



Tolerability and pharmacokinetics of oxaloacetate 100 mg capsules in Alzheimer's subjects



Russell H. Swerdlow^{a,b,c,d,*}, Rebecca Bothwell^a, Lewis Hutfles^a, Jeffrey M. Burns^{a,b,d}, Gregory A. Reed^e

^a University of Kansas Alzheimer's Disease Center, Kansas City, KS, USA

^b Department of Neurology, University of Kansas School of Medicine, Kansas City, KS, USA

^c Department of Biochemistry and Molecular Biology, University of Kansas School of Medicine, Kansas City, KS, USA

^d Department of Molecular and Integrative Physiology, University of Kansas School of Medicine, Kansas City, KS, USA

^e Department of Pharmacology, Toxicology, and Therapeutics, University of Kansas School of Medicine, Kansas City, KS, USA

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ABSTRACT

Bioenergetics and bioenergetic-related functions are altered in Alzheimer's disease (AD) subjects. These alterations represent therapeutic targets and provide an underlying rationale for modifying brain bioenergetics in AD-affected persons. Preclinical studies in cultured cells and mice found that administering oxaloacetate (OAA), a Krebs cycle and gluconeogenesis intermediate, enhanced bioenergetic fluxes and upregulated some brain bioenergetic infrastructure-related parameters. We therefore conducted a study to provide initial data on the tolerability and pharmacokinetics of OAA in AD subjects. Six AD subjects received OAA 100 mg capsules twice a day for one month. The intervention was well-tolerated. Blood level measurements following ingestion of a 100 mg OAA capsule showed modest increases in OAA concentrations, but pharmacokinetic analyses were complicated by relatively high amounts of endogenous OAA. We conclude that OAA 100 mg capsules twice per day for one month are safe in AD subjects but do not result in a consistent and clear increase in the OAA blood level, thus necessitating future clinical studies to evaluate higher doses.

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1. Introduction

Alzheimer's disease (AD) is clinically characterized by cognitive decline [1]. While hypotheses postulate its potential causes and propose various therapeutic targets, no clearly effective disease-modifying interventions are currently recognized.

The single greatest AD risk factor is advancing age. Brain bioenergetic function and mitochondrial integrity decline with advancing age and to a further extent when AD is present [2]. Energy metabolism-associated changes in AD include decreased glucose utilization, as indicated by fluoro-deoxyglucose positron emission tomography (FDG PET) studies that reliably reveal early and neuroanatomically predictable

hypometabolic brain regions [3–5]. Activities of several mitochondria-localized enzymes, including enzymes of the Krebs cycle and the respiratory chain, are also reduced in AD subject brains and in some cases even peripheral tissues [2]. Some brain regions show an overall reduction in the number of normal-appearing mitochondria, apparently increased mitochondrial debris in autophagosomes, and low levels of the mitochondrial biogenesis-promoting peroxisome proliferator-activated receptor gamma coactivator (PGC1 α) protein [6,7]. Some AD investigators believe energy metabolism functional and structural changes may contribute to the progression of disease and perhaps even initiate it, and constitute reasonable therapeutic targets [8,9].

We previously reported changes in bioenergetic fluxes and infrastructure when cells or animals are exposed to various energy metabolism pathway intermediates. One intermediate we evaluated is oxaloacetate (OAA), a dicarboxylic acid found in Krebs cycle and gluconeogenesis fluxes. Administering OAA to cultured neuronal SH-SY5Y cells enhances glycolysis and respiratory fluxes, increases PGC1 α mRNA and protein, and increases mRNA and protein levels of a mitochondrial DNA (mtDNA)-encoded cytochrome oxidase (COX) subunit [10]. The brains of mice that received a two-week course of intraperitoneal (IP) OAA showed increased levels of PGC1 α mRNA, an increase in the nuclear to cytosolic PGC1 α protein ratio, and higher amounts of the COX subunit 4 protein [11]. Compared to saline injected

