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Recombination as a motor of host switches and virus emergence: geminiviruses as case studies

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Genetic recombination facilitates the transfer of genetic information in a parasexual reproduction manner even between distantly related species. Within the *Geminiviridae* family, a group of plant-infecting viruses that severely constrain cropping systems worldwide, it is highly suspected that recombination was pivotal in the emergence as a devastating phytopathological problem. Whereas extensive evidence of recombination suggests that this mechanism might be adaptive in this family, direct demonstration remains scarce. Here we assemble lines of evidences indicating that recombination was crucial in driving host switches and further emergence of geminiviruses, making these viruses such successful plant pathogens.

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Introduction

Genetic recombination allows parental viruses to derive genetic information to their progeny in a parasexual reproduction manner. This mechanism is a key process in the evolution of many virus families and has been extensively recorded for members of the family *Geminiviridae* [1–4]. Geminiviruses are plant-infecting circular single stranded (ss) DNA viruses that severely constrain production in a variety of cropping systems throughout the world [5–7]. They comprise seven genera that differ in terms of phylogenetic relationships, genome structure, host range and insect vector [8]. Besides evolving at substitutions rates equivalent to those found for RNA viruses [9], in this virus family, extensive evidence of recombination is available at every level of the diversity spectrum [10]. As recorded also for other virus groups [11], this mechanism was most probably essential in the macroevolution and the emergence process (see $[1,12^{\circ},13]$ for a review of recombination for the ssDNA viruses in general).

Recombination in geminiviruses facilitates the transfer of genetic information even between distantly related species in such extent that their genomic organization is thought to have evolved to maximize its adaptive value and minimize potentially deleterious effects [1]. In fact, the risk to produce defective progeny is high for geminiviruses because of their highly compacted genome $(\sim 2.7 \text{ kb})$ with overlapping genes, multifunctional proteins and optimized interactions between different parts and proteins it encodes. While the preservation of coevolved interactions is supported by wide experimental and analytical information, in some cases recombinants with novel abilities to interact with hosts or vector might arise. Thus, for example, the ability to infect a particular host depends in part on the balance between host defenses and virus counterdefense. Among host defenses, it is now apparent that gene silencing is an essential plant antiviral mechanism [14]. However, as for other plant viruses, geminiviruses encode proteins that can suppress gene silencing [15] in a host dependent manner [16[•]]. Genetic exchanges involving these proteins might drive host shift and emergence [16[•],17].

The abilities of geminiviruses to rapidly generate polymorphisms through mutations [9] and combine new genetic forms through recombination were probably essential in responding to the numerous niche extensions events offered within the context of modern agriculture [1,5-7,18-21]. However, despite a large amount of indirect evidence and some degree of speculation, there are few well supported examples of this having occurred in nature (see $[4,22,23,24^{\circ}]$ for good examples).

Recombination patterns in geminiviruses

Whereas non-homologous recombination (during which genome regions get rearranged, duplicated, deleted) is mostly apparent above the genus level in the *Geminiviridae* family, homologous recombination is a widespread phenomenon as almost every geminivirus is the descendent of some inter-species or inter-strain recombinant [10]. Importantly, analyses of viral sequences obtained from both the environment and experimental studies revealed that recombination breakpoints are generally not randomly distributed with conserved recombination hot spots and cold spots [10]. Studies demonstrated that these distribution patterns are strongly influenced by mechanistical factors preserving interaction networks [1,25,26,27^{••}]. Remarkably, recombination is intimately associated with the replication process of geminiviruses. It notably involves the so-called 'recombination dependent replication' mechanism that is able to recover fragments of geminiviral DNA that may result from incomplete synthesis or from nucleolytic attack to create recombinant viruses [28,29]. Doing so, it has the potential to create large number of recombinant viruses displaying specific recombination patterns (e.g. the notorious recombination hot-spot around the origin of replication [10]) when multiple viruses co-infect the same cell [1].

