



## Four new C<sub>18</sub>-diterpenoid alkaloids with analgesic activity from *Aconitum weixiense*

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### ABSTRACT

Four new C<sub>18</sub>-diterpenoid alkaloids, weisaconitines A–D (**1–4**), were isolated from *Aconitum weixiense*. Based on extensive UV, IR, MS, 1D and 2D NMR analyses, their structures were elucidated as 8-O-ethylolaconine (**1**), 4-demethylgenicunine B (**2**), 14-oxoaconosine (**3**), and 8-O-ethylaconosine (**4**). The analgesic activity of compound **4** was studied with CH<sub>3</sub>COOH-induced writhing model in mice. Compound **4** showed writhing inhibitions of 24% (50 mg/kg), 26% (100 mg/kg) and 34% (200 mg/kg), respectively, as compared to the reference drug aspirin (63%) at a dose of 200 mg/kg.

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## 1. Introduction

*Aconitum* L. (Ranunculaceae), annual or perennial herbs, mainly distribute in the temperate regions of the northern hemisphere. There are about 300 species all over the world, of which about 76 *Aconitum* spp. have been used as poisonous and medicinal plants in China [1]. The diterpenoid alkaloids, including 'C<sub>18</sub>-, C<sub>19</sub>- and C<sub>20</sub>-diterpenoid alkaloids', are main chemical constituents with analgesic, cardiotoxic, anti-inflammatory, and anti-arrhythmic activities [2–5]. Three diterpenoid alkaloids, 3-acetylaconitine, lappaconitine, and crassicauline A, used as analgesic drugs taking effects on sodium channel, are clinically employed for the treatment of various pains in China [3].

The roots of *Aconitum weixiense* W. T. Wang, endemic to Weixi county of Yunnan province in China, are utilized to treat pains and rheumatism by natives [6]. Furthermore, we observed that the plant was severely grazed by cattle, but

causing no death to them. Therefore, it was indicative that *A. weixiense* was nontoxic, being prominently different from other *Aconitum* spp.. To find safer and biologically active substances, the roots of *A. weixiense* were phytochemically investigated to afford four new C<sub>18</sub>-diterpenoid alkaloids, weisaconitines A–D (**1–4**, Fig. 1). Compound **4** showed an analgesic activity by CH<sub>3</sub>COOH-induced writhing model in mice. This paper described their isolation, structural elucidation and analgesic activity.

## 2. Experimental

### 2.1. General experimental procedures

Optical rotations were determined on a Jasco model 1020 polarimeter (Horiba, Tokyo, Japan). UV spectra were measured on a Shimadzu UV2401PC spectrophotometer (Shimadzu, Kyoto, Japan). IR (KBr) spectra were recorded on a Bio-Rad FTS-135 spectrometer (Bio-Rad, Hercules, California, USA). 1D and 2D NMR were recorded on Bruker AM-400, Bruker DRX-500 or AVANCE III-600 spectrometers (Bruker, Bremerhaven, Germany). Mass spectra were run on a VG

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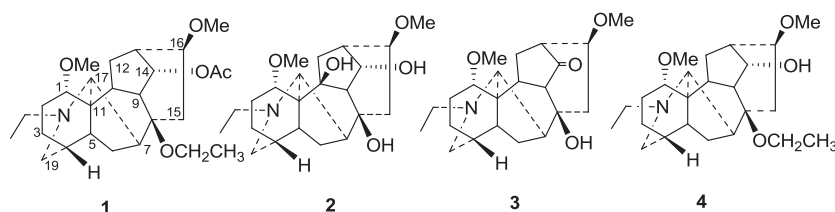


Fig. 1. The structures of compounds 1–4, isolated from *Aconitum weixiense*.

Spec-3000 spectrometer (VG, Manchester, UK) and Waters AutoSpec Premier P776 (Waters, USA). Silica gel (200–300 mesh) for column chromatography and TLC plates (GF<sub>254</sub>) were obtained from Qingdao Makall Chemical Company (Makall, Qingdao, China). Al<sub>2</sub>O<sub>3</sub> for column chromatography was purchased from Shanghai Wusi Chemical Reagents Company, Ltd. (Wusi, Shanghai, China). Fractions were visualized by silica gel plates sprayed with Dragendorff's reagent.

## 2.2. Plant material

The roots of *A. weixiense* W. T. Wang were collected at Weixi county of Yunnan Province, P. R. China, in October, 2012, and authenticated by Prof. Li-Gong Lei (Kunming Institute of Botany, Chinese Academy of Sciences). The voucher specimen (No. YNS2012-26) had been deposited in the Yunnan Research Center on Good Agricultural Practice for Dominant Chinese Medicinal Materials, College of Agriculture and Biotechnology, Yunnan Agricultural University.

