ELSEVIER

Contents lists available at ScienceDirect

## **Chemico-Biological Interactions**

journal homepage: www.elsevier.com/locate/chembioint



# Aldehyde dehydrogenase homologous folate enzymes: Evolutionary switch between cytoplasmic and mitochondrial localization



Natalia I. Krupenko <sup>a</sup>, Roger S. Holmes <sup>b</sup>, Yaroslav Tsybovsky <sup>c</sup>, Sergey A. Krupenko <sup>a,\*</sup>

- <sup>a</sup> Department of Nutrition, UNC-Chapel Hill, UNC Nutrition Research Institute, Kannapolis, NC 28081, United States
- <sup>b</sup> The Eskitis Institute for Drug Discovery and School of Natural Sciences, Griffith University, Nathan, 4111 Brisbane, Queensland, Australia
- <sup>c</sup> Department of Pharmacology, Case Western Reserve University, Cleveland, OH 44106, United States

#### ARTICLE INFO

Article history: Available online 27 December 2014

Keywords: Folate metabolism ALDH1L enzymes Mitochondria Aldehyde dehydrogenases Enzyme mechanism Evolution

#### ABSTRACT

Cytosolic and mitochondrial 10-formyltetrahydrofolate dehydrogenases are products of separate genes in vertebrates but only one such gene is present in invertebrates. There is a significant degree of sequence similarity between the two enzymes due to an apparent origin of the gene for the mitochondrial enzyme (ALDH1L2) from the duplication of the gene for the cytosolic enzyme (ALDH1L1). The primordial ALDH1L gene originated from a natural fusion of three unrelated genes, one of which was an aldehyde dehydrogenases. Such structural organization defined the catalytic mechanism of these enzymes, which is similar to that of aldehyde dehydrogenases. Here we report the analysis of ALDH1L1 and ALDH1L2 genes from different species and their phylogeny and evolution. We also performed sequence and structure comparison of ALDH1L enzymes possessing aldehyde dehydrogenase catalysis to those lacking this feature in an attempt to explain mechanistic differences between cytoplasmic ALDH1L1 and mitochondrial ALDH1L2 enzymes and to better understand their functional roles.

© 2014 Elsevier Ireland Ltd. All rights reserved.

#### 1. Introduction

Folate metabolism is crucial for several biosynthetic processes including de novo purine and thymidylate generation, synthesis of methionine from homocysteine and biosynthesis of glycine from serine [1,2]. It is also involved in the degradation of histidine and glycine and metabolism of betaine and dimethylglycine, which donate carbon groups into folate pool [1,2]. Enzymes involved in folate pathways are compartmentalized in the cell between cytoplasm and mitochondria [2]. Of note, several folate-dependent reactions take place in both compartments and are catalyzed by cytoplasmic and mitochondrial isozymes. Corresponding cytosolic and mitochondrial forms of folate enzymes are products of separate genes, which have likely arisen from gene duplication [3]. In recent years, presence of several folate enzymes in the nucleus has also been established [4,5]. This phenomenon, however, is the result of translocation of cytosolic enzymes under certain conditions to allow thymidylate generation directly at DNA replication sites [6,7]. Overall, folate-dependent nucleotide and methionine

Abbreviations: ALDH, aldehyde dehydrogenase; C<sub>t</sub>-FDH, carboxyl terminal domain of 10-formyltetrahydrofolate dehydrogenase; THF, tetrahydrofolate.

E-mail address: sergey\_krupenko@unc.edu (S.A. Krupenko).

biosynthesis takes place outside of mitochondria and it has been proposed that the mitochondrial folate metabolism plays a supportive role providing cytoplasmic folate metabolism with additional one-carbon groups derived from glycine degradation and betaine/dimethylglycine conversion [8,9].

One of the folate reactions duplicated between cytosol and mitochondria is the conversion of 10-formyltetrahydrofolate to tetrahydrofolate (THF) and CO<sub>2</sub> [10]. This reaction is catalyzed by two similar enzymes, cytosolic and mitochondrial 10-formyl-THF dehydrogenases, which are products of separate genes [11]. While the precise roles of these enzymes are not clear at present, the cytosolic isoform is likely to serve as a regulator of the overall folate metabolism since it irreversibly removes one-carbon groups from folate pool thus restricting the capacity of folate-dependent biosynthetic reactions [12,13]. In agreement with this regulatory function, *ALDH1L1* is ubiquitously silenced in human cancers apparently as a mechanism favoring limitless proliferation [14–16]. The function of ALDH1L2 enzyme is even less clear, but it could be involved in the production of formate instead of CO<sub>2</sub> [17].