While mechanistic predisposition profoundly affects the recombination patterns, these also bear severe imprints of selection. Recombination breakpoint distribution patterns are partially attributable to natural selection disfavouring survival of recombinants in which co-evolved intra-genome interaction networks are disrupted [25,30]. Whereas the vast majority of recombinant forms, as most mutants, are likely of poorer fitness than the parents [26,31], recombinants bearing favourable combinations [27] might outcompete the parental viruses and become prevalent in the population.

It is clearly apparent from the literature that genetic material is exchanged between viruses participating into epidemics on distinct hosts. Although it does not prove that recombination allowed host shift or range extension, it clearly demonstrated exchanges of genetic material between distinct viral diversity pools.

Additionally in the *Begomovirus* genus of the *Geminiviridae* family, where bipartite genomes occur, mixed infections provide opportunities for heterologous genome component reassortment, also-called pseudo-recombination. Evidences of exchange, loose or gain of components during the evolution of begomoviruses is shown in natural infections [32–34]. Importantly, pseudorecombination involving non-essential components such as betasatellites, can modulate the expression of the disease in a host-dependent manner, and drive host shift [35–37] providing another determinant for host adaptation.

Recombination as a driving force of emergence

Besides experimental studies demonstrating alteration of ecology and fitness following recombination between geminiviruses [22,38–45], evidences accumulate for recombination as *in natura* mechanism of modification of host ranges [4,22,32,40,46–48,49^{••}]. These notably include the invasive spread of tomato yellow leaf curl disease (TYLCD)-associated begomoviruses across the Western Mediterranean during the past three decades [22,50] or the emergence of maize streak disease as a major agricultural threat throughout the African continent [4,51]. It should be highlighted that these diseases were absent in the centre of origin of the host plants (both from the Americas) and have emerged in the areas where these plants where later introduced. Recombination might have contributed to the likely host switches events and adaptation of locally circulating geminiviruses.

Recombination as a driving force in dicot geminiviruses: begomoviruses

Begomovirus is one of the largest plant virus genus comprising almost 300 viral species [52]. Several lines of evidence indicate that the emergence of a highly polyphagous and invasive type of the vector of begomoviruses, the whitefly (Hemiptera: Aleyrodidae) Bemisia tabaci, is contributing to transferring begomoviruses among a large set of distinct hosts [7,53], increasing the opportunities of mixed infection and further diversification through recombination [21]. Some instances of such events have been recorded with, for example, the recent pandemic on cassava in Africa caused by a recombinant virus with an extremely increased severity [54,45], or the emergence of the resistance-breaking recombinant virus named cotton leaf curl Burewala virus (CLCuBuV) that causes devastating damage to cotton production in India and Pakistan [24]. One of the best studied examples is the one that refers to TYLCD-associated begomoviruses. The TYLCD has emerged during the last decades in almost every region of the world where tomato is commercially grown [50] and is caused by a complex of begomovirus species related to the highly invasive tomato yellow leaf curl virus (TYLCV) [55]. Complex recombinationderived progenies were shown to arise at high frequencies and in relatively short periods during mixed infections of TYLCD-related species [3,56] or with other distantly related begomoviruses [30,57]. Interestingly, although natural selection preserves co-evolved intra-genome interaction networks [30], numerous and diverse viable recombinants can be produced during mixed infections [30,57[•]] or artificially [58], highlighting the tolerance to disruption of begomoviruses genomes. Scarce information, however, is available on the fitness of recombinants arising in natura. Again, studies conducted on natural populations of TYLCD-associated viruses inform about this aspect. In Spain and Italy, a diversity of recombinant viruses between TYLCD-associated begomoviruses was described [22,23,59]. Importantly, both field and experimental studies demonstrated that successful recombinant viruses exhibited host shift in respect to parental viruses, being able to infect a larger set of hosts [22,23]. This was the case, for example, of the recombinant virus named tomato yellow leaf curl Málaga virus (TYLCMalV) that has an enlarged host range and outcompeted the parental viruses in *Phaseolus vulgaris* epidemics [3,22].

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