



## Short communication

Occurrence of  $\epsilon$ -proteobacterial species in rabbits (*Oryctolagus cuniculus*) reared in intensive and rural farmsJ. Revez<sup>a,b,\*</sup>, M. Rossi<sup>b</sup>, S. Piva<sup>a</sup>, D. Florio<sup>a</sup>, A. Lucchi<sup>c</sup>, A. Parisi<sup>d</sup>, G. Manfreda<sup>c</sup>, R.G. Zanoni<sup>a</sup><sup>a</sup> Department of Veterinary Medical Sciences, Alma Mater Studiorum, University of Bologna, Via Tolara di Sopra 50, 40064 Ozzano dell'Emilia, Bologna, Italy<sup>b</sup> Department of Food Hygiene and Environmental Health, Faculty of Veterinary Medicine, University of Helsinki, P.O. Box 66, FI-00014 Helsinki, Finland<sup>c</sup> Department of Food Science, Alma Mater Studiorum – University of Bologna, Via Del Florio 2, 40064 Ozzano Emilia, Bologna, Italy<sup>d</sup> Experimental Zooprophyllactic Institute of Apulia and Basilicata, Via Chiancolla 1, 70017 Putignano (BA), Italy

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## ABSTRACT

In order to investigate the occurrence of *Campylobacter*, *Helicobacter* and *Arcobacter* species in caecal contents of rabbits reared in intensive and rural farms, a total of 87 samples from animals belonging to 29 farms were analysed by both cultural and PCR analyses.

PCR analysis directly from faecal samples detected 100% positive samples for *Campylobacter* genus, 3.4% for *Helicobacter* genus and none for *Arcobacter* genus. 83 out of 87 animals (95.4%) and all the 29 farms were positive for *Campylobacter cuniculorum* as also determined by cultural examination. *Campylobacter coli* and *Campylobacter jejuni* were isolated only from three animals reared in two rural farms. No *Helicobacter* and *Arcobacter* species were isolated. To evaluate a possible genetic variability, one strain of *C. cuniculorum* from each farm was analysed by Pulsed Field Gel Electrophoresis (PFGE) and Amplified Fragment Length Polymorphism (AFLP). Genotyping revealed that *C. cuniculorum* population is heterogeneous among the different sources and no dominant clone has spread in the investigated farms.

This survey highlighted a high presence of *C. cuniculorum* with a high rate of intestinal colonization, low presence of *C. jejuni*-*coli*, *Helicobacter* spp. and any *Arcobacter* spp. in farmed rabbits.

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## 1. Introduction

*Campylobacter* and *Helicobacter* species are frequently found in the gastrointestinal tracts of mammals and birds. *Campylobacter jejuni* and *Campylobacter coli* are worldwide known as major food-borne enteropathogens causing enteric diseases in humans (Engberg et al., 2000) and *Helicobacter* species have been related with gastritis, gastric ulceration, enteric disease and bacteraemia in humans (De Groote et al., 2000; Moyaert

et al., 2008). Recently, *Arcobacter* species have been recognized as emerging foodborne pathogens and also detected in livestock animals (Lehner et al., 2005). Although the rabbit (*Oryctolagus cuniculus*) is considered an important source of meat for humans (Kohler et al., 2008) and is markedly gaining in importance as pet animal, microbiological data on the occurrence of  $\epsilon$ -proteobacteria in rabbits is limited.

Gastric *Helicobacter* have been detected in the stomach of rabbits (Van den Bulck et al., 2005) and only one study reports the isolation of *Arcobacter butzleri* from rabbit meat (Collado et al., 2009). In addition, few studies have reported low prevalence of *Campylobacter* species (mainly *C. jejuni*), in healthy and diarrheic rabbits as well as from rabbit meat (Prescott and Bruin-Mosch, 1981; Kohler et al., 2008; Little et al., 2008; Piccirillo et al., 2011). Recently a

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new *Campylobacter* species, *C. cunicolorum*, was described in rabbit caecal contents (Zanoni et al., 2009), but the occurrence of this *Campylobacter* in rabbits is nowadays unknown. The aim of this study was to evaluate the occurrence of different species of  $\epsilon$ -proteobacteria in rabbits caecal content by both cultural and PCR analyses.

2. Materials and methods

2.1. Sampling

29 farms of rabbits (18 intensive and 11 rural) were sampled from April 2007 to November 2008. Twenty-seven farms (18 intensive and 9 rural) were located in seven different Italian regions, while 2 rural farms were located in Portugal. Data relative to the farms is reported in Fig. 1. Among 87 rabbits (3 animals per farm), 57 were healthy and were sampled at the slaughterhouse (intensive farms) or during private slaughter (rural farms), while 30 animals, with enteritis, were sampled during routine necropsy. The complete intestinal tract from each rabbit

was removed avoiding cross-contamination, collected into a separate sterile plastic bag using fresh disposable gloves, kept at  $5 \pm 3^\circ\text{C}$  and examined within 4 h after sampling.

