



Biofabrication, characterization, and possible bio-reduction mechanism of platinum nanoparticles mediated by agro-industrial waste and their catalytic activity



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ARTICLE INFO

Article history:

Received 25 March 2014
Received in revised form 2 July 2014
Accepted 9 July 2014
Available online 17 July 2014

Keywords:

Punica granatum
Biofabrication
Platinum
Nanoparticle
Reduction

ABSTRACT

The present study showed biofabrication of platinum nanoparticles (Pt-NPs) using agro-industrial waste *Punica granatum*'s peel extract. Appearance of the broad spectrum from visible to the ultraviolet region, confirmed the biofabrication of Pt-NPs. Pt-NPs were spherical, within size range of 16–23 nm. XRD suggested the fabrication of crystalline Pt-NPs with (111) plane in predominant orientation. The negative ζ potential value of colloidal Pt-NPs revealed high stability. FTIR confirmed the role of hydroxyl and carbonyl groups of polyphenolic compounds of peel extract for biofabrication. The reduction of anthropogenic pollutant, 3-nitrophenol, by NaBH_4 using colloidal Pt-NPs established it as an efficient “green catalyst.”

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Introduction

Today, transition metal nanoparticles, particularly platinum nanoparticles (Pt-NPs) are subject to intensive research. The emerging catalytic applications of Pt-NPs in variety of reactions such as hydrogenation [1], oxidation [2], reduction [3], and for the synthesis of organic dyes [4] has attracted the focus of researchers. The biofabricated nanoparticles for their eco-friendly properties are the preferred option for variety of applications. The development of simple and environment-friendly methods for controlled synthesis of Pt-NPs is therefore, important, not only for the fascinating utilization of Pt-NPs but also for the demands of green chemistry.

In recent years, significant efforts have been made toward fabrication of nanoparticles using biogenic resources, which also represents a growing connection between biotechnology and nanotechnology [5]. This approach for nanoparticle fabrication shows several benefits concerning biocompatibility, thermal and chemical stability, high efficiency, fast process, cost efficiency, and eco-friendly nature [6,7]. However, till date plant-based synthesis of Pt-NPs has been reported only by leaf extract of *Ocimum sanctum*

[8], *Diospyros kaki* [9], *Cacumen platycladi* [10], and *Anacardium occidentale* [11]. Few other biological resources are also reported for biological synthesis of Pt-NPs such as horse spleen apoferritin [12] and honey [4].

Recently, use of *Punica granatum* (*P. granatum*) peel has been reported for cost-effective and environmentally benign synthesis of Ag [13] and Au-NPs [14]. Earlier, leaf [15] and fruit [16] of *P. granatum* were also utilized for biofabrication of Au and Ag-NPs. *P. granatum* peel is one of the most valuable by-products of the food and agriculture industry, which is mainly composed of ellagic tannins, punicalagin, gallic acid, ellagic acid, and quercetin [17,18]. It is also shown antioxidant [19,20], antimutagenic [21], and chemo-preventive potential [22]. An extensive literature survey revealed that there is no report available for the biosynthesis of Pt-NPs using *P. granatum* peel. In the present study, antioxidant potential of *P. granatum* peel is utilized for biofabrication of Pt-NPs.

The present study involves biofabrication of Pt-NPs using agro-industrial waste *P. granatum* peel as the bio-reducing agent. Further, catalytic activity of biofabricated colloidal Pt-NPs is also investigated for reduction of anthropogenic pollutant 3-nitrophenol (3-NP) using sodium borohydride (NaBH_4) as a hydrogen or electron donor. Nowadays, nitrophenols are extensively used as raw materials in pharmaceutical, dye, and insecticide industries [23]. Since nitrophenols are readily soluble and stable in water [24] they are abundantly present in agricultural and industrial waste water. Further, they are perilous to public

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health as they possess mutagenic and carcinogenic properties harmful for humans [25]. It is therefore, necessary to eliminate nitrophenols from industrial waste water. The reduction of 3-NP to 3-aminophenol (3-AP) is therefore, of significant importance. Use of eco-friendly biofabricated colloidal Pt-NPs as a catalyst in this direction is of enormous value in environmental and industrial aspect. Biofabricated Pt-NPs are characterized by UV-visible spectrophotometer (UV-visible), dynamic light scattering (DLS), X-ray diffraction (XRD), transmission electron microscopy (TEM), energy-dispersive X-ray spectroscopy (EDX), and Fourier transform infrared spectroscopy (FTIR) techniques. Further, the reduction of 3-NP is monitored by UV-visible spectrophotometer.

Experimental

Materials

Chemicals, including hexachloroplatinic acid ($\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$) 99.9%, 3-nitrophenol ($\text{C}_6\text{H}_5\text{NO}_3$) 98%, and sodium borohydride (NaBH_4) 98% were procured from HiMedia Pvt. Ltd, Mumbai, India. Peels of the *P. granatum* fruit were collected from the local market of Surat, Gujarat, India. Double-sterilized Milli-Q water was used throughout the experiments.

