Short communication: Dietary conjugated linoleic acid down-regulates fatty acid transporters in the mammary glands of lactating rats

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ABSTRACT

Recent studies indicated that reduction of milk triacylglycerol concentrations by dietary conjugated linoleic acid (CLA) involves an impairment of both de novo fatty acid synthesis and uptake of fatty acids from circulating triacylglycerol-rich lipoproteins into the mammary gland. However, nonesterified fatty acids (NEFA) in the plasma released from adipose tissue and taken up into the mammary gland by fatty acid transporters are a further important source of fatty acids available for milk triacylglycerol synthesis. Therefore, the aim of the present study was to investigate the effect of dietary CLA on plasma concentrations of NEFA and the expression of fatty acid transporters in the mammary glands of lactating rats fed either a CLA diet or a control diet. Dams fed diets with CLA had a greater concentration of NEFA in plasma than those fed the control diet. In addition, relative mRNA concentrations of fatty acid transporters (fatty acid translocase/CD36, fatty acid transport protein, and plasma membrane fatty acid binding protein) were about 45, 75, and 70% lower, respectively, in the mammary gland of dams fed diets with CLA compared with those fed the control diet. In conclusion, the present findings indicate that reduced uptake of circulating NEFA released from white adipose tissue into the mammary gland could also contribute to the reduction of milk triacylglycerol concentrations by dietary CLA in rats. The mechanism through which CLA inhibits expression of fatty acid transporters deserves further study.

Key words: conjugated linoleic acid, fatty acid transporters, lactation, mammary gland

Conjugated linoleic acid (**CLA**) is a collective term for a group of positional and geometric isomers of linoleic acid with conjugated double bonds. Conjugated linoleic acid is naturally found in significant amounts in milk, dairy products, and meat of ruminants (Steinhart

et al., 2003). Several studies showed that dietary CLA exerts many biological effects in humans and animals (Martin et al., 2000; Baumgard et al., 2001; Masters et al., 2002; Terpstra et al., 2002; Toomey et al., 2006). For instance, in lactating rats (Ringseis et al., 2004; Hayashi et al., 2007), sheep (Lock et al., 2006), cows (Baumgard et al., 2001, 2002; Peterson et al., 2004; Harvatine and Bauman, 2006), and humans (Masters et al., 2002) dietary CLA causes a reduction in milk triacylglycerol concentration.

Milk triacylglycerol synthesis depends on the availability of fatty acids in the mammary gland, which are derived from 3 different sources. The first source represents de novo biosynthesis of fatty acids within the mammary gland by the activity of lipogenic enzymes. Medium-chain fatty acids with 8 to 14 carbon atoms are the main products of this process, which is controlled by the lipogenic transcription factor sterol regulatory element-binding protein (SREBP)-1c (Barber et al., 2003). Fatty acids released from triacylglycerol-rich lipoproteins by lipoprotein lipase and taken up into the mammary gland by fatty acid transporters are a second important source for milk triacylglycerol synthesis (Scow et al., 1977). Nonesterified fatty acids in the plasma released from adipose tissue by hormone-sensitive lipase and taken up into the mammary gland by fatty acid transporters are a third source of fatty acids available for milk triacylglycerol synthesis. Fatty acids with 16 carbon atoms and long-chain fatty acids with 18 to 22 carbon atoms, either saturated or unsaturated, largely reflect the second and third sources of fatty acids for milk triacylglycerol synthesis (Green et al., 1981; Ross et al., 1985). The contribution of NEFA to milk fat synthesis in lactating cows is especially important during the early lactation stage, because the capacity for feed intake is limited at the beginning of lactation and therefore adipose tissue depots, which have accumulated during pregnancy, are actively mobilized during this stage. In mid and late lactation NEFA are of minor importance for milk fat synthesis in dairy cows because feed intake is sufficient to provide enough substrates for milk fat synthesis and even to replenish lipid stores during this lactation stage. In contrast, NEFA

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are an important source for milk fat synthesis during the whole lactation in species with a high milk yield such as the rat, whose demand for lactation is so large that food intake is dramatically increased (up to 3- to 4-fold compared with nonlactating rats; Peterson and Baumgardt, 1971). In spite of the large increase in feed consumption, however, rats are generally in negative energy balance during lactation, in particular during peak lactation (d 12 to 14 postpartum), where mobilization of body fat and protein is greatest (Sampson and Janson, 1984). Consequently, rats usually lose weight during lactation regardless of the feeding regimen (e.g., ad libitum) or the energy content of the diet (Sainz et al., 1986).