2.2. Cultural examination

Approximately 5 g of caecal content from each gut was diluted into 5 mL of sterile saline and homogenized by vortex mixer. Ten microlitres of the diluted samples were streaked onto plates with the following media: *Campylobacter* blood-free selective agar (mCCDA, Oxoid), Blaser-Wang's Agar (Oxoid), Skirrow's Agar (Oxoid), and C.A.T. Selective Medium (Oxoid). The last three media were prepared using Nutrient Broth N° 2 (Oxoid) with 1.5% of Bacto Agar (Difco) as base media. In addition a modified filter technique of Steele and McDermott was performed as described previously (Zanoni et al., 2007). All plates were incubated at  $37 \pm 1^\circ\text{C}$  under a microaerobic atmosphere with hydrogen (Bolton et al., 1992) and examined daily up to 12 days. From each plate different types of colonies of gram

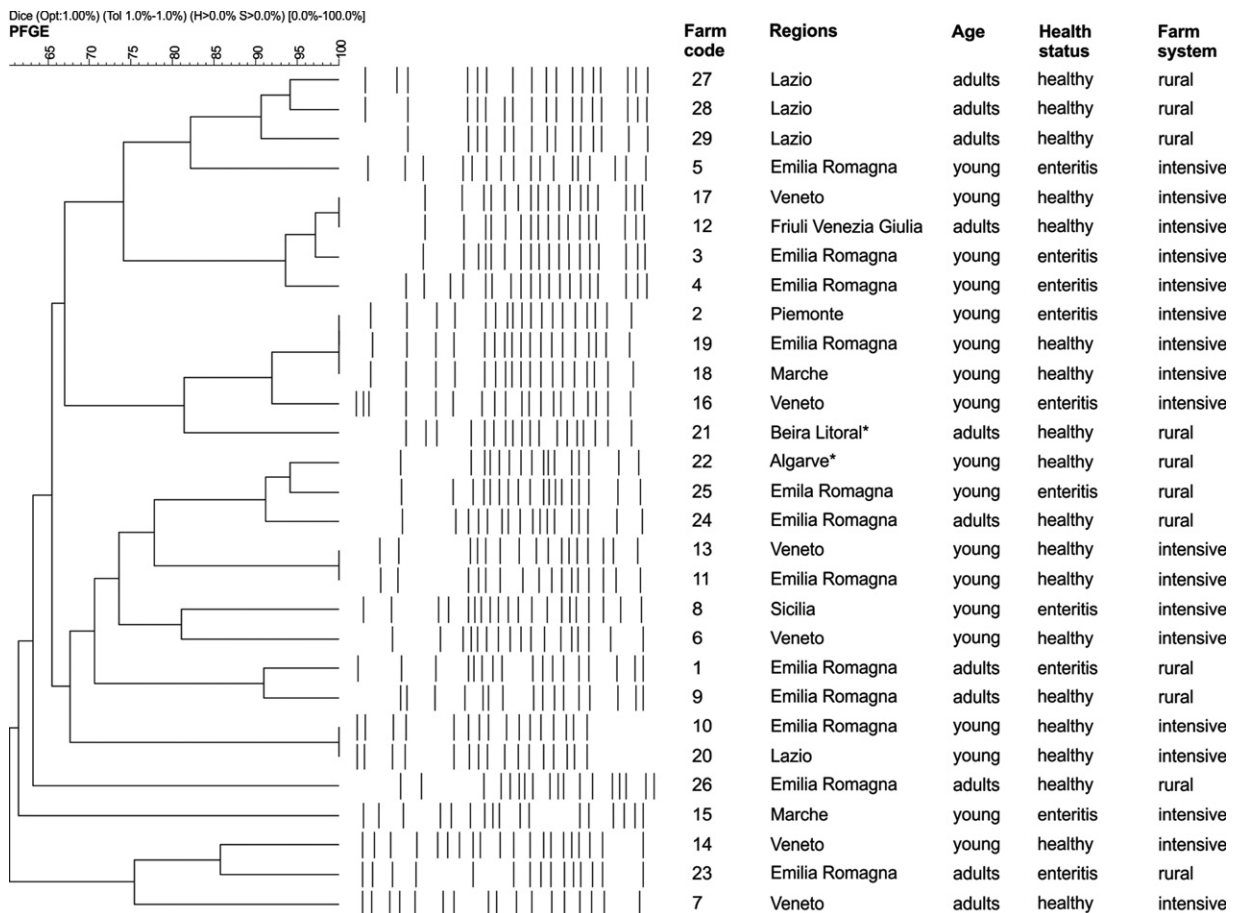


Fig. 1. Distribution of *HhoI* digested PFGE patterns of *Campylobacter cunicolorum* (1–29). Information of the origin of the strains (all from Italian regions except those indicated with \*, which are from Portuguese regions), age (adults > 6 months; young < 6 months), health status (considered only if animals had or not enteritis) and farm system is presented on the side of the dendrogram. The number of rabbit does in the intensive farms ranged from 300 to 700 subjects while in rural from 5 to 15. The numbers on the horizontal axis indicate the percentage of similarities as determined by Dice correlation coefficient and UPGMA clustering.

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