## 2.3. Extraction and isolation

The air-dried roots of *A. weixiense* (3 kg) were powdered and extracted three times with MeOH for 2 h under reflux. Being removed solvent under reduced pressure, the crude extract was dissolved with 3 L 1.5% HCl solution. After filtration, the acidic solution was basified to pH 9.0 with ammonia (25%) and extracted with CHCl<sub>3</sub> to obtain crude alkaloidal extract (65 g). The alkaloidal extract was subjected to silica column chromatography (Si CC, 800 g, 8 × 50 cm) and eluted with petroleum ether–acetone–diethylamine (100:1:1, 80:1:1, 50:1:1, 25:1:1, 25:5:1, 25:10:1, 25:25:1, 10:20:1, v/v, each 1 L) gradient to afford five fractions (A–E). Fr. B (25 g) was further separated to obtain five subfractions (B1–B5) by Si CC with petroleum ether–diethylamine (50:1, 40:1, 30:1, 20:1, each 500 mL) as the eluent. Fr. B2 (2.7 g) was performed on Al<sub>2</sub>O<sub>3</sub> CC (2.0 × 25 cm, 30 g) and eluted with petroleum ether–EtOAc (15:1, each 100 mL) to yield compound 1 (7 mg). Fr. B3 (5.8 g) was applied to Al<sub>2</sub>O<sub>3</sub> CC (4 × 22 cm, 112 g) with an eluent of petroleum ether–EtOAc (10:1, each 500 mL), and then purified through Sephadex LH-20 (CHCl<sub>3</sub>/MeOH, 1:1) to produce compounds 3 (10 mg) and 4 (1.8 g). Fr. D (15 g) was further separated to obtain five subfractions (D1–D5) by Si CC with petroleum ether–acetone–diethylamine (15:1:1, 10:1:1, 10:2:1, 10:5:1, v/v, each 500 mL) as the eluent. Fr. D3 (2.1 g) was applied to Al<sub>2</sub>O<sub>3</sub> CC (2.0 × 25 cm, 30 g) with an eluent of petroleum ether–EtOAc (1:3, each 100 mL), and further purified through Sephadex LH-20 (CHCl<sub>3</sub>/MeOH, 1:1) to provide compound 2 (13 mg). All obtained compounds had a degree of purity

>90%, based on the TLC method in three different solvent systems exhibiting one spot with Dragendorff's reagent, and NMR spectra with the smooth baseline and no impurity peak.

**Weisaconitine A (1):** Colorless gum; [ $\alpha$ ]<sub>D</sub><sup>13.5</sup>: –0.90 (c 0.20, MeOH); IR (KBr)  $\nu_{\max}$ : 2925, 1739, 1063 cm<sup>–1</sup>; <sup>1</sup>H NMR and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS  $m/z$  448.3047 ([M + H]<sup>+</sup>, C<sub>26</sub>H<sub>42</sub>NO<sub>5</sub><sup>+</sup>, calcd for 448.3058).

**Weisaconitine B (2):** Colorless powder; [ $\alpha$ ]<sub>D</sub><sup>15.9</sup>: –11.37 (c 0.20, MeOH); IR (KBr)  $\nu_{\max}$ : 3436, 2924, 1059 cm<sup>–1</sup>; <sup>1</sup>H NMR and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS  $m/z$  394.2588 ([M + H]<sup>+</sup>, C<sub>22</sub>H<sub>36</sub>NO<sub>5</sub><sup>+</sup>, calcd for 394.2588).

**Weisaconitine C (3):** Colorless powder; [ $\alpha$ ]<sub>D</sub><sup>13.5</sup>: –35.47 (c 0.10, MeOH); IR (KBr)  $\nu_{\max}$ : 3432, 2922, 1745, 1064 cm<sup>–1</sup>; <sup>1</sup>H NMR and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS  $m/z$  376.2483 ([M + H]<sup>+</sup>, C<sub>22</sub>H<sub>34</sub>NO<sub>4</sub><sup>+</sup>, calcd for 376.2482).

**Weisaconitine D (4):** Colorless powder; [ $\alpha$ ]<sub>D</sub><sup>16.1</sup>: –16.07 (c 0.20, MeOH); IR (KBr)  $\nu_{\max}$ : 3431, 2924, 1064 cm<sup>–1</sup>; <sup>1</sup>H NMR and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS  $m/z$  406.2948 ([M + H]<sup>+</sup>, C<sub>24</sub>H<sub>40</sub>NO<sub>4</sub><sup>+</sup>, calcd for 406.2952).

Table 1  
<sup>13</sup>C NMR data (100 MHz, in CDCl<sub>3</sub>) of compounds 1–4.

Position	1	2	3	4
1	85.8 d	78.6 d	85.5 d	86.2 d
2	28.3 t	29.6 t	27.9 t	29.8 t
3	36.7 t	34.6 t	37.2 t	35.8 t
4	35.8 d	36.2 d	36.0 d	36.6 d
5	44.8 d	41.3 d	45.1 d	45.6 d
6	27.3 t	26.0 t	26.1 t	28.4 t
7	46.7 d	45.2 d	46.0 d	45.9 d
8	77.7 s	72.4 s	83.0 s	77.9 s
9	44.3 d	56.1 d	55.4 d	45.5 d
10	38.2 d	81.1 s	46.2 d	40.7 d
11	49.0 s	53.8 s	50.2 s	50.2 s
12	26.5 t	37.7 t	25.4 t	26.4 t
13	40.9 d	38.1 d	43.8 d	39.2 d
14	75.0 d	74.1 d	216.5 s	75.2 d
15	29.6 t	40.1 t	29.2 t	30.4 t
16	81.3 d	81.6 d	86.4 d	82.7 d
17	61.1 d	64.4 d	64.2 d	62.9 d
19	51.0 t	50.0 t	49.6 t	49.5 t
NCH <sub>2</sub> CH <sub>3</sub>	49.3 t	49.7 t	48.8 t	48.8 t
NCH <sub>2</sub> CH <sub>3</sub>	12.9 q	13.6 q	13.6 q	13.5 q
OCH <sub>2</sub> CH <sub>3</sub> -8	56.3 t	–	–	55.9 t
OCH <sub>2</sub> CH <sub>3</sub> -8	16.0 q	–	–	16.1 q
OMe-1	56.1 q	56.0 q	56.1 q	56.3 q
OMe-16	56.6 q	56.4 q	56.4 q	56.5 q
COMe-14	170.8 s	–	–	–
COMe-14	21.2 q	–	–	–

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