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# Three new sesquiterpenes from *Tithonia diversifolia* and their anti-hyperglycemic activity

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#### ABSTRACT

Three new germacrane sesquiterpenes (1), (2), (3), along with eleven known sesquiterpenes, namely, tirotundin-3-O-methyl ether (4), deacetylvguiestin (5), 1 $\beta$ -hydroxydiversifolin-3-O-methyl ether (6), tagitinin C (7), 1 $\beta$ -hydroxytirotundin-3-O-methyl ether (8), 1 $\beta$ -hydroxytirotundin-1,3-O-dimethyl ether (9), tagitinin F-3-O-methyl ether (10), tagitinin F (11), tagitinin A (12), 3 $\beta$ -acetoxy-4 $\alpha$ -hydroxyeduesm-11(13)-en-12-oic acid (13) and ilicic acid (14) were isolated from the aerial parts of *Tithonia diversifolia*. Their structures were established by spectroscopic analysis, while the relative configuration of compound 1 was confirmed by X-ray diffraction analysis. In addition, compounds 1–14 were evaluated *in vitro* for their anti-hyperglycemic activity by glucose uptake in 3T3-L1 adipocytes. It was found that 10 µg/mL 1, 3, 6 and 8 could significantly increase glucose uptake without significant toxic effects.

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#### 1. Introduction

*Tithonia diversifolia* (Hemsl.) A. Gray is an impressive member of the sunflower family, Asteraceae. It is a perennial native of Mexico and Central America and was introduced to the southern part of Asia, Africa and the Pacific region [1]

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because of ornamental purposes or pharmacological action. T. diversifolia was brought to Yunnan province of China in the 1900s. Traditionally, T. diversifolia is used for the treatment of malaria and other forms of fever or wound in Mexico. Presently, T. diversifolia is of particular interest in health care phytomedical research with respect to its antimalarial [2,3], antidiabetic [4–6], anti-inflammatory [7–10], and anticancer [11,12] activities. Chemical studies on this species have resulted in sesquiterpene lactones, chromene, and flavone [13–16]. The present study is our ongoing research of antidiabetic active metabolites from T. diversifolia, involving the isolation and identification of 14 sesquiterpenes, including three new sesquiterpenes: tagitinin G (1), tagitinin H (2), tagitinin I (3), and 11 known sesquiterpenes: tirotundin 3-O-methyl ether (4) [15], deacetylvguiestin (5) [16], 1 $\beta$ -hydroxydiversifolin-3-O-methyl ether (6) [17], tagitinin C (7) [16], 1 $\beta$ -hydroxytirotundin 3-O-methyl ether (8)

Table 1

<sup>1</sup> H	I (600 MHz	) and	<sup>13</sup> C (15	0 MHz	) NMR	date fo	r compounds	<b>1, 2</b> and <b>3</b> .
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No.	1 <sup>a</sup>		2 <sup>a</sup>		3 <sup>a</sup>	
	δ <sub>H</sub> mult (J in Hz)	$\delta_{C}$	δ <sub>H</sub> mult (J in Hz)	$\delta_{C}$	δ <sub>H</sub> mult (J in Hz)	$\delta_{C}$
1α	4.00 dd (9.0, 9.0)	78.4	1.96 m	38.1	1.72 m	36.6
1β			1.78 m		1.73 m	
2α	2.22 dd (12.6, 9.0)	44.3	1.76 m	33.5	2.39 m	24.6
<b>2</b> β	1.63 dd (12.6, 9.0)		1.70 m		2.39 m	
3		110.4		110.3		150.1
4	1.98 dq (6.6, 19.2)	35.8	1.96 m	35.8		106.6
5α	2.24 ddd (2.4, 5.4, 7.8)	38.3	2.20 m	38.6	2.84 dd (10.8, 13.2)	39.5
5β	1.66 ddd (5.4, 12.0, 18.6)		1.68 m		2.14 d (13.2)	
6	4.40 ddd (2.4, 10.2, 12.6)	79.4	4.32 ddd (3.0, 10.8, 12.0)	80.0	4.31 dd (5.4, 10.8)	78.4
7	2.50 ddd (7.8, 10.2, 18.0)	60.3	2.47 ddd (7.8, 10.2, 18.0)	60.1	3.97 dddd (2.4, 3.0, 3.6, 5.4)	49.5
8	3.81 dd (8.4, 7.8)	69.8	3.75 dd (8.4, 7.8)	69.5	5.43 ddd (2.4, 5.4, 10.8)	70.2
9α	2.91 dd (8.4, 13.8)	38.6	2.22 m	47.2	1.64 dd (5.4, 13.8)	37.4
<b>9</b> β	1.60 d (8.4)		1.79 m		1.71 dd (10.8, 13.8)	
10	(0.4)	82.7		78.5	(10.8, 15.8)	80.1
11	2.37 dq (6.6, 19.2)	43.0	2.32 dq (6.6, 13.2)	43.0		136.9
12		178.4		178.6		169.4
13α	1.23 d (6.6)	13.6	1.18 d (6.6)	13.9	6.27 d (3.6)	122.0
<b>13</b> β					5.56 d (3.0)	
14	1.40 s	26.8	1.39 s	28.8	1.47 s	23.2
15	0.95 d (7.2)	15.6	0.97 d (7.2)	15.9	1.69 s	18.5
1'						176.3
2'					2.40 m	34.0
3'					1.02 d (6.6)	19.1
4'					1.04 d (6.6)	18.7

