



## Antimicrobial compounds isolated from *Haematoxylon brasiletto*

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### ABSTRACT

**Ethnopharmacological relevance:** The decoction of the bark of *Haematoxylon brasiletto* Karst. (Leguminosae), commonly known as “Palo Brasil”, is used in the Mexican traditional medicine to treat mouth and kidney infections, hypertension, stomach upsets, gastric ulcers and diabetes.

**Aim of the study:** The present study was performed to evaluate the antimicrobial effects of the methanolic extract of the bark of *Haematoxylon brasiletto*.

**Materials and methods:** A panel of 12 bacteria and the yeast *Candida albicans* were used. Minimum inhibitory concentrations (MICs) were determined by the standard broth microdilution method.

**Results:** The results indicate that the extract of *Haematoxylon brasiletto* inhibited the growth of eight of the tested microorganisms at a concentration limit of 128 µg/mL. For the tested compounds the MIC values ranged from 8.7 to 128 µg/mL.

**Conclusions:** The overall results provide promising baseline information for the potential use of the extracts from *Haematoxylon brasiletto* as well as some of the isolated compounds in the treatment of bacterial infections.

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## 1. Introduction

*Haematoxylon brasiletto* Karst. (Leguminosae) is a large tree very abundant in the south-east Mexico. It is commonly called “azulillo”, “corteza de Brasil”, “palo Brasil”, “palo de tinta” and “palo tinto”. The wood derives its name from the word *baza*, meaning fiery red, and has no geographic connection with Brazil or with common brazilwood, *Caesalpinia echinata* or *Caesalpinia brasiliensis* (Craig et al., 1965). As a home remedy, the bark prepared as a tea is employed for treating hypertension, stomach upsets, mouth infections, diarrhea, gastric ulcers and diabetes (Argueta et al., 1994). An ethanolic extract of the stem bark of *Haematoxylon brasiletto* was found to inhibit the growth of *Escherichia coli* O157:H7 (EHEC), verotoxin production, and adhesion of *Escherichia coli* O157:H7 to HeLa cells (Heredia et al., 2005). In previous work, the bark has afforded hematoxylin (1) and brazilin (2) (Sanchez-Marroquin et al., 1958; Pratt and Yuzuriha, 1959). The plant was selected for fractionation after its methanol-soluble extract exhibited significant antibacterial activity against 12 bacteria and the yeast *Candida albicans*.

## 2. Methodology

### 2.1. Plant material

The air-dried bark of the *Haematoxylon brasiletto* Karst. were obtained in the Mercado de Sonora, México, D.F. Voucher specimens (Bye 20339) have been deposited at the ethnobotanical collection of the National Herbarium (MEXU), Instituto de Biología, UNAM.

### 2.2. Activity guided compound isolation

The air-dried stem bark (1.0 kg) of *Haematoxylon brasiletto* was extracted by maceration with methanol (3 × 2 L) at room temperature (24 h each) and the combined MeOH extracts were evaporated under reduced pressure, yielding a residue (100 g), which was partitioned with petroleum ether, dichloromethane and ethyl acetate. Water was added to the MeOH extract to afford a 50% aqueous MeOH solution before partitioning with dichloromethane. The resulting four extracts (petroleum ether, dichloromethane, ethyl acetate and aqueous) were evaluated against 12 bacteria and the yeast *Candida albicans*. The crude ethyl acetate-soluble extract (35.7 g) was found to exhibit the stronger antibacterial activity (Table 1). Hence the ethyl acetate-soluble extract was subjected to silica gel column chromatography and eluted with gradient mixtures of hexanes–dichloromethane and then acetone and acetone–methanol of increasing polarity to give 15 pooled

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**Table 1**  
Minimum inhibitory concentrations in of the extract ( $\mu\text{g/mL}$ ), the compounds isolated from the bark of *Haematoxylon brasiletto* bark and reference antibiotics ( $\mu\text{g/mL}$  and mM)