Abbreviations: AD, Alzheimer's disease; ADASCog, Alzheimer's Disease Assessment Scale-Cognitive subset; AUC, area under the curve; CDR, Clinical Dementia Rating; CBC, complete blood count; COX, cytochrome oxidase; FDG PET, fluoro-deoxyglucose positron emission tomography; HOMA-IR, homeostatic model assessment of insulin resistance; IP, intraperitoneal; KU ADC, University of Kansas Alzheimer's Disease Center; LC-MS/MS, liquid chromatography–tandem mass spectrometry; LFT, liver function tests; MMSE, mini-mental state exam; mtDNA, mitochondrial DNA; OAA, oxaloacetate; PGC1 α , peroxisome proliferator-activated receptor gamma coactivator; PK, pharmacokinetic.

* Corresponding author at: Landon Center on Aging, MS 2012, 3901 Rainbow Blvd, Kansas City, KS 66160, USA.

E-mail address: rswerdlow@kumc.edu (R.H. Swerdlow).

mice, the brains of the OAA-treated mice also showed higher hippocampal neurogenesis activity and changes suggesting enhanced brain insulin signaling and reduced neuroinflammation [11]. For these reasons we want to determine the effects of OAA on persons with AD.

2. Methods

This study was approved by the Kansas University Medical Center Human Subjects Committee and informed consent was obtained for all subjects. We recruited six AD subjects from the University of Kansas Alzheimer's Disease Center (KU ADC) Clinical Core cohort; APOE genotype status and clinical dementia rating scale (CDR) scores are independently acquired for KU ADC cohort members. Subjects met the McKhann et al. AD criteria [12], had CDR scores of 0.5 or 1, and had mini-mental state exam (MMSE) scores between 15 and 28. Subjects with a syndromic diagnosis of mild cognitive impairment (as opposed to a full AD diagnosis), as well as subjects with diabetes, were excluded. Each subject had a study partner who was already engaged in their daily social and medical care.

OAA capsules were obtained from Terra Biological LLC (San Diego, California). Terra Biological LLC markets OAA capsules produced using good manufacturing practice procedures, and OAA batches used to prepare these capsules are tested to ensure the integrity of the OAA. The capsules contained 100 mg OAA and 150 mg of ascorbic acid. Because 100 mg of OAA was the lowest amount that could be administered at a given time, we predicted OAA would elevate the blood level for a limited period, and safety was a major focus of this study OAA dosing was set at 100 mg twice per day.

Participation in the study required two study visits, an initial visit and a final study visit 4 weeks later (Fig. 1). For both visits a 36 item review of symptoms inventory was also completed by the subject and the study partner. During the initial visit the first 100 mg OAA capsule was administered and subject assessments were performed. At the conclusion of the initial visit a bottle containing 60 OAA capsules was provided with instructions for the subject to take one capsule each morning and one capsule each evening. Approximately one month later the final study visit was conducted, at which time subjects were assessed at the time of taking their final 100 mg OAA capsule. The OAA bottles were collected and any remaining capsules were counted to assess compliance.

For each visit subjects presented at 8 AM to the Clinical Trials Unit of the Kansas University Medical Center having fasted since midnight. The

subjects were weighed and vital signs determined. A heparin lock was inserted, and blood was obtained to determine the subject's complete blood count (CBC), electrolytes, liver function tests (LFTs), plasma amino acids, and insulin level. A 3–5 ml baseline blood sample for pharmacokinetic (PK) testing was then obtained, and a 100 mg OAA capsule was orally administered with 200 ml of water. Additional 3–5 ml PK blood samples were obtained at 30, 45, 60, 75, 90, 105, 120, 135, 150, and 240 min after dosing. The PK blood samples were immediately placed on ice and processed into plasma within 10 min of acquisition. Aliquots of plasma (0.5 ml each) were frozen immediately and stored at -80°C until analysis. During the time lag between the 150 and 240 min PK phlebotomies, a psychometrician administered an MMSE and Alzheimer's Disease Assessment Scale–Cognitive subscale (ADASCog) test to each subject. The ADASCog version in our study is the version used by the Alzheimer's Disease Cooperative Study Group, which scores 11 items. Higher scores indicate worse performance and the highest maximum possible score is 70 points.

