



## Four new neolignans from the leaves of *Tripterygium wilfordii*

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### ABSTRACT

Four new neolignans, wilfordiols A–D (1–4), together with five known compounds (5–9), were isolated from an aqueous extract of the dried leaves of *Tripterygium wilfordii*. Their structures were determined by spectroscopic methods, including 1D and 2D NMR, HRESIMS, and CD experiments. The anti-inflammation activities of compounds 1–9 were evaluated by the inhibitory effect on NO production, *in vitro*.

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## 1. Introduction

*Tripterygium wilfordii* Hook f. (Celastraceae) is widespread in southern China. The root of *T. wilfordii* has been used as a traditional medicine for the treatment of rheumatoid arthritis, skin disorders, male-fertility control, and other inflammatory autoimmune diseases [1]. Previous investigations revealed that the root of *T. wilfordii* mainly contained triterpenoids, diterpenoids, and sesquiterpene alkaloids [2]. Some of them exhibited interesting pharmacological activities including anti-inflammatory, antitumor, antifertility and insecticidal [3]. However, few studies have been performed on the chemical constituents and bioactivities of the leaves of *T. wilfordii*, compared with its roots. As a part of our search for bioactive constituents from *T. wilfordii*, the aqueous extract of the leaves of *T. wilfordii* has been investigated. In this paper, we described

herein the isolation, structure elucidation and biological assays of four new neolignans, named as wilfordiols A–D (1–4) (Fig. 1).

## 2. Experimental procedure

### 2.1. General

CD spectra were measured on a Jasco J-815 CD spectrophotometer. UV spectra were recorded in MeOH on a Jasco V650 spectrophotometer. IR spectra were measured on an IMPACT-400 FT-IR spectrophotometer (Thermo Nicolet Corporation, America). NMR spectra were recorded on an INOVA-500 spectrometer with tetramethylsilane (TMS) as internal standard. ESI-MS experiments were performed on an Agilent 1100 Series LC/MSD Trap-SL mass spectrometer. HRESIMS data were measured by an Agilent Technologies 6520 Accurate Mass Q-ToF LC/MS spectrometer. TLC was performed on precoated silica gel GF<sub>254</sub> plates (Qingdao Haiyang Chemical Co. Ltd.). Column chromatography was carried out with silica gel 100–200 and 200–300 (Qingdao Haiyang Chemical Co. Ltd.) and Sephadex LH-20 (Pharmacia, Sweden).

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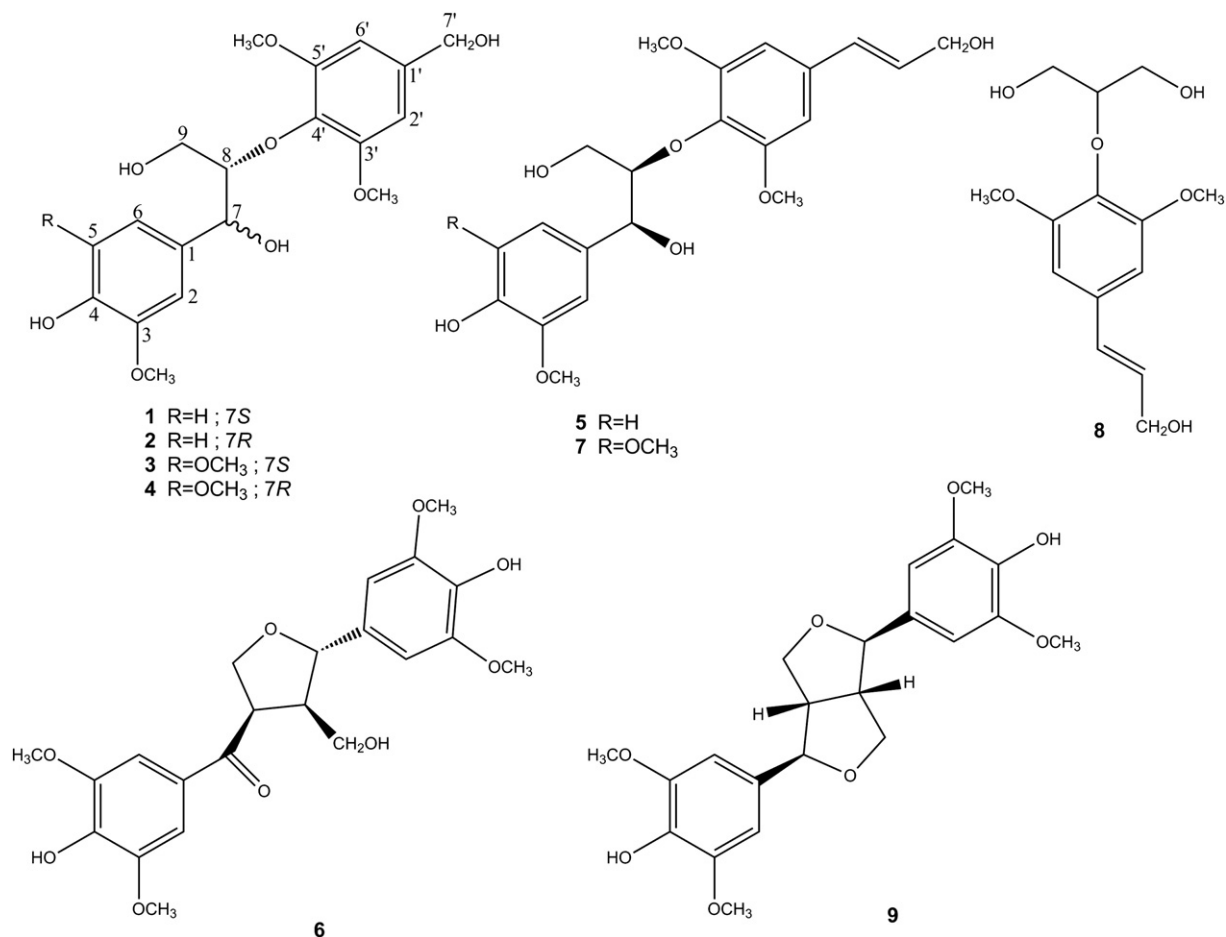