<sup>a</sup> Spectra were measured in CDCl<sub>3</sub>.

[12], 1 $\beta$ -hydroxytirotundin-1,3-O-dimethyl ether (9) [18], tagitinin F-3-O-methyl ether (10) [17], tagitinin F (11) [19], tagitinin A (12) [16], 3 $\beta$ -acetoxy-4 $\alpha$ -hydroxyeduesm-11(13)-en-12-oic acid (13) [20], ilicic acid (14) [21,22]. In addition, the anti-hyperglycemic activities of compounds 1-14 were evaluated by glucose uptake in 3T3-L1 adipocytes. The purity of the compounds ranged from 90.5% (14) to 97.3% (1) as determined by analytical HPLC with DAD and ELSD detection.

#### 2. Experimental

#### 2.1. General experimental procedures

Optical rotation was measured on a Perkin Elmer polarimeter (serial No. 9903). <sup>1</sup>H NMR (600 MHz), <sup>13</sup>C NMR (150 MHz) spectra and all 2D NMR spectra were obtained on a Bruker Avance 600 NMR spectrometer (Bruker Co., Germany). Chemical shifts are expressed in  $\delta$  (ppm) referring to the solvent peaks  $\delta_H$  7.26 and  $\delta_C$  77.0 for CDC<sub>13</sub>,  $\delta_H$  7.22 and  $\delta_C$  135.5 for pyridine, and the coupling constant (J) is given in Hz. HRESI-MS was recorded on a Varian MAT-212 mass spectrometer and a Agilent Technologies 6538 UHD Accurate-Mass Q-TOF LC/MS spectrometer (Agilent Technologies, MA, USA); GC/ MS(Termo Finnigan Trace GC apparatus, L-Chirasil-Val column, 25 m×0.32 mm). IR was recorded on a Bruker Vector 22 spectrometer with KBr pellet. The purity (%) of compounds were tested with an Agilent 1200 instrument using using a diamonsil C18 column (250 mm  $\times$  4.6 mm, 5  $\mu$ m) eluting with MeOH-H<sub>2</sub>O (30%-60%) at 1.0 mL/min detected by SEDEX 75 ELSD (Dikma Technologies, USA) and DAD detector (Agilent Technologies, MA, USA). Crystallographic data were collected on a Bruker SMART diffractometer using a graphitedater with Mo K $\alpha$  radiation. Other materials included Sephadex LH-20 (Pharmacia), Silica gel GF254 (Luyou company of Yantai, 100-200 and 200-300 mesh); RP-C18 (Merck, 43-60 µm), TLC GF254 and preparative TLC GF254 (Luyou company of Yantai); TLC Silica gel 60 RP-18F254 (Merck); MCI gel (Mitsubishi chemical corporation); 3T3-L1 cell line (Tsucuba Cell Bank, Japan); Pioglitazone (Sigma Aldrich, P4120; purity $\geq$ 99.9%) was used as a positive control.

#### 2.2. Plant material

The aerial parts of *T. diversifolia* (Hemsl.) A. Gray were collected in Mengzi of Yunnan province, China in September 2007 and identified by Prof. Wansheng Chen (Department of Pharmacy, Changzheng Hospital, Second Military Medical University). A voucher specimen (no. 20070820) was deposited in the Department of Pharmacognosy of the Second Military Medical University in Shanghai, China.

#### 2.3. Extraction and isolation

The dried aerial parts of *T. diversifolia* (21 kg) were percolated with 80% EtOH at room temperature. The EtOH extract was concentrated to an aqueous residue and suspended with water. The water layer was extracted with petroleum ether, EtOAc and n-BuOH, and the EtOAc soluble fraction (128.0 g) was separated by column chromatography using silica gel with Petroleum ether-EtOAc (30:1; 15:1; 8:1; 4:1; 2:1; 1:1), resulting in six major fractions (1–6). Fraction 2 (6.5 g) was further separated

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