



## New *neo*-clerodane diterpenoids from *Scutellaria barbata* with cytotoxic activities

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### ABSTRACT

Two new *neo*-clerodane diterpenoids have been isolated from the whole plant of *Scutellaria barbata* D. Don, and their structures were established by detailed spectral analyses as scutehenanine H (**1**) and 6-(2,3-epoxy-2-isopropyl-n-propoxyl)barbatin C (**2**). *In vitro*, the isolated two new compounds showed significant cytotoxic activities against three human cancer lines, and gave IC<sub>50</sub> values in the range of 2.0–4.2 μM.

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### 1. Introduction

*Scutellaria* L. (Labiatae) is a large subcosmopolitan genus with 360 currently recognized species [1]. Recently, plants belonging to this genus have attracted attention owing to interesting biological activities observed for some *neo*-clerodane diterpenoids isolated from them. *Scutellaria barbata* D. Don is a perennial herb which is natively distributed throughout Korea and southern China. This herb is known in traditional Chinese Medicine as Ban-Zhi-Lian and traditional Korean medicine as Banjiryun, respectively, and has been used as an anti-inflammatory and antitumor agent [2–4]. In previous phytochemical studies on *S. barbata* collected from Linyi district (Shandong Province, China), we reported the isolation of a series of *neo*-clerodane diterpenoids [5–7]. From a chemotaxonomic point of view, it is of interest to note that these *neo*-clerodane diterpenoids lack an oxygenated sub-

stituent at C-19 found in almost all of the *neo*-clerodane diterpenoids from European *Scutellaria* species [8]. The oxidation pattern of the *neo*-clerodane diterpenoids indicates a phylogeographical and an evolutionary relation between some Asiatic and European *Scutellaria* species. This relation has been proposed previously, based on both botanical considerations and the global distribution of the genus in the reported literature [9].

As part of our ongoing search for more novel *neo*-clerodane diterpenoid, we have investigated the aerial parts of *S. barbata* collected from the Zhumadian district, Henan Province, China. The EtOH extract of this species was successively partitioned with CHCl<sub>3</sub> and EtOAc. The CHCl<sub>3</sub> fraction was subjected to extraction with 3% HCl, and the aqueous layer was basified with NH<sub>4</sub>OH and extracted with CHCl<sub>3</sub>. The CHCl<sub>3</sub> fraction was concentrated in vacuo and sequentially subjected to column chromatography over silica gel, silica gel RP-18, and Sephadex LH-20 to give two new *neo*-clerodane diterpenoids. By means of extensive spectroscopic analyses, the structures of two new compounds **1–2** were elucidated as scutehenanine H (**1**), and 6-(2,3-epoxy-2-isopropyl-n-propoxyl)barbatin C (**2**). In addition, the isolated

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two new compounds were screened for cytotoxicity against three tumor cell lines (HONE-1 nasopharyngeal, KB oral epidermoid carcinoma, and HT29 colorectal carcinoma cells), with IC<sub>50</sub> values being in the range of 2.0–4.2  $\mu$ M. Herein we report on the isolation, structure elucidation, as well as the evaluation of cytotoxic effects of these two new neoclerodane diterpenoids.

## 2. Experimental

### 2.1. General experimental procedures

Melting points were measured on an XT-4 micro-melting point apparatus and are uncorrected. Optical rotations were measured on a Perkin-Elmer 241 polarimeter. UV spectra were obtained on a Shimadzu UV-160 spectrophotometer. IR spectra were recorded on a Perkin-Elmer 683 infrared spectrometer with KBr disks. FABMS and HR-FABMS were recorded on an Autospec-Ultima ETOF MS spectrometer. NMR spectra were recorded on a Varian Unity BRUKER 400 at 400 MHz (<sup>1</sup>H) and 100 MHz (<sup>13</sup>C), with TMS as the internal standard. Silica gel (200–300 mesh) for column chromatog-

raphy was obtained from Qingdao Marine Chemical Factory, Qingdao, People's Republic of China. Precoated plates of silica gel GF254 and silica gel RP-18 F254s (Merck) were used for TLC, and detected under UV light.

### 2.2. Plant material

*S. barbata* D. Don was collected in Zhumadian district, Henan Province, People's Republic of China, in September 2008, and identified by Professor Yan-yan Zhao, School of Pharmaceutical Science, Yantai University. The whole plants of *S. barbata* was harvested and air-dried at room temperature in the dark. A voucher specimen (YP08077) has been deposited at the Herbarium of School of Pharmaceutical Science, Yantai University.

### 2.3. Extraction and isolation

The air-dried whole plant (49.6 kg) of *S. barbata* was finely cut and extracted three times with refluxing EtOH. Evaporation of the solvent under reduced pressure provided the ethanolic extract. The extract was dissolved and suspended in H<sub>2</sub>O, and

**Table 1**  
NMR data of compounds **1–2** (in CDCl<sub>3</sub>)<sup>a,b</sup>.

No.	<b>1</b>		<b>2</b>	
	$\delta_H$	$\delta_C$	$\delta_H$	$\delta_C$
1	1.76 (m, H <sub>a</sub> -1) 2.03 (m, H <sub>b</sub> -1)	18.3 CH <sub>2</sub>	1.31 (m, H <sub>a</sub> -1) 1.65 (m, H <sub>b</sub> -1)	19.4 CH <sub>2</sub>
2	2.68 (m, 2H)	26.2 CH <sub>2</sub>	2.02 (m, 2H)	26.2 CH <sub>2</sub>
3	5.31 (br s)	123.1 CH	5.21 (br s)	123.0 CH
4		141.5 C		141.1 C
5		42.8 C		43.0 C
6	5.33 (d, 10.0)	77.7 CH	5.34 (d, 9.8)	81.4 CH
7	3.72 (d, 10.0)	74.7 CH	3.85 (d, 9.8)	73.9 CH
8		84.7 C		77.5 C
9		43.3 C		47.6 C
10	2.46 (dd, 2.4, 12.2)	40.3 CH	2.24 (dd, 2.3, 12.0)	42.5 CH
11	6.05 (dd, 3.9, 12.0)	75.6 CH	6.39 (d, 16.8)	147.8 CH
12	2.09 (m, H <sub>a</sub> -12) 2.35 (m, H <sub>b</sub> -12)	30.7 CH <sub>2</sub>	6.36 (d, 16.8)	121.5 CH
13		79.9 C		162.3 C
14	4.49 (s)	76.2 CH	5.91 (br s)	114.6 CH
15		175.3 C		173.9 C
16	4.26 (d, 8.7, H <sub>a</sub> -16) 4.30 (d, 8.7, H <sub>b</sub> -16)	75.8 CH <sub>2</sub>	4.99 (dd, 1.3, 8.7, H <sub>a</sub> -16) 5.02 (dd, 1.3, 8.7, H <sub>b</sub> -16)	70.7 CH <sub>2</sub>
17	1.72 (s, 3H)	18.0 CH <sub>3</sub>	1.16 (s, 3H)	22.5 CH <sub>3</sub>
18	1.61 (s, 3H)	20.9 CH <sub>3</sub>	1.61 (s, 3H)	20.8 CH <sub>3</sub>
19	1.38 (s, 3H)	17.2 CH <sub>3</sub>	1.23 (s, 3H)	17.5 CH <sub>3</sub>
20	1.14 (s, 3H)	17.0 CH <sub>3</sub>	1.12 (s, 3H)	15.4 CH <sub>3</sub>
1'		164.8 C	4.17 (br s, 2H)	61.6 CH <sub>2</sub>
2'		125.9 C		78.4 C
3'	9.22 (br s)	150.7 CH	2.71 (d, 5.9, H <sub>a</sub> -3') 2.98 (d, 5.9, H <sub>b</sub> -3')	38.3 CH <sub>2</sub>
4'			2.31 (sept, 6.8)	34.9 CH
5'	8.85 (br d, 4.5)	153.6 CH	0.97 (3H, d, 6.8)	17.1 CH <sub>3</sub>
6'	7.44 (dd, 4.5, 7.6)	123.5 CH	1.07 (3H, d, 6.8)	16.9 CH <sub>3</sub>
7'	8.29 (br d, 7.6)	137.5 CH		
1''		167.5 C		
2''		130.7 C		
3'' and 7''	8.07 (m, 2H)	129.7 CH		
4'' and 6''	7.48 (m, 2H)	128.4 CH		
5''	7.57 (br t, 7.7)	133.1 CH		

<sup>a</sup> Coupling constants (in Hz) were presented in parentheses.

<sup>b</sup> The assignments were based on DEPT, HMQC, HMBC, and <sup>1</sup>H–<sup>1</sup>H COSY experiments.

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