Preparation of the *P. granatum* peel extract and biofabrication of colloidal Pt-NPs

A total of 30 g of fresh peel of *P. granatum* was extracted with 120 mL of distilled water at 60 °C for 10 min and filtered. The solution was decanted and stored at 4 °C for further use. About 100 mL of aqueous *P. granatum* peel extract was added to 400 mL of 1×10^{-3} M $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ solution. The mixture was maintained at 90 °C in a sealed flask for 30 min under shaking conditions on a rotary shaker (500 rpm). The reduced Pt-NPs were sonicated for 10 min to separate Pt-NPs from the biomolecules present in *P. granatum* peel extract. After sonication, Pt-NPs were purified by repeated centrifugation at 14,000 rpm for 10 min and the pellets were washed thrice with distilled water to remove the impurities. Control reactions, in which $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ solution and peel extract was kept in a separate conical flask, under the same reaction conditions.

Evaluation of catalytic activity of biofabricated colloidal Pt-NPs

In order to find out the catalytic activity of biofabricated colloidal Pt-NPs (Pt-NPs dispersed in *P. granatum* peel extract) three typical reactions were carried. In the first reaction, 1 mL of 1×10^{-3} M 3-NP was mixed with 0.5 mL of water. In the second reaction, 0.5 mL of 1 M NaBH_4 was added to the first reaction mixture. In the third reaction, 1 mL of colloidal Pt-NPs was mixed with reaction mixture obtained from the second reaction. All the three reactions were monitored by UV-visible spectrophotometer (DR 5000, HACH, USA). Temperature and concentration dependent catalytic activity of biofabricated colloidal Pt-NPs were also carried out for 3-NP reduction. Catalytic activity of *P. granatum* peel extract and bulk $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ was also evaluated for 3-NP reduction.

Characterization of Pt-NPs

Biofabrication of Pt-NPs was monitored first by visual inspection and then by using UV-visible spectrophotometer (DR 5000, HACH, USA). Baseline correction was made with *P. granatum* peel extract. Hydrodynamic size distribution and ζ potential of the colloidal Pt-NPs were determined by using DLS (Zetasizer Nano

ZS90, Malvern, UK). TEM and selected area electron diffraction (SAED) pattern data were obtained by using TEM (CM-200, Philips, UK). TEM image was recorded at 100 kV accelerating voltage with resolution of 2.4 Å. XRD pattern of Pt-NPs on the glass substrate was recorded by using XRD (X'Pert Pro, PANalytical, Holland) operated at a voltage of 45 kV and current of 35 mA with $\text{Cu-K}\alpha$ radiation ($K = 1.5406 \text{ \AA}$). The scanning range (2θ) was selected from 20 ° to 80 ° at 0.045 °/min continuous speed. The crystallite size of the Pt-NPs was calculated using Scherrer's formula. The natures of elements were identified by EDX (INCA X-sight, Oxford Instruments, UK) coupled with scanning electron microscopy (SEM) (JSM-6380LV, JEOL, Japan). In order to identify the phytochemicals responsible for bio-reduction and stabilization of Pt-NPs, FTIR analysis of peel extract before and after bio-reduction and biogenic Pt-NPs were carried out using FTIR (Magna-550, Nicolet, USA). All spectra was taken in the mid-IR region of 600–3600 cm^{-1} .

Results and discussion

UV-visible spectroscopy analysis for biofabrication of Pt-NPs

The biofabrication of colloidal Pt-NPs was confirmed by UV-visible spectroscopy analysis. The $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ solution (pale yellow) showed an absorption peak at around 260 nm in its UV-visible spectrum due to the ligand-to-metal charge-transfer transition between Pt^{4+} and Cl^- ions [26], displayed in Fig. 1. As the bio-reduction reaction was carried out, colloidal Pt-NPs were formed simultaneously with a change in pale yellow color of $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ solution into light brown. The absorption peak present at 260 nm disappeared and was replaced by a broad continuous absorption spectrum, which gradually increases in intensity from visible to the ultraviolet region, suggested Pt^{4+} ions were completely reduced to Pt^0 [27]. Control reaction mixture recovered as pale yellow in color, with no light-brown, being observed suggested Pt-NPs were formed only in presence of *P. granatum* peel extract.

DLS and ζ potential analysis

Fig. 2a revealed that the Z-average diameter of the biofabricated Pt-NPs was 30 nm with a polydispersity index (PDI) 0.270. The corresponding average ζ potential value of -15.7 mV confirmed the stability of Pt-NPs in colloidal solution (Fig. 2b). This low value of ζ potential of biofabricated nanoparticles was attributed to the additional influence by the electric charge of bio-organics present in peel extract. Various process parameters such as biomaterial dosage, temperature, and pH (change the electric charge of bio-organics), which might further affect their capping and stabilizing

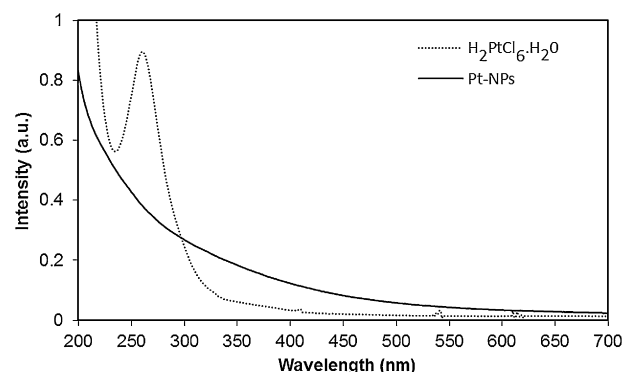


Fig. 1. UV-visible spectra of aqueous $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ and colloidal Pt-NPs.

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