The cloning of *ALDH1L1* gene in 1991 immediately revealed the fact that it is the product of natural fusion of three unrelated genes [18]. One of these genes was an aldehyde dehydrogenase (ALDH) and another was similar to two 10-formyl-THF utilizing enzymes, GARFT and FMT [19]. Such gene organization results in the enzyme

<sup>\*</sup> Corresponding author. Tel.: +1 704 250 5053.

with two distinct catalytic domains, the amino-terminal folate-binding domain and carboxyl-terminal ALDH domain [20,21]. These domains are connected by a short (about a 100 amino acid residues) linker, which is not a part of either domain. We later demonstrated that the linker domain is a structural and functional homolog of acyl carrier proteins [22]. The characteristic feature of these proteins, the 4'-phosphopantetheinyl prosthetic group, allows the transfer of the intermediate of the ALDH1L1 catalytic reaction from folate binding site to the ALDH catalytic center [19,23].

Compared to canonical *ALDHs*, which are ancient genes and present in all kingdoms of life, the *ALDH1L1* gene appeared later in evolution: it is not found in plant, bacteria or yeast [3]. Our previous phylogenetic analysis pointed to the conclusion that mitochondrial ALDH1L2 has emerged after cytosolic ALDH1L1 and the appearance of the former was traced to bony fish [3]. The annotation of additional genomes in recent years indicated the necessity to re-evaluate the evolutionary relationship between *ALDH1L1* and *ALDH1L2*. Here we performed the extended phylogenetic analysis of ALDH1L1 and ALDH1L2 enzymes and compared structures of the enzymes to understand differences in their catalytic abilities.

#### 2. Materials and methods

#### 2.1. ALDH1L1 and ALDH1L2 gene and protein identification

ALDH1L1 and ALDH1L2 sequences for representative vertebrate and invertebrate species were retrieved from ExPASy (http://www.expasy.org) [24] and NCBI (http://blast.ncbi.nlm.nih.gov/Blast.cgi) databases using human (Homo sapiens) [11] and zebrafish (Danio rerio) [3] ALDH1L1 and ALDH1L2 sequences to seed

searches. Identification of these genes was based on high predictive scores (>850) and sequence coverage (>98%) for ALDH1L-like protein sequences listed by NCBI, in each case (Table 1). BLAT searches were performed using relevant ALDH1L1 and ALDH1L2 protein sequences to confirm the presence or absence of these genes among the species examined using the UCSC Genome Browser [25]. Predicted gene structures, gene locations and ALDH1L1 and ALDH1L2 amino acid sequences were obtained for each protein identified (Table 1). Prediction of the ALDH1L-like protein N-terminal sequence that may serve as a mitochondrial targeting peptide, and the cleavage site for this peptide, was undertaken using MITOPROT [26].

#### 2.2. Amino acid sequence alignments and phylogenetic analyses

Vertebrate and invertebrate ALDH1L-like sequences were subjected to phylogenetic analysis using the <a href="http://www.phylogeny.fr/">http://www.phylogeny.fr/</a>
portal to enable alignment (MUSCLE), curation (Gblocks), phylogeny (PhyML) and tree rendering (TreeDyn) to reconstruct phylogenetic relationships [27]. Vertebrate sequences were identified as members of the ALDH1L1 (cytosolic) or ALDH1L2 (mitochondrial) groups, whereas invertebrate sequences were identified as a members of a single group, designated as ALDH1L1.

#### 2.3. Homology modeling

Homology models of the C-terminal domains of human mtFDH and zebrafish cytosolic FDH (zFDH) were generated using the SWISS-MODEL server as described earlier [17].

Table 1
Invertebrate ALDH1L1 and vertebrate ALDH1L1 and ALDH1L2 genes and proteins.