Microorganisms	Extract	Compounds								Reference antibiotics	
	MeOH	1	2	3	4	5	6	7	8	PG <sup>a</sup>	CHX <sup>a</sup>
<i>Staphylococcus aureus</i> 375 <sup>b</sup>	64	16 (4.83)	16 (4.57)	32 (5.76)	32 (5.88)	32 (5.44)	32 (4.03)	128 (20.99)	64 (13.82)	0.06 (0.02)	
<i>Staphylococcus aureus</i> 310 (MR) <sup>b</sup>	128	32 (9.86)	32 (9.15)	64 (11.52)	64 (11.76)	128 (21.76)	128 (16.13)	128 (20.99)	128 (27.64)	64 (21.40)	nt
<i>Staphylococcus aureus</i> ATCC 25923 <sup>b</sup>	16	16 (4.83)	16 (4.57)	32 (5.76)	16 (2.94)	32 (5.44)	64 (8.06)	64 (10.49)	32 (6.91)	0.06 (0.02)	nt
<i>Enterococcus faecium</i> 379 (VR) <sup>b</sup>	16	16 (4.83)	16 (4.57)	32 (5.76)	16 (2.94)	32 (5.44)	32 (4.03)	64 (10.49)	128 (27.64)	32 (10.70)	nt
<i>Bacillus subtilis</i> 327 <sup>b</sup>	16	32 (9.66)	64 (18.30)	64 (11.52)	64 (11.76)	64 (10.88)	64 (80.6)	64 (10.49)	>128 (27.64)	32 (10.70)	nt
<i>Escherichia coli</i> ipm 389 <sup>b</sup>	>128	64 (19.32)	128 (36.61)	>128 (23.04)	>128 (23.52)	>128 (21.76)	>128 (16.13)	>128 (20.99)	>128 (27.64)	2 (0.66)	nt
<i>Escherichia coli</i> 442 <sup>b</sup>	>128	64 (19.32)	64 (36.61)	>128 (23.04)	>128 (23.52)	>128 (21.76)	>128 (16.13)	>128 (20.99)	>128 (27.64)	32 (10.7)	nt
<i>Escherichia coli</i> ATCC 25922 <sup>b</sup>	>128	128 (38.64)	128 (36.61)	128 (23.04)	>128 (23.52)	>128 (21.76)	>128 (16.13)	>128 (20.99)	>128 (27.64)	128 (42.80)	nt
<i>Klebsiella pneumonia</i> 425 <sup>b</sup>	128	128 (38.64)	128 (36.61)	>128 (23.04)	>128 (23.52)	>128 (21.76)	>128 (16.13)	>128 (20.99)	>128 (27.64)	128 (42.80)	nt
<i>Pseudomonas aeruginosa</i> 339 <sup>b</sup>	>128	128 (38.64)	>128 (36.61)	128 (23.04)	>128 (23.52)	>128 (21.76)	>128 (16.13)	>128 (20.99)	>128 (27.64)	>128 (42.80)	nt
<i>Candida albicans</i> 54 <sup>b</sup>	>128	>128 (38.64)	>128 (36.61)	>128 (23.04)	>128 (23.52)	>128 (21.76)	>128 (16.13)	>128 (20.99)	>128 (27.64)	>128 (42.80)	nt
<i>Streptococcus mutans</i> <sup>c</sup>	36.3	17.8 (5.47)	13.5 (3.83)	14.8 (2.66)	20.6 (3.79)	76.8 (13.05)	18.9 (2.38)	23.6 (3.87)	52.4 (11.32)	nt	1.7 (0.85)
<i>Porphyromonas gingivalis</i> <sup>c</sup>	19.5	13.5 (4.07)	8.7 (2.48)	17.8 (1.56)	17.8 (3.27)	60.7 (10.32)	17.0 (2.14)	17.8 (2.92)	44.7 (9.65)	nt	0.6 (0.30)

Hematoxylin (**1**), brazilin (**2**), caffeic acid (**3**), methyl gallate (**4**), gallic acid (**5**), phloroglucinol (**6**), 4-hydroxycinnamic acid (**7**), and 5-methoxypsoralen (**8**).

<sup>a</sup> Reference antibiotics—PG: penicillin; CHX: chlorhexidine gluconate.

<sup>b</sup> Standard broth microdilution.

<sup>c</sup> *In vitro* growth inhibition assay performed in 96-well microtiter plates.

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