Several studies have shown that the reduced milk triacylglycerol concentrations by dietary CLA were accompanied by a diminished activation of SREBP-1c and reduced expression and activity of lipogenic enzymes and lipoprotein lipase in the mammary gland (Baumgard et al., 2002; Peterson et al., 2004; Ringseis et al., 2004; Harvatine and Bauman, 2006). These findings clearly indicate that dietary CLA reduces milk triacylglycerol concentrations by impairment of both de novo fatty acid synthesis and uptake of fatty acids from circulating triacylglycerol-rich lipoproteins into the mammary gland. However, whether the reduced concentrations of long-chain fatty acids and triacylglycerols in the milk by dietary CLA are also the consequence of diminished uptake of NEFA by fatty acid transporters from plasma into the lactating mammary gland has not been investigated yet. Therefore, the aim of the present study was to investigate the effect of dietary CLA on plasma concentrations of NEFA and the expression of the most important fatty acid transporters, fatty acid translocase/CD36 (FAT/CD36), fatty acid transport protein (FATP), and plasma membrane fatty acid binding protein (**FABPpm**), in the mammary gland of lactating rats. For this purpose, we used samples of a recently performed feeding experiment with lactating rats that were fed either a CLA diet or a control diet containing sunflower oil (SFO; Ringseis et al., 2004). In this experiment, the rats of the CLA group had a 46% lower milk fat content and markedly lower absolute concentrations of medium-chain fatty acids, fatty acids with 16 carbon atoms, and long-chain fatty acids in the milk at d 10 postpartum [C8 to C14: CLA group: $56 \pm$ 13 mmol/L, n = 8, SFO group: 149 ± 6 mmol/L, n = 10, means \pm SEM, P < 0.05; C16: CLA group: 47 ± 9 mmol/L, n = 8, SFO group: 85 ± 4 mmol/L, n = 10, means \pm SEM, P < 0.05; C18 to C22: CLA group: 99 \pm 17 mmol/L, n = 8, SFO group: 140 \pm 8 mmol/L, n = 10, means \pm SEM, P < 0.05, (Ringseis et al., 2004)]. Relative proportions of medium-chain fatty acids in the milk were also lower in the CLA group than in the SFO

group (CLA group: $17.6 \pm 1.6 \text{ g}/100 \text{ g}$ of fatty acids, n = 8, SFO group: 28.5 ± 2.2 g/100 g of fatty acids, n = 10, means \pm SEM, P < 0.05), whereas proportions of C16:0 and long-chain fatty acids did not significantly differ between the groups (C16:0: CLA group: 23.6 \pm 0.5 g/100 g of fatty acids, n = 8, SFO group: 22.8 ± 0.8 g/100 g of fatty acids, n = 10, means \pm SEM; C18 to C22: CLA group: 49.0 ± 3.2 g/100 g of fatty acids, n = 8, SFO group: 41.2 ± 1.5 g/100 g of fatty acids, n = 10, means \pm SEM). The rats, at a mean BW of 64 \pm 1 g (mean \pm SEM), were randomly assigned to 2 groups (n = 12) and fed the diets with 30 g/kg diet of either sunflower oil (SFO group) or CLA oil (CLA group). The fatty acid compositions of total lipids of SFO and the CLA oil were similar except for the concentrations of 18:2 n-6 and CLA. The CLA oil contained 54 g of CLA isomers/100 g of CLA oil, whereas the CLA concentration in the SFO was <0.1 g per 100 g of total fatty acids. The CLA isomer distribution of the CLA oil was as follows (g/100 g of total CLA): trans(t)-10 cis (c)-12 (18.5), c11t13 (15.8), c9t11 (15.6), t8c10 (14.9), t10t12 (5.61), t9t11 (5.41), t7t9 (3.12), c13t15and t13c15 (2.85), t11t13 (2.75), t8t10 (2.63), c10c12(2.58), t12t14 (2.22), c9c11 (2.08), c11c13 (1.99), c8c9(1.30), c12t13 and t12c13 (0.97), t7c9 (0.79), t11c13(0.62), t6t8 (0.18), and t13t15 (0.13). At 11 wk of age, the rats were paired with adult male Sprague-Dawley rats for 6 d. At the day of parturition, designated as d 1 of lactation, litters were weighed and then adjusted to 10 pups per dam without differentiation of sex. The experimental diets were fed for a total of 13 wk, starting at 5 wk of age. During growth and pregnancy, the rats were fed identical amounts of the experimental diets, increasing from 7 to 19 g/d, except for wk 11. In wk 11, when the rats were paired with the male rats, they had free access to the experimental diets. Throughout the period of lactation, the rats also had free access to the experimental diets; however, daily food intake during lactation did not differ between groups [CLA group: $33.7 \pm 1.4 \text{ g/d}, \text{ n} = 8$; SFO group: $34.9 \pm 0.9 \text{ g/d}, \text{ n} =$ 10, means \pm SEM (Ringseis et al., 2004)]. Based on an energy content of the diets of 17.5 MJ/kg of diet, the daily food intake during lactation was $590 \pm 25 \text{ kJ/d}$ in the CLA group (n = 8) and 611 \pm 16 kJ/d in the SFO group (n = 10), means \pm SEM. Plasma and samples of mammary gland were obtained on d 17 of lactation, when the dams were anesthetized with diethyl ether and killed by decapitation. Day 17 of lactation in rats corresponds to the late lactation stage where milk yield is slowly declining because pups suckle less milk. During these final days of lactation, the energy and nutrient demands of the pups are partially met by the consumption of solid food. Further details regarding animals, diets, feeding regimen, and sample collec-

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