Animal	Species	Gene	RefSeq ID <sup>a</sup> Prediction	GenBank ID	<sup>b</sup> Exons (strand)	Gene size (bp)	Amino acids	Localization	Leader peptide
Human	Homo sapiens	ALDH1L1	NM_012190.3	AF052732	22 (-ve)	57,186	902	Cytosol	NA
Human	Homo sapiens	ALDH1L2	NM_001034173.3	BC103934	23 (-ve)	60,010	923	Mitochondria	120
Mouse	Mus musculus	Aldh1l1	NM_027406.1	BC024055	22 (+ve)	41,715	902	Cytosol	NA
Mouse	Mus musculus	Aldh1l2	NM_153543.2	BC034531	23 (-ve)	43,433	923	Mitochondria	120
Chicken	Gallus gallus	ALDH1L2	XP_416314.2 <sup>a</sup>	NA	23 (+ve)	27,506	922	Mitochondria	120
Lizard	Anolis carolinensis	ALDH1L2	XP_003220962.1 <sup>a</sup>	NA	23 (-ve)	30,373	924	Mitochondria	120
Frog	Xenopus tropicalis	ALDH1L1	NM_001011027.1	BC082822	22 (+ve)	16,496	902	Cytosol	NA
Frog	Xenopus tropicalis	ALDH1L2	XP_002938116.1 <sup>a</sup>	NA	23 (+ve)	27,966	924	Mitochondria	133
Zebra fish	Danio rerio	ALDH1L1	NM_001198772.1	NA	22 (+ve)	27,616	904	Cytosol	NA
Zebra fish	Danio rerio	ALDH1L2	XP_002661418.2 <sup>a</sup>	NA	22 (+ve)	19,873	923	Mitochondria	144
Shark	Callorhinchus milii	ALDH1L1	XP_007888551.1 <sup>a</sup>	NA	22 (-ve)	15,784	901	Cytosol	NA
Shark	Callorhinchus milii	ALDH1L2	XP_007907882.1 <sup>a</sup>	JW862169	23 (+ve)	27,862	922	Mitochondria	119
Sea squirt	Ciona intestinalis	ALDH1L1a	XP_002130073.1	NA	17 (+ve)	7083	898	Cytosol	NA
Sea squirt	Ciona intestinalis	ALDH1L1b	XP_002130073.2	NA	18 (+ve)	7339	921	Mitochondria	112
Sea urchin	Strongylocentrotus purpuratus	ALDH1L1	XP_784777.3 <sup>a</sup>	NA	22 (-ve)	25,243	927	Mitochondria	118
Sea hare	Aplysia californica	ALDH1L1	XP_005090853.1ª	NA	23 (+ve)	19,195	900	Cytosol	NA
Trichoplax	Trichoplax adhaerens	ALDH1L1	XP_002111368.1 <sup>a</sup>	NA	NA	NA	921	Mitochondria	118
Worm	Caenorhabditis elegans	ALDH1L1	NM_069653.6	NA	7 (+ve)	3128	908	Cytosol	NA
Round worm	Caenorhabditis brenneri	ALDH1L1	GL379933.1 <sup>a</sup>	EGT36278.1	7 (-ve)	3102	908	Cytosol	NA
Fruit fly	Drosophila melanogaster	ALDH1L1	NP_610107.1	CG8665	2 (+ve)	3149	913	Cytosol	NA
Mosquito	Anopheles gambiae	ALDH1L1	XP_318614.3 <sup>a</sup>	NA	2 (-ve)	2820	916	Cytosol	NA
House fly	Musca domestica	ALDH1L1	XP_005181895.1a	NA	NA	NA	912	Cytosol	NA
Bee	Apis mellifera	ALDH1L1	XM_623795 <sup>a</sup>	NA	6 (-ve)	3404	900	Cytosol	NA
Butterfly	Danaus plexippus	ALDH1L1	NA	EHJ79154.1	NA	NA	927	Cytosol	NA
Water flea	Daphnia pulex	ALDH1L1	NA	EFX71787.1	NA	NA	924	Cytosol	NA
Wasp	Nasonia vitripennis	ALDH1L1	XP_001602871.1 <sup>a</sup>	NA	NA	NA	902	Cytosol	NA
Beetle	Tribolium castaneum	ALDH1L1	XP_969916.1 <sup>a</sup>	NA	NA	NA	915	Cytosol	NA
Ant	Camponotus floridanus	ALDH1L1	ENF71966.1 <sup>a</sup>	NA	NA	NA	900	Cytosol	NA
Termite	Zootermopsis nevadensis	ALDH1L1	KDR07781.1 <sup>a</sup>	NA	NA	NA	922	Mitochondria	121

NA, not available; "bp" refers to base pairs of nucleotide sequence; the length of the predicted mitochondrial leader sequence is shown. RefSeq refers to the NCBI reference sequence;

<sup>&</sup>lt;sup>a</sup> Predicted NCBI sequence.

<sup>&</sup>lt;sup>b</sup> The number of translatable exons is shown.

### Download English Version:

# https://daneshyari.com/en/article/2580217

Download Persian Version:

https://daneshyari.com/article/2580217

<u>Daneshyari.com</u>