- Macy E, Mangat R, Bruchette RJ. Penicillin skin testing in advance of need: multiyear follow-up in 568 test result-negative subjects exposed to oral penicillins. J Allergy Clin Immunol 2003;111:1111-5.
- Bittner A, Greenberger PA. Incidence of resensitization after tolerating penicillin treatment in penicillin-allergic patients. Allergy Asthma Proc 2004;25:161-4.
- Kelkar PS, Li JT-C. Cephalosporin allergy. N Engl J Med 2001;345: 804-9.
- Greenberger PA. Utility of penicillin major and minor determinants for identification of allergic reactions to cephalosporins. J Allergy Clin Immunol 2005;115(suppl):S182.
- Romano A, Guenant-Rodriquez R-M, Viola M, Pettinato R, Gueant J-L. Cross-reactivity and tolerability of cephalosporins in patients with immediate hypersensitivity to penicillins. Ann Intern Med 2004;141: 16-22.
- Gruchalla RS. Drug allergy. J Allergy Clin Immunol 2003;111(suppl): S548-59.
- Shear NH, Spielberg SP, Grant DM, Tang BK, Kalow W. Differences in metabolism of sulfonamides predisposing to idiosyncratic toxicity. Ann Intern Med 1986;105:179-84.
- Strom BL, Schinnar R, Apter AJ, Margolis DJ, Lautenbach E, Hennessy S, et al. Absence of cross-reactivity between sulfonamide antibiotics and sulfonamide nonantibiotics. N Engl J Med 2003;349: 1628-35.
- 36. Greenberger PA. Drug allergy part B: allergic reactions to individual drugs: low molecular weight. In: Grammer LC, Greenberger PA, editors. Patterson's allergic diseases. 6th ed. Philadelphia: Lippincott, Williams & Wilkins; 2002. p. 335-59.
- Gollapudi RR, Teirstein PS, Stevenson DD, Simon RA. Aspirin sensitivity: implications for patients with coronary artery disease. JAMA 2004; 292:3017-23.
- Mathison DA, Lumry WR, Stevenson DD, Curd JG. Aspirin in chronic urticaria and/or angioedema: studies of sensitivity and desensitization. J Allergy Clin Immunol 1982;69:135.
- Lam N-S, Yang Y-H, Wang L-C, Lin Y-T, Chiang B-L. Clinical characteristics of childhood erythema multiforme, Stevens-Johnson syndrome and toxic epidermal necrolysis in Taiwanese children. J Microbiol Immunol Infect 2004;37:366-70.
- Rzany B, Correia O, Kelly J, Naldi L, Auquier A, Stern R. Risk of Stevens-Johnson syndrome and toxic epidermal necrolysis during first weeks of epileptic therapy: a case-control study. Lancet 1999;353: 2190-4.
- Nassif A, Bensussan A, Boumsell L, Deniaud A, Moslehi H, Wolkenstein P, et al. Toxic epidermal necrolysis: effector cells are drug-specific cytotoxic T cells. J Allergy Clin Immunol 2004;114:1209-15.
- Viard I, Wehrli P, Bullani R, Schneider P, Holler N, Salomon D, et al. Inhibition of toxic epidermal necrolysis by blockade of CD95 with human intravenous immunoglobulin. Science 1998;282:490-3.
- 43. Abe R, Shimizu T, Shibaki A, Nakamura H, Watanabe H, Shimizu H. Toxic epidermal necrolysis and Stevens-Johnson syndrome are induced by soluble Fas ligand. Am J Pathol 2003;162:1515-20.

- Bachot N, Revuz J, Roujeau J-C. Intravenous immunoglobulin treatment for Stevens-Johnson syndrome and toxic epidermal necrolysis. Arch Dermatol 2003;139:33-6.
- Allam J-P, Paus T, Reichel C, Bieber T, Novak N. DRESS syndrome associated with carbamazepine and phenytoin. Eur J Dermatol 2004;14: 339-42.
- 46. Tripathi A, Peters NT, Patterson R. Erythema multiforme, Stevens-Johnson syndrome, and toxic epidermal necrolysis. In: Grammer LC, Greenberger PA, editors. Patterson's allergic diseases. 6th ed. Philadelphia: Lippincott, Williams & Wilkins; 2002. p. 289-94.
- Tripathi A, Ditto AM, Grammer LC, Greenberger PA, McGrath KG, Zeiss CR, et al. Corticosteroid therapy in an additional 13 cases of Stevens-Johnson syndrome: a total series of 67 cases. Allergy Asthma Proc 2000;21:101-5.
- Levine BB, Zolov DM. Prediction of penicillin allergy by immunological tests. J Allergy 1969;43:231-44.
- Saxon A, Adelman DC, Patel A, Hajdu R, Calandra GB. Imipenem cross-reactivity with penicillin in humans. J Allergy Clin Immunol 1988;82:213-7.
- Canaday BR. Anticonvulsant cross-sensitivity. Am J Health Syst Pharm 1997;54:2616-7.
- Brown NJ, Snowden M, Griffin MR. Recurrent angiotensin-converting enzyme inhibitor-associated angioedema. JAMA 1997;278:232-3.
- Slater EE, Merrill DD, Guess HA, Roylance PJ, Cooper WD, Inman WHW, et al. Clinical profile of angioedema associated with angiotensin converting-enzyme inhibition. JAMA 1988;260:967-70.
- Samter M, Beers RF Jr. Intolerance to aspirin. Clinical studies and consideration of its pathogenesis. Ann Intern Med 1968;68:975-83.
- Adkinson NF Jr, Thompson WL, Maddrey WC, Lichtenstein LM. Routine use of penicillin skin testing on an inpatient service. N Engl J Med 1971;285:22-4.
- Stevenson DD, Simon RA. Lack of cross-reactivity between rofecoxib and aspirin in aspirin-sensitive patients with asthma. J Allergy Clin Immunol 2001;108:47-51.
- Szczeklik A, Nizankowska E, Bochenek G, Nagraba K, Mejza F, Swierczynska M. Safety of a specific COX-2 inhibitor in aspirin-induced asthma. Clin Exp Allergy 2001;31:219-25.
- Woessner KM, Simon RA, Stevenson DD. Safety of high-dose rofecoxib in patients with aspirin-exacerbated respiratory disease. Ann Allergy Asthma Immunol 2004;93:339-44.
- 58. Quiralte J, Delgado J, Saenz de San Pedro B, Lopez-Pascual E, Nieto MA, Ortega N, et al. Safety of the new selective cyclooxygenase type 2 inhibitors rofecoxib and celecoxib in patients with anaphylactoid reactions to nonsteroidal anti-inflammatory drugs. Ann Allergy Asthma Immunol 2004;93:360-4.
- Cicardi M, Zingale LC, Bergamaschini L, Agostoni A. Angioedema associated with angiotensin-converting enzyme inhibitor use. Arch Intern Med 2004;164:910-3.
- Greenberger PA, Patterson R. The prevention of immediate generalized reactions to radiocontrast media in high risk patients. J Allergy Clin Immunol 1991;87:867-72.

9. Food allergy

Scott H. Sicherer, MD, and Hugh A. Sampson, MD New York, NY

This activity is available for CME credit. See page 5A for important information.

Food allergy, defined as an adverse immune response to food proteins, affects as