FISHVIER

Contents lists available at ScienceDirect

Aquaculture

journal homepage: www.elsevier.com/locate/aqua-online



Identification of single nucleotide polymorphism cytochrome P450-c19a and its relation to reproductive traits in Japanese flounder (*Paralichthys olivaceus*)

Feng He, Hai S. Wen*, Shuang L. Dong, Bao Shi, Cai F. Chen, Lian S. Wang, Jun Yao, Xing J. Mu, Yu G. Zhou

Fisheries College, Ocean University of China, 5 Yushan Road, Qingdao 266003, China

ARTICLE INFO

Article history:
Received 1 August 2007
Received in revised form 22 March 2008
Accepted 24 March 2008

Keywords: Japanese flounder CYP19a Diplotype SNPs Reproductive traits

ABSTRACT

CYP19 is considered as an important factor affecting reproductive endocrinology in many fishes, and plays an important role in ovarian development, reproductive function and sexual differentiation. In this study, three single nucleotide polymorphisms (SNPs) within CDS of the CYP19a gene were tested and the associations between their genotypes and four reproductive traits were analyzed in 65 Japanese flounder individuals with Polymerase chain reaction and Single-stranded conformational polymorphism (PCR–SSCP). Results indicated that a SNP in the exon7 of CYP19a gene, SNP2, was significantly associated with 17 β -estradiol (E_2) (P<0.05) and gonadosomatic index (GSI) (P<0.05). Individuals with genotype AB of SNP2 had significantly higher serum E_2 levels (P<0.05) and GSI (P<0.05) than those of genotype AA or BB. In addition, there was significant association between one diplotype based on three SNPs and reproductive trait. The genetic effects for both serum E_2 of diplotype D9 and GSI of diplotype D1 were respectively much higher than those of other diplotypes (P<0.05). The evidence of the associations between genetic variants with serum E_2 and GSI may help explain effects of C1919a gene in reproductive endocrinology of Japanese flounder.

© 2008 Elsevier B.V. All rights reserved.

1. Introduction

Cytochrome P450arom, as a member of the cytochrome P450 superfamily, is a key component of the enzymatic aromatase complex converting androgens to estrogens in vertebrates. The protein that catalyzes the aromatization of steroid hormones is encoded by the CYP19 gene (Thompson and Siiteri, 1974; Simpson et al., 1994). Estrogens, especially 17β -estradiol (E₂), have been shown to play a key role in ovarian development, reproductive function and sexual differentiation in various species (Miyashita et al., 2000; Miyata and Kubo, 2000; Kuntz et al., 2003a; Kato et al., 2004). There are two isoforms of CYP19 genes including CYP19a and CYP19b present in Japanese flounder (Paralichthys olivaceus). They are primarily expressed in the ovary and brain, respectively (Kitano et al., 1999). Both aromatase CYP19 isoforms are involved in the sexual differentiation, regulation of the reproductive cycle and male reproductive behavior in Japanese flounder (Kitano et al., 1999, 2000). The gene mutation or disruption of either activity or production of this enzyme is likely to result in altered development or reproductive biology of organisms. Due to its key function in estrogen biosynthesis and association with reproductive processes, aromatase has been considered as an important factor to affect reproductive endocrinology in many fishes (Sanderson et al., 2002; Hayes et al., 2002; Rotchell and Ostrander, 2003).

Single nucleotide polymorphisms (SNP), one base variant including deletion, insertion, and substitution, can greatly influence gene expression and the functions of proteins. In agricultural and aquaculture species, SNPs are especially important if they cause differences in economic traits, or are linked to the mutations that do so. For example, in the centromeric region of Bos taurus autosome (BTA) 14, the acyl-CoA: diacylglycerol acyltransferasel gene (DGAT1) has been identified as the most likely causative gene underlying a OTL for milk fat yield and content (Grisart et al., 2002; Winter et al., 2002; Thaller et al., 2003). This information can be used to increase the accuracy of selection for these traits, thereby increasing the rate of genetic gain and production efficiency. While a large number of SNPs have been reported in some important livestock species (e.g. Kim et al., 2003; Jungerius et al., 2003), significant numbers of SNPs in the aquaculture species have been few reported by He et al. (2003), Heikki and Craig (2006) and Hayes et al. (2007). Those SNPs were only distributed in EST sequence. With regard to polymorphism of CYP19a gene, many studies focused mainly on diseases in different human populations (Kristensen and Borresen-Dale, 2000; Mitrunen and Hirvonen, 2003; Dunning et al., 1999; Ribeiro et al., 2006). While, polymorphisms in CYP19 gene of fish were firstly reported by Galay-Burgos et al. (2006).