Fig. 1. Structures of compounds 1–9.

## 2.2. Plant material

The leaves of *Tripterygium wilfordii* were collected in Taining County, Fujian Province (2009), and identified by Prof. Lin Ma. A voucher specimen (Herbarium No. 20090034) was deposited at Institute of Materia Medica, Chinese Academy of Medical Sciences.

## 2.3. Extraction and isolation

Air-dried leaves of *Tripterygium wilfordii* Hook f. (8 kg) were powdered and refluxed by water for three times, then defatted with petroleum ether, and extracted with chloroform. The extraction was chromatographed on a silica gel, eluted with cyclohexane–ethyl acetate (50:1 to 5:1, v/v) to afford five fractions (A–E).

The Fr. B (55 g) was chromatographed on a silica gel column using a gradient of CHCl<sub>3</sub>–MeOH (50:1 to 5:1, v/v) as eluent and gave 122 fractions (1–122). Fr. 21–26 (17.54 g) was further chromatographed again on a silica gel column eluting with a gradient of CHCl<sub>3</sub>–MeOH (100:1 to 50:1, v/v) and yielded 9 fractions (21-1–21-9). Fraction 21-8 (2.6 g) was further isolated by Sephadex LH-20 (MeOH). Then fraction 21-8-6 (662 mg) was separated by HPLC (35% MeOH–H<sub>2</sub>O for

the first time, and then 15%–18% CH<sub>3</sub>CN–H<sub>2</sub>O) to obtained compounds 1 (10 mg), 2 (8 mg), 3 (7 mg), 4 (4 mg), 5 (37 mg), 6 (45 mg), 7 (4 mg), 8 (6 mg) and 9 (20 mg).

Compound 1: light yellow oil;  $[\alpha]_D^{20}$  –2.9 (MeOH, c 0.10); UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ) 204 (4.64), 233 (3.98), 279 (3.50) nm; CD (MeOH) 217 ( $\Delta\epsilon$  +1.08), 238 ( $\Delta\epsilon$  +0.38), 269 ( $\Delta\epsilon$  +0.29), 309 ( $\Delta\epsilon$  +0.27) nm; IR (KBr)  $\nu_{\max}$  3409, 1594, 1516 and 825 cm<sup>–1</sup>; ESI-MS:  $m/z$  403 [M+Na]<sup>+</sup>, HRESIMS:  $m/z$  403.1361 [M+Na]<sup>+</sup> (calc. for C<sub>19</sub>H<sub>24</sub>NaO<sub>8</sub> 403.1363). <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD) and <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD); see Tables 1 and 2.

Compound 2: light yellow oil;  $[\alpha]_D^{20}$  +1.9 (MeOH, c 0.10); UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ) 204 (4.54), 230 (3.91), 279 (3.34) nm; CD (MeOH) 222 ( $\Delta\epsilon$  +0.73), 242 ( $\Delta\epsilon$  +0.26), 264 ( $\Delta\epsilon$  +0.19), 306 ( $\Delta\epsilon$  +0.17) nm; IR (KBr)  $\nu_{\max}$  3366, 1594, 1515 and 826 cm<sup>–1</sup>; ESI-MS:  $m/z$  403 [M+Na]<sup>+</sup>, HRESIMS:  $m/z$  403.1368 [M+Na]<sup>+</sup> (calc. for C<sub>19</sub>H<sub>24</sub>NaO<sub>8</sub> 403.1363). <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD) and <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD); see Tables 1 and 2.

Compound 3: light yellow oil;  $[\alpha]_D^{20}$  +15.6 (MeOH, c 0.10); UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ) 207 (4.42), 229 (3.87), 242 (3.65), 273 (3.13) nm; CD (MeOH) 218 ( $\Delta\epsilon$  +2.10), 238 ( $\Delta\epsilon$  +0.47), 261 ( $\Delta\epsilon$  +0.71), 307 ( $\Delta\epsilon$  +0.35) nm; IR (KBr)  $\nu_{\max}$  3403, 1594, 1511 and 836 cm<sup>–1</sup>; ESI-MS:  $m/z$  433 [M+Na]<sup>+</sup>, HRESIMS:

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