Contents lists available at SciVerse ScienceDirect



Review



journal homepage: www.elsevier.com/locate/biochempharm

# Structural insights into Cys-loop receptor function and ligand recognition

## Mieke Nys\*, Divya Kesters, Chris Ulens

Laboratory of Structural Neurobiology, Department of Cellular and Molecular Medicine, KU Leuven, Herestraat 49, PB 601, B-3000 Leuven, Belgium

#### ARTICLE INFO

#### ABSTRACT

Article history: Received 7 May 2013 Received in revised form 3 July 2013 Accepted 3 July 2013 Available online 10 July 2013

Keywords: Pentameric ligand-gated ion channel Cys-loop receptor Ligand recognition Channel gating Ion permeation This review outlines recent insights into ligand recognition, channel gating and ion permeation for the family of pentameric ligand-gated ion channels (pLGICs). These receptors are involved in the fast inhibitory and excitatory neurotransmission. Prototypical anion-selective members are the  $\gamma$ -amino butyric acid type A (GABA<sub>A</sub>),  $\gamma$ -amino butyric acid type C (GABA<sub>C</sub>) and glycine receptor. The cation-selective members are the 5-HT<sub>3</sub> serotonin and nicotinic acetylcholine (nACh) receptors. They are the target for a wide variety of drugs and dysfunction in these receptors is associated with several diseases.

We summarize recent structural knowledge in combination with electrophysiological data and molecular dynamic simulations, thereby describing key features of ligand binding, channel gating and ion permeation. A conserved cation- $\pi$  interaction between ligand and aromatic residues of the ligand binding site critically contributes to ligand recognition, as revealed by X-ray crystal structures of acetylcholine binding proteins, as well as the integral pLGICs, ELIC and GluCl. In addition, we summarize the possible downstream effects on gating of structural rearrangements in the extracellular ligand-binding domain, which mainly occur in loop C and loop F. These data are discussed in the context of different conformational states of the pore-forming domain observed in crystal structures of GLIC and GluCl, which likely represent the open pore conformation, and ELIC, which likely corresponds to a closed pore conformation. We conclude with a current structural view on the determinants of ion selection and permeation.

© 2013 Elsevier Inc. All rights reserved.

CrossMark

#### Contents

1.	Introduction			1043
2.	Ligand recognition in eukaryotic Cys-loop receptors			1044
	2.1. The ligand binding pocket			1044
	2.2.	Molecular determinants for ligand recognition		1044
		2.2.1.	Conserved cation- $\pi$ interactions in AChBPs and pLGICs (Fig. 4)	1044
		2.2.2.	Water-mediated binding of partial agonists (Fig. 5)	1045
		2.2.3.	Loop D and E are important determinants for ligand efficacy	1046
	2.3.	Structural rearrangements upon ligand binding		1046
		2.3.1.	The conformational change of loop C: A contracted, open or intermediate conformation	1047
		2.3.2.	Loop F	1048
3.	Channel gating and ion permeation			1048
	3.1.	3.1. The overall architecture of the transmembrane domain		
	3.2.	3.2. Structural rearrangements during channel gating		
	3.3.	3.3. Ion conduction and selectivity		
4.	Conclusions and future challenges			1053
	References			

\* Corresponding author. Tel.: +32 16 330113. *E-mail address:* mieke.nys@med.kuleuven.be (M. Nys).

0006-2952/\$ - see front matter © 2013 Elsevier Inc. All rights reserved. http://dx.doi.org/10.1016/j.bcp.2013.07.001