Moreover, there is no report describing polymorphisms of CYP19a gene in Japanese flounder, and few studies about the relationship between mutants and reproductive traits. In this study, SNPs in Japanese flounder CYP19a and its effect on the synthesis of the E_2 or sex steroids were studied. Single-stranded conformational polymorphism (SSCP) analysis is one of the simplest, most reliable, and most sensitive methods for detecting mutations based on PCR (Orita

^{*} Corresponding author. Tel./fax: +86 532 82031825. E-mail address: wenhaishen@ouc.edu.cn (H.S. Wen).

et al., 1989; Sheffield et al., 1993). We have optimized the SSCP procedure to detect single nucleotide polymorphisms (SNPs) and used this method to evaluate polymorphisms of *CYP19a* and their associations with reproductive traits.

2. Materials and methods

2.1. Animals

Japanese flounder were reared in sea water at room temperature. Fish were decapitated and the gonads were removed and weighed. Gonads were fixed in 10% neutral buffered formalin, dehydrated, embedded in paraffin, sectioned at 5 μ m and stained with hematoxylin and eosin for histological examination. Animal populations were 65 female individuals of Japanese flounders, individual weight approximately 239.23±74.93 (g). Four reproductive traits, serum testosterone (T), serum 17 β -estradiol (E₂), Hepatosomatic index (HSI) and gonadosomatic index (GSI), were used for association analysis. Table 1 presented the mean and standard deviations of four traits.

2.2. Hepatosomatic index (HSI) and gonadosomatic index (GSI)

The Hepatosomatic index or gonadosomatic index of each animal was calculated as the ratio of the gonad or liver wet weight to the whole body net weight. Gonadosomatic or Hepatosomatic index= (Gonad or liver weight/(body weight-viscera weight)) × 100.

2.3. T and E_2 assays by radioimmunoassay

The blood was sampled by puncturing the caudal vasculature with a 25-guage 1.3-cm needle attached to a 1.0-ml disposable syringe. Blood samples were allowed to clot on ice for several hours, and then separated the serum by centrifugation (15,000 rpm) for 5 to 7 min and stored at -40 °C. The serum testosterone and estradiol-17 β were quantified by ¹²⁵I radioimmunoassay basing on double antibody assay, using diagnostic kits from Diagnostic Products Corporation (Tianjin Nine Tripods Medical & Bioengineering Co., Ltd., Sino-US jointventure enterprise). Steroids were assayed directly on the serum, the antisera are highly specific with an extremely low crossreactivity to other naturally occurring steroids, the crossreactivity was less than 0.1% to most circulating steroids. Intraassay variability was 7.4% for the estradiol-17\beta assay and 8.0\% for the testosterone assay. Any sample with coefficient of variation higher than 10% was not included in the analyses. The assay sensitivity reached to 2 ng/dl for T and 4 pg/ml for E₂ in a modified protocol provided by Wen et al. (2006).

2.4. PCR-SSCP analysis

Genomic DNA was isolated from blood sample by the phenol-chloroform method. Nine pairs of primers were designed to amplify eight exons of Japanese flounder *CYP9a* based on its cDNA sequence (GenBank Accession No. AB017182) using the Oligo6.0 software (Table 2). PCR reactions were carried out in a total of 25 µl volume containing 50 ng of genomic DNA, 0.20 mM each dNTP, 2.5 mM MgCl₂, 0.20 mM primers and 0.5 U Taq DNA polymerase. Amplification condition was 94 °C for 5 min followed by 35 cycles of 94 °C for 45 s, 57 °C for 45 s, 72 °C for 45 s and a final extension at 72 °C for 10 min. The PCR

Table 1Means and standard deviations of reproductive traits

Traits	Mean	SD ¹
T (pg/ml)	17.638	7.36
E_2 (pg/ml)	6.306	3.739
HIS	1.548	0.567
GSI	0.155	0.122

¹ Standard deviation.