Quantitation of OAA in plasma was performed using 3 different approaches. We used a commercially available coupled enzyme method assay kit (Sigma-Aldrich, St. Louis, MO) that utilizes a fluorescence detection protocol to measure OAA concentrations. This kit also provides an absorption-based protocol for OAA measurements, and the absorption-based approach was used as well. Finally, because OAA undergoes some degree of decarboxylation to pyruvate when it is placed in solution, we adapted and applied a liquid chromatography–tandem mass spectrometry (LC–MS/MS)-based assay for OAA quantitation and to measure plasma pyruvate concentrations [13].

The glucose and insulin levels from each visit were used to calculate the homeostatic model assessment of insulin resistance (HOMA-IR) values for each subject using the following equation: $(\text{glucose in mg/dl} \times \text{insulin in mcu/ml})/405$. Initial and final visit HOMA-IR, weight, plasma amino acid, and cognitive score values were compared using a paired t-test approach. CBC, electrolyte, and LFT studies were analyzed for the appearance of clinical abnormalities.

3. Results

Subject characteristics are shown in Table 1. Mean age (with standard deviation) was 76.2 ± 8.2 . Only one subject lacked an APOE4 allele. All of the subjects completed the study. During the course of the study no adverse events, treatment-induced symptoms, or clinically significant changes in safety labs were observed. Compliance estimates for the six subjects ranged from 76 to 100% (Table 2).

Weight, fasting glucose, fasting insulin, and HOMA-IR values for subjects varied between visits, but as a group no consistent changes in weight or HOMA-IR were observed during the course of the study (Table 3). Although there was not a statistically significant reduction in post-treatment fasting glucose levels, post-treatment fasting glucose levels were slightly lower than pre-treatment levels in 5 of the 6 subjects. MMSE and ADASCog values also varied between visits, but as a group no consistent changes in the MMSE or ADASCog scores were observed during the course of the study (Table 3). The average visit 1 and 2 MMSE scores (with standard deviations) were, respectively, 21.2 ± 4.7 and 19.5 ± 5.9 . The average visit 1 and visit 2 ADASCog scores were 21.5 ± 5.1 and 23.7 ± 6.6 (Table 3).

Table 1
Subject characteristics.

Subject	Age	Sex	APOE	CDR
1	64	M	4/4	0.5
2	68	M	3/4	1
3	78	M	3/3	1
4	83	F	3/4	0.5
5	82	F	3/4	1
6	82	M	3/4	1

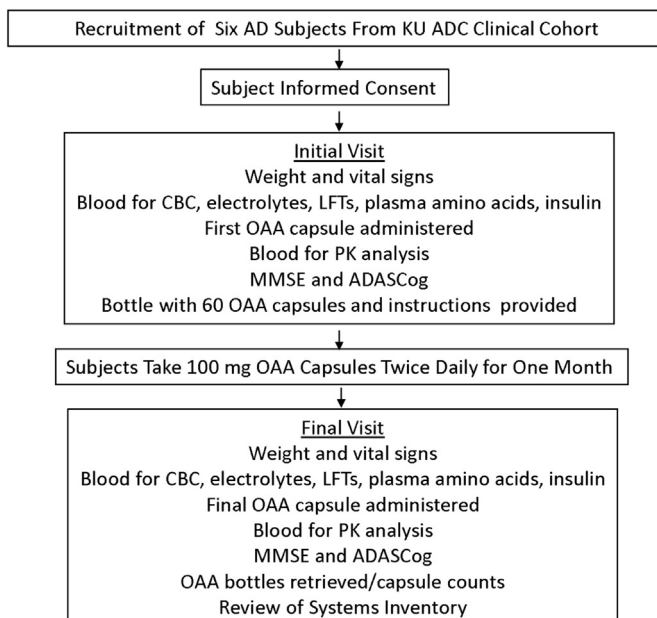


Fig. 1. Pharmacokinetics of Oxaloacetate (POX) study organization.

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