#### 1. Introduction

Our expanding structural knowledge of membrane proteins, including ion channels, G-protein coupled receptors, transporters and pumps, progressively shifts the drug design process for these important drug targets from high-throughput screening toward a structure-based discovery approach. The treatment of neurological disorders could greatly benefit from rational drug development since many of these disorders could be better treated with more potent or efficacious drugs with fewer side effects. However, structure-based drug discovery approaches require a thorough insight into the structural features of target receptors, including the family of pentameric ligand-gated ion channels (pLGICs). This family consists of allosterically regulated membrane proteins located at synapses in the nervous system. They convert binding of a specific neurotransmitter released in the synaptic cleft into an ion flux over the postsynaptic membrane, which subsequently triggers excitatory or inhibitory postsynaptic potentials. Known vertebrate anion-selective pLGICs are the  $\gamma$ -amino butyric acid type A (GABA<sub>A</sub>),  $\gamma$ -amino butyric acid type C (GABA<sub>C</sub>) and glycine receptor, whereas the cation-selective pLGICs are the 5-HT<sub>3</sub> serotonin and nicotinic acetylcholine (nACh) receptors. pLGICs serve as targets for a wide variety of frequently prescribed drugs, including smoking cessation aids, anxiolytics, anticonvulsants, muscle relaxants, hypnotics and anti-emetics. Dysfunction of pLGICs also plays an important role in several disorders of the central nervous system, including hyperekplexia [1], myasthenia gravis [2], epilepsy [3], irritable bowel syndrome (IBS) [4], Alzheimer's disease [5], schizophrenia [6] and Parkinson's disease [7].

The current understandings concerning structure, gating mechanism and ligand recognition of pLGICs (Fig. 1) are derived from electron microscopic imaging of the nACh receptor from the *Torpedo marmorata* (*Tm*) electric organ [8] and X-ray crystallographic studies of the extracellular domain (ECD) from the muscle  $\alpha$ 1 nACh receptor [9] and the water-soluble acetylcholine binding proteins (AChBP) [10]. Until 2005 it was

assumed that this class of membrane proteins was uniquely expressed in multicellular eukaryotic organisms, but Tasneem et al. also identified several homologues in unicellular prokaryotic organisms (Fig. 2) [11]. In 2008 the first X-ray crystal structure of an integral prokaryotic pLGIC, derived from Erwinia chrysanthemi (ELIC) was determined [12]. This structure likely corresponds to a non-conducting conformation of the ion channel. About one year later a second crystal structure was determined for another prokarvotic homologue, derived from the cyanobacterium *Gloeobacter violaceus* (GLIC) [13,14], which likely displays an open ion-conducting conformation. Both ELIC and GLIC form cation-selective ion channels, similar to the nACh receptor (nAChR) and the 5-HT<sub>3</sub> receptor (5-HT<sub>3</sub>R). ELIC can be activated by primary amines such as GABA [15,16], GLIC on the other hand responds to an increase in extracellular proton concentration [17]. In 2011 the first 3-dimensional structure of a eukaryotic, anion-selective pLGIC was determined. This glutamate-gated chloride (GluCl) channel is derived from Caenorhabditis elegans and reveals an open pore conformation, similar to GLIC [18].

Based upon this structural information, the overall architecture of pLGICs appears to be conserved and consists of five homologous or identical membrane-spanning subunits. Each of these subunits is composed of a N-terminal extracellular ligand binding domain, four transmembrane domains, an intracellular loop between the 3rd and 4th transmembrane domain and an extracellular Cterminus. They are also referred to as the family of Cys-loop receptors (CLRs) due to a highly conserved disulfide bond in one of the loops forming the interface between the extracellular ligand binding domain and the pore-forming transmembrane channel domain. This disulfide bond is not present in the prokaryotic homologues, but the architectural fold of this loop is similar to their eukaryotic homologues.

This review will outline the current structural knowledge and understandings concerning the molecular recognition of ligands, channel gating and ion permeation for this class of ion channels.



**Fig. 1.** Overview of the current structural knowledge concerning pentameric ligand-gated ion channels. From left to right: The electron-microscopic structure of the *Torpedo marmorata* nAChR [8], followed by X-ray structures of the water-soluble AChBP [10], ELIC [12], GLIC [13,14] and GluCl [18]. Torpedo nAChR, *Torpedo marmorata* nicotinic acetylcholine receptor; AChBP, Acetylcholine binding protein; ELIC, *Erwinia chrysanthemi* ligand-gated ion channel; GLIC, *Gloeobacter violaceus* ligand-gated ion channel; GluCl, Glutamate-gated chloride channel derived from *Caenorhabditis elegans*.

Download English Version:

### https://daneshyari.com/en/article/2512540

Download Persian Version:

https://daneshyari.com/article/2512540

Daneshyari.com