Table 2Primer sequences and information of Japanese flounder *CYP19a* gene

Names	Sequences	Length (bp)	Tm (°C)	Amplicons
Primer1	5-GTCGTCCAGTTTGTGCAG-3	240	57	5'-UTR-exon1
	5-TCTCCTGTCTGTGTGGCT-3			
Primer2	5-GTCCACCTTTCTGTTTGG-3	150	57	Exon2
	5-TGCTGAGGATGAGTGTCT-3			
Primer3	5-CATGTACTGAAGAATGGA-3	139	49	Exon3
	5-CTTTGGAGAAATAGTTGC-3			
Primer4	5-TCTGACAGGTCCAGGTTTG-3	159	60	Exon4
	5-GGGCACATCAAGGAAGAGT-3			
Primer5	5-GAGCTGCTGCTGAAGATT-3	108	56	Exon5
	5-TGCTGTCTTATGCCTCTG -3			
Primer6	5-CTGTCTGCGGAGAACGTGGT-3	145	57	Exon7
	5-CAGTGTCAATCTCTCGCAGC-3			
Primer7	5-CTGGAGAGCTTCATCAAC-3	193	57	Exon8
	5-TCTCAAAGTTGTCCAGGC-3			
Primer8	5-GACGTTACTTCCAGCCAT-3	250	55	Exon9
	5-TCAGAGTGTTTGCCAGCT-3			
Primer9	5-TGATCCACACTGCTTCAT-3	237	53	3'-UTR
	5-TTCCTACTTGGAAAGTGC-3			

products of *CYP19a* were genotyped by single-stranded conformation polymorphism (SSCP) method. Two μl PCR products of each individual were mixed with 5 μl denaturing buffer (98% formamide, 0.09% xylene cyanole FF, and 0.09% bromophenol blue), and then denatured at 94 °C for 5 min followed by a rapid chill on ice for 10 min. The denatured PCR products were separated on 12% polyacrylamide gel for 14 h at 4 V/cm. The DNA bands were stained by silver staining (Qu et al., 2005). Individual genotypes were defined according to band patterns.

PCR products of each type of homozygotes were purified with DNA Fragment Quick Purification/Recover Kit. The purified PCR products were ligated to the PMD 18-T vector and transformed into DH5- α Escherichia coli. Positive recombinant colonies were sequenced on the ABI 377 sequencer.

2.5. Statistical models and analysis

The genotype frequencies of each polymorphism were calculated by Excel. The diplotypes were constructed on the base of 3 SNPs with phase 2.0. Associations between genotypes and diplotypes of 3 SNPs of Japanese flounder CYP9a gene and four reproductive traits (T, E₂, HSI and GSI) and genetic effects were respectively analyzed using GLM procedure of SAS 8.02 software. The following models were used.

$$Y = \mu + G (or H) + e$$

where Y is value measured of four reproductive traits; μ is mean value of four reproductive traits, G or H is fixed effects of genotypes of each SNP or diplotype, e is random error effect. Considering the all experimental fish were female from the same site and slaughtered at the same age, so other effects were not taken in this model such as sex, generation and site. Significant differences among least-square means of different genotypes or diplotypes were calculated using Duncan's multiple-range test, and P values of 0.05 were considered statistically significant.

3. Results

3.1. Polymorphisms within exons of CYP19a gene

Among the nine sets of primers used to amplify the gene fragments by PCR–SSCP analysis, the PCR products of primer1, primer6, and primer7 were polymorphic, respectively (Fig. 1). Three SNPs, namely SNP1, SNP2 and SNP3, were located at positions of A193C, T993G and A1297G of Japanese flounder *CYP19a* gene (Fig. 2). Three genotypes were found for each SNP and named as AA, AB, and BB, respectively (Fig. 1).

Download English Version:

https://daneshyari.com/en/article/2424725

Download Persian Version:

https://daneshyari.com/article/2424725

<u>Daneshyari.com</u>