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Genetic network driven control of PHBV copolymer composition

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Abstract

We developed a detailed mathematical model describing the coupling between the molecular weight distribution dynamics of poly(3-hydroxybutyrate-co-3hydroxyvalerate) (PHBV) copolymer chains with those of hydroxybutyrate (HB) and hydroxyvalerate (HV) monomer formation. Sensitivity analysis of the model revealed that both the monomer composition and the molecular weight distribution of the copolymer chains are strongly affected by the ratio between the rates at which the two-monomer units are incorporated into the chains. This ratio depends on the relative HB and HV availability, which in turn is a function of the expression levels of genes encoding enzymes that catalyze monomer formation. Regulation of gene expression was accomplished through the aid of an artificial genetic network, the patterns of expression of which can be controlled by appropriately tuning the concentration of an extracellular inducer. Extensive simulations were used to study the effects of operating conditions and parameter uncertainties on the range of achievable copolymer compositions. Since the predicted conditions fell in the range of feasible bioprocessing manipulations, it is expected that such strategy could be successfully employed. Thus, the presented model constitutes a powerful tool for designing genetic networks that can drive the formation of PHBV copolymer structures with desirable characteristics.

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Keywords: Poly(3-hydroxybutyrate-co-3-hydroxyvalerate); PHBV; Control; Mathematical modeling; Genetic toggle; Population balance; Molecular weight distribution

1. Introduction

Polyhydroxyalkanoates (PHAs) represent a broad class of polyesters produced by many bacterial

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species under nutrient-limited conditions (Anderson and Dawes, 1990). Recombinant DNA technology has allowed expressing PHA genes in non-natural producers, thus enlarging the range of suitable microorganisms. PHAs are biodegradable in both aerobic and anaerobic conditions and in different environments (Mergaert et al., 1993, 1995; Tanio et al., 1982) and biocompatible with various human tissues and blood

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Nomenclature

$a_{\rm HB}(x_{\rm HB},t)$ active PHB homopolymer chains		
$a_{\rm HV}(x_{\rm HV},t)$ active PHV homopolymer chains		
$\hat{a}_{\text{HB}}(x_{\text{HB}})$ time invariant active PHB homopoly-		
mer chains		
$\hat{a}_{\rm HV}(x_{\rm HV})$	<i>(</i>) time invariant active PHV homopoly-	
mer chains		
$a_{0,\mathrm{HB}}$	total number of active PHB chains	
$a_{0,\mathrm{HV}}$	total number of active PHV chains	
$A(x_{\rm HB}, x)$	HV,t) active PHBV copolymer chains	
$\hat{A}(x_{\text{HB}}, x_{\text{HV}})$ time invariant active PHBV		
	copolymer chains	
A_0	total number of active PHBV chains	
$A_{1 \text{ HB}}$	HB concentration of active PHBV	
1,112	chains	
A_{1} HV	HV concentration of active PHBV	
1,111	chains	
C_{ACACCO}	A AcAcCoA intracellular concentration	
C_{AcCoA}	AcCoA intracellular concentration	
C_{Ace}	acetate intracellular concentration	
Сатр	ATP intracellular concentration	
C_{CoA}	co-enzyme A intracellular concentration	
Снв	HB intracellular concentration	
Сну	HV intracellular concentration	
Спарри	NADPH intracellular concentration	
CPrCoA	PrCoA intracellular concentration	
$C_{\rm Pro}$	propionate intracellular concentration	
CvalCoA	ValCoA intracellular concentration	
Снв	time invariant HB intracellular concen-	
- 110	tration	
Ĉнv	time invariant HV intracellular concen-	
- 11 v	tration	
<i>e</i> 1	Acs concentration	
e1 e2	prpE concentration	
Eur	time invariant elongation rate HB addi-	
	tion	
Euv	time invariant elongation rate HV addi-	
2111	tion	
F	PHBV copolymer mass fraction	
h_1	Pr s1con-clts repressor Hill cooperativ-	
1	ity	
ha	Ptrc-2-LacI repressor Hill cooperativity	
h_3	IPTG-LacI repressor Hill cooperativity	
$i_{\rm LLB}(r_{\rm LLB})$	t) inactive PHB homopolymer chains	
$i_{\rm HV}(x_{\rm HV},t)$ inactive PHV homopolymer chains		
I m m	x_{t} inactive PHRV copolymer chains	
(AHB, AHV, i) mactive i i ib v coporymer chams		

$I_{1,\mathrm{HB}}$	HB concentration of inactive PHBV
$I_{1,\mathrm{HV}}$	HV concentration of inactive PHBV
	chains
IPTG	IPTG inducer concentration
k _{Ace}	acetate Michealis–Menten constant
$k_{\rm Acs}$	Acs specific activity
$k_{\rm ATP}$	ATP Michealis–Menten constant
$k_{\rm CoA}$	CoA Michealis–Menten constant
$k_{\rm elo,HB}$	homopolymer PHB elongation rate con- stant
$k_{\rm elo,HV}$	homopolymer PHV elongation rate con-
<i>I</i>	homonolymor DUD initiation rate con
Kini,HB	stant
$k_{\rm ini,HV}$	homopolymer PHV initiation rate con- stant
<i>k</i> NADPH	NADPH Michealis–Menten constant
k _{nrnE}	prpE specific activity
$k_{\rm Pro}$	propionate Michealis-Menten constant
KIPTG	IPTG-LacI repressor dissociation con-
1110	stant
K TCA	TCA cycle flux constant
M	PHBV molecular weight distribution
k12	P ₁ s1con-cIts repressor dissociation con-
12	stant
<i>k</i> 21	Ptrc-2-LacI repressor dissociation con-
-21	stant
N_{Λ}	Avogadro's number
rup	time invariant PHB initiation rate
ruv	time invariant PHV initiation rate
$\frac{n_1}{n_1}$	LacI repressor concentration
p_1	cIts repressor concentration
$\frac{P^2}{R_{a1a}}$ IID	copolymer PHBV elongation rate HB
тею,нв	addition
$R_{\rm elo,HV}$	addition addition
R _{ter}	copolymer PHBV termination rate
t	Time
t _d	doubling time
tr _{HB}	time invariant PHB transition rate
tr _{HV}	time invariant PHV transition rate
T_1	first copolymer PHBV termination rate
-	constant
T_2	second copolymer PHBV termination rate constant

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