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# Glycyrrhizin derivative inhibits eotaxin 1 production via STAT6 in human lung fibroblasts

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#### Abstract

We recently demonstrated that glycyrrhizin (GL) and its derivatives down-regulate  $TNF\alpha$ - and IL-4-induced eotaxin 1 production by the human fetal lung fibroblast line HFL-1 at protein or mRNA levels. In particular, the GL derivative hetero-30-OH-GL (3 $\beta$ -[(2-O- $\beta$ -D-glucopyranuronosyl- $\beta$ -D-glucopyranuronosyl)oxy]-olean-11,13(18)-dien-30-ol) showed marked inhibition of eotaxin 1 production with less cytotoxicity than 18 $\beta$ -GL. To identify the molecular mechanism of this effect, we focused on the inhibition of the transcriptional factors NF- $\kappa$ B and signal transducer and activator of transcription 6 (STAT6), which regulate eotaxin 1 gene activation. STAT6 phosphorylation and translocation of phospho-STAT6 from cytosol to nuclei were slightly inhibited by 18 $\beta$ -GL and significantly inhibited by hetero-30-OH-GL. While I $\kappa$ B $\alpha$  degradation and translocation of NF- $\kappa$ B p65 to nuclei were not significantly affected by either compound, the stability of eotaxin-1 mRNA was decreased with hetero-30-OH-GL. In addition, eotaxin 1 promoter activity was markedly inhibited by hetero-30-OH-GL. Electrophoretic mobility shift assay (EMSA) confirmed these results. Thus, hetero-30-OH-GL significantly inhibited eotaxin 1 expression by the selective inhibition of IL-4 signal transduction as well as by enhanced mRNA degradation.

Keywords: Glycyrrhizin derivatives; Eotaxin 1; Human lung fibroblast; Signal transducer and activator of transcription 6; Nuclear factor-kappa B

#### 1. Introduction

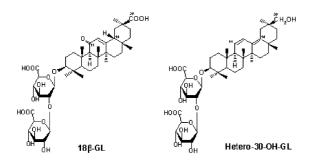
Eotaxin 1 is a typical eosinophil chemoattractant cytokine belonging to the C–C chemokine family [1]. A large number of studies have indicated that eotaxin 1 plays an important role in Th2-type diseases like asthma and atopic dermatitis. IL-4, one of the Th2-type cytokines, promotes the production of eotaxin 1 and several other chemokines, namely eotaxin 2, eotaxin 3,

RANTES, MCP-4, MDC and TARC, by tissue resident non-immune cells such as lung fibroblasts, bronchial epithelial cells and dermal fibroblasts [2–6], and by these means attracts immune cells toward the inflammatory site. Further, stimulation with IL-4 and TNFα in combination synergistically increases the production of eotaxin 1 by activating the transcriptional factors STAT6 and NF-κB [7,8]. A number of studies have reported that glucocorticoids, PPARγ agonists and retinoic acid attenuate eotaxin 1 production [4,6,9–11], but no pathway for this effect has been identified, including any role of STAT6 and NF-κB [10,11].

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More recently, we have shown that glycyrrhizin (GL) and its derivatives inhibit eotaxin 1 production at protein or mRNA expression levels [12]. GL is an aqueous extract of licorice root which consists of glycyrrhetic acid and two molecules of glucuronic acid at the C-3 position (Fig. 1). A GL preparation, Stronger Neo-Minophagen C (SNMC), has long been used for the treatment of chronic hepatitis in Japan [13], and the activity of GL and its derivatives in their targeting of liver [14–16] and other diseases [12,17,18] has been widely investigated.

In the present study, we investigated the mechanism of action of  $18\beta$ -GL and  $(3\beta$ -[(2-O- $\beta$ -D-glucopyranuronosyl- $\beta$ -D-glucopyranuronosyl)oxy]-olean-11,13(18)-dien-30-ol) (hetero-30-OH-GL), a GL derivative considered to have therapeutic potential, on the inhibition of TNF $\alpha$ -and IL-4-induced eotaxin 1 production mediated by the activation of transcriptional factors NF- $\kappa$ B and STAT6.



