

Short communication

Role of cannabinoid CB₂ receptors in glucose homeostasis in rats

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Received 19 August 2006; received in revised form 12 February 2007; accepted 13 February 2007

Available online 20 April 2007

Abstract

Here we show that the activation of cannabinoid CB₂ receptors improved glucose tolerance after a glucose load. Blockade of cannabinoid CB₂ receptors counteracted this effect, leading to glucose intolerance. Since blockade of cannabinoid CB₁ receptors mimics the actions of cannabinoid CB₂ receptor agonists, we propose that the endocannabinoid system modulates glucose homeostasis through the coordinated actions of cannabinoid CB₁ and CB₂ receptors. We also describe the presence of both cannabinoid CB₁ and CB₂ receptor immunoreactivity in rat pancreatic β - and non- β -cells, adding the endocrine pancreas to adipose tissue and the liver as potential sites for endocannabinoid regulation of glucose homeostasis.

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Keywords: Endocannabinoid; Cannabinoid receptor; Glucose intolerance; Diabetes; Pancreatic islet

1. Introduction

Endocannabinoids are lipid mediators that counteract satiety signals at the gastrointestinal and hypothalamic levels, promoting overfeeding and lipid biosynthesis and storage (Cota et al., 2003; Gomez et al., 2002). A role for cannabinoid CB₁ receptors has been established for these actions (Di Marzo and Matias, 2005). As confirmation, a clinical trial involving obese patients treated with Rimonabant, a cannabinoid CB₁ receptor antagonist, resulted in a relevant reduction in body weight, waist circumference and insulin resistance (Van Gaal et al., 2005). Recent reports have established that in rats cannabinoid CB₁ receptors modulate glucose homeostasis after a glucose load (Bermudez-Silva et al., 2006), and that glucose levels modulate anandamide and 2-arachidonoyl glycerol production by the pancreatic beta-cell line RIN-m5F (Matias

et al., 2006). Additionally, a recent report suggest that there is overactivation of the endocannabinoid system in obese humans and in humans with eating disorders (Engeli et al., 2005; Monteleone et al., 2005). The effects of natural and endogenous cannabinoids on lipid and glucose metabolism might be mediated by cannabinoid CB₁ receptors located in insulin-sensitive tissues such as adipose tissue (Cota et al., 2003) and the liver (Osei-Hyiaman et al., 2005). The endocrine pancreas is an additional potential target for exogenously administered cannabinoids since there are cannabinoid CB₁ receptors in pancreatic islets (Juan-Pico et al., 2006) and in rat insulinoma β -cell line RIN-m5F (Matias et al., 2006). Besides cannabinoid CB₁ receptors, we have described that cannabinoid CB₂ receptors modulate calcium oscillations and insulin secretion in mouse pancreatic islets *in vitro* (Juan-Pico et al., 2006). However, we do not know whether cannabinoid CB₂ receptors modulate glucose homeostasis *in vivo*, a question addressed in this study. To this end we analyzed whether cannabinoid CB₂ receptor-acting drugs modulate glucose homeostasis in rats receiving an i.p. glucose load. We compared these actions with those described for cannabinoid CB₁ receptor antagonists and analyzed the presence of cannabinoid CB₁ and CB₂ receptors in rat pancreatic islets.

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2. Materials and methods

2.1. Animals

We carried out the experiments with male Wistar rats (250 g), in strict compliance with the European Communities directive 86/609/EEC regulating animal research. Animals were housed in groups of four in a room with controlled temperature (20 ± 2 °C) and humidity ($55 \pm 5\%$) with free access to water and standard food pellets. Food was withdrawn in the early morning (4 h before the procedure of glucose tolerance test).

2.2. Glucose tolerance test

This was carried out by injecting an intraperitoneal glucose load of 2 g/kg body wt. Tail blood samples were collected before (0 min) and 5, 10, 15, 30, 60 and 120 min after glucose

administration. Glucose was determined using a standard glucose oxidase method as described previously (Bermudez-Silva et al., 2006).

2.3. Drugs and treatments

Awake rats ($n=8$ per group) were injected i.p. with either vehicle (5% Tween 80 in saline), the mixed cannabinoid CB_1/CB_2 receptor agonists 2-arachidonoylglycerol (2-AG, 4 mg/kg) and (*R*)-(+)-(2,3-dihydro-5-methyl-3-[(morpholinyl)-methyl]-pyrrolo-[1,2,3-de]-1,4-benzoxazinyl)-(1-naphthalenyl) methanone mesylate (WIN 33,212-2, 5 mg/kg), the cannabinoid CB_2 receptor agonist 3-(1',1'-dimethylbutyl)-1-deoxy-delta8-THC, JWH 133 (0.1; 1 and 2 μ g/kg, Huffman et al., 1999), the selective cannabinoid CB_2 receptor antagonist 6-iodo-2-methyl-1-[2-(4-morpholinyl)ethyl]-1*H*-indol-yl](4-methoxyphenyl) methanone (AM630, 0.001, 0.01 and 0.05 mg/kg, Ross et al.,

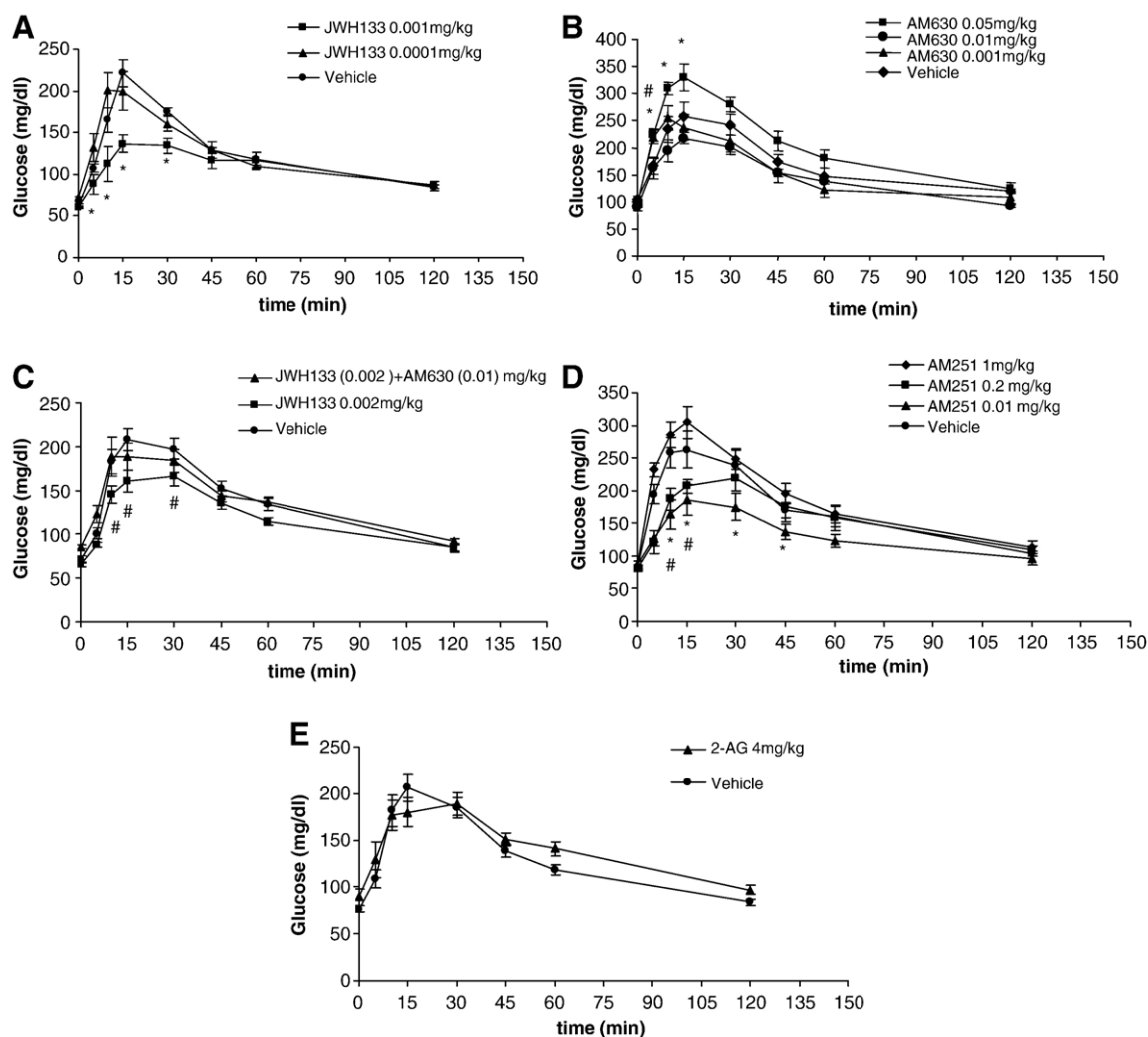


Fig. 1. Administration of the cannabinoid CB_2 receptor agonist JWH 133 improved glucose homeostasis after a glucose load (A). The opposite effect was observed when a cannabinoid CB_2 receptor antagonist, AM630, was administered before glucose (B). Pretreatment with AM630 antagonized the effects of JWH 133, confirming the role of cannabinoid CB_2 receptors on the effects of JWH 133 (C). The effects of administration of a cannabinoid CB_1 receptor antagonist, AM251, resembled the effects of JWH 133, at doses selective for CB_1 receptors, but not at the highest dose tested (1 mg/kg) that targets both cannabinoid CB_1 and CB_2 receptors (D). Administration of 2-arachidonoylglycerol, a full agonist at both cannabinoid CB_1 and CB_2 receptors, did not modify glucose homeostasis (E). Data are means $\pm$ S.E.M. of 8–10 determinations per group. (*, # and &) $P < 0.01$ different doses versus vehicle-treated animals, Newman–Keul's.

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