediate in the synthesis of precursors, β -CFT served as the standard compound. β -CFT ^1H NMR (CDCl₃): δ 7.23–7.19 (m, 2H), 6.98–6.92 (m, 2H), 3.57–3.55 (m, 1H), 3.50 (s, 3H, OCH₃), 3.38–3.37 (d, br, 1H, *J*=3.0 Hz), 3.02–2.94 (m, 1H), 2.88–2.86 (d, br, 1H, *J*=6.0 Hz), 2.63–2.53 (m, 1H), 2.23 (s, 3H, NCH₃), 2.18–2.03 (m, 2H), 1.78–1.56 (m, 3H). α -CFT ^1H NMR (CDCl₃): δ 7.25–7.20 (m, 2H), 6.97–6.91 (m, 2H), 3.51 (s, 3H, OCH₃), 3.44–3.42 (d, br, 1H, *J*=6.0 Hz), 3.28–3.26 (m, 1H), 3.12–3.09 (m, 2H), 2.42 (s, 3H, NCH₃), 2.14–2.08 (ddd, 1H, *J*=18.0 Hz), 1.98–1.82 (m, 3H), 1.76–1.58 (m, 2H).

Hydrolysis of β -CFT (Compound **3**) under neutral conditions (50% dioxane/H₂O) [21] provided precursor β -CFT-acid (Compound **4**) in 97% yield. ^1H NMR (CDCl₃): δ 7.24–7.19 (m, 2H), 7.01–6.95 (m, 2H), 3.61–3.58 (m, 2H), 3.21–3.12 (m, 1H), 2.67–2.56 (m, 2H), 2.51 (s, 3H, NCH₃), 2.31–2.28 (m, 2H), 2.01–1.94 (m, 2H), 1.82–1.74 (m, 1H).

N-Desmethylation of Compound **3** with 1-chloroethyl chloroformate [19] afforded another precursor, nor- β -CFT (Compound **5**), in 75% yield. ^1H NMR (CDCl₃): δ 7.18–7.15 (t, 2H, *J*=9.0 Hz), 6.97–6.94 (t, 2H, *J*=9.0 Hz), 3.78–3.59 (m, 2H), 3.40 (s, 3H, OCH₃), 3.24–3.21 (m, 1H), 2.68–2.39 (m, 3H), 2.26–2.20 (m, 1H), 1.98–1.95 (m, 1H), 1.80–1.58 (m, 3H).

2.1.2. Radiochemistry of [*O*-methyl- ^{11}C] β -CFT and [*N*-methyl- ^{11}C] β -CFT

Radiosynthesis was performed in an automated, multi-purpose ^{11}C -radiosynthesis module, which allows measurement of SA during synthesis [22,23]. [^{11}C]CO₂ was produced by the $^{14}\text{N}(\text{p},\alpha)^{11}\text{C}$ nuclear reaction in a small-volume (9.5 cm³) aluminum gas target (CTI) from 11 MeV proton cyclotron on research purity nitrogen (+1% O₂) in a Siemens radionuclide delivery system (Eclipse RDS-111). The precursor β -CFT-acid (Compound **4**, 0.1 mg) was dissolved in CH₃CN (300 μl). To this solution was added 3 N NaOH (2 μl). The mixture was transferred to a small reaction vial. No-carrier-added (high SA) [^{11}C]CH₃OTf that was produced by the gas-phase production method [18] from [^{11}C]CO₂ through [^{11}C]CH₄ and [^{11}C]CH₃Br with silver triflate (AgOTf) column was passed into the reaction vial, which was cooled to 0°C, until radioactivity reached a maximum (~2 min) and then the reaction vial was isolated and heated at 80°C for 3 min. The contents of the reaction tube were diluted with NaHCO₃ (1 ml, 0.1 M) and injected onto the semipreparative HPLC column with 2 ml injection loop. The product fraction was collected, the solvent was removed by rotatory evaporation under vacuum and the final product, [*O*-methyl- ^{11}C] β -CFT (*O*-[^{11}C]**3**), was formulated in saline, sterile filtered through a sterile vented Millex-GS 0.22- μm cellulose acetate membrane and collected into a sterile vial containing 8% NaHCO₃ solution (0.25 ml) for adjustment of pH of the product. Total radioactivity was assayed and total volume was noted for

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