Eotaxin 1 production (ng/mL) by TNF $\alpha$  and L-4 stimuli for 48 hr

Concentration (µg/mL)	18 <b>β-</b> GL F	Hetero-30-OH-GL
0	12	12
1	$11 \pm 3.4$	$8.1 \pm 0.6^{**}$
10	$10.1 \pm 0.9$	$7.9 \pm 1.3^*$
100	$9.3 \pm 0.7$	$0.6 {\pm} 0.8^{**}$
1000	7.2±0.7 <sup>**</sup>	
IC <sub>50</sub>	>1000	33
CC30	>1000	>100

\* P<0.05 \*\*P<0.01

Fig. 1. Structure of 18β-GL and Hetero-30-OH-GL and their effects on TNFα plus IL-4-induced eotaxin 1 production. Eotaxin 1 levels released during stimulation of HFL-1 cells ( $5 \times 10^4$  cells) with TNFα (10 ng/mL) plus IL-4 (20 U/mL) and various concentrations of each GL compound for 48 h at 37 °C were calculated from ELISA data (n=3) reported previously [12]. Statistically different from the effect of TNFα plus IL-4 at \* P < 0.05, \*\* P < 0.01 level according to Student's t-test. Inhibitory concentration (IC<sub>50</sub>) against eotaxin 1 production of each GL compound and their cytotoxicity concentration (CC<sub>30</sub>) values assessed by LDH assay were also excerpted from the

#### 2. Materials and methods

#### 2.1. Cell culture and materials

Human fetal lung fibroblasts (HFL-1, from normal embryonic lung) were purchased from Riken Cell Bank (Saitama, Japan). The cells were cultured in Ham' F-12 (Gibco, Grand Island, NY) supplemented with 15% heat-inactivated fetal bovine serum, penicillin G 100 units/mL, and streptomycin 100 μg/mL in a humidified 5%  $CO_2/95\%$  air atmosphere at 37 °C. Human recombinant TNFα was kindly provided by Dainippon Pharmaceutical Co. (Suitashi, Osaka, Japan). Human recombinant IL-4 was purchased from Peprotech EC (London, UK). 18β-GL and hetero-30-OH-GL were prepared as described previously [12].

#### 2.2. Western blot analysis

HFL-1 cells (approximately  $3 \times 10^6$ ) were treated with TNFa (10 ng/mL) plus IL-4 (20 U/mL) and each GL compound for 0-60 min and washed with PBS. Whole cell lysates and cytoplasmic and nuclear extracts were prepared as described elsewhere [19-21]. Sample protein concentrations were determined by Bradford assay, and the samples were stored at -80 °C. Whole cell lysate (20 µg), cytoplasmic extract (20 µg) or nuclear extract (10 µg) were boiled for 5 min in an equal volume of 2 × Laemmli's sample buffer, then separated by electrophoresis on 10-12% SDS-PAGE and transferred to a nitrocellulose transfer membrane (Protoran, Schleicher and Schuell BioScience, Germany). Membranes were blocked in 5% nonfat milk powder in TPBS (0.05% Tween 20 containing PBS) at 4 °C overnight, then incubated with the following antibodies diluted in TPBS for 1 h at room temperature with continuous shaking: rabbit polyclonal anti-phospho-STAT6 Ab (Tyr641) (1:1000, Cell Signaling); rabbit polyclonal anti-STAT6 Ab (1:500, SC-621, Santa Cruz); rabbit polyclonal anti-NF-κB p65 Ab (1:1000, SC-372, Santa Cruz); rabbit polyclonal anti-IκBα Ab (1:500, SC-371, Santa Cruz); or goat polyclonal antiactin Ab (1:500, SC-1615, Santa Cruz). After washing with TPBS (5 times  $\times$  5 min each; room temperature), membranes were incubated with horseradish peroxidase-conjugated antirabbit or anti-goat immunoglobulin (DAKO) diluted 1:1000 in TPBS containing 1% nonfat milk powder for 1 h at room temperature with continuous shaking. After washing with TPBS (5 times × 5 min each; room temperature), immunoreactive bands were visualized with an ECL detection system (Amersham).

#### 2.3. Plasmids, transient transfection and luciferase assay

A 442-bp eotaxin 1 promoter fragment (site – 406 to +36) was amplified by PCR using a sense primer (5'-GGTAC-CATGTGAACACAGGAATC-3') containing a restriction site for *Kpn*I and an antisense primer (5'-AAGCTTTGGCGT-GAGAGGTGG-3') containing a restriction site for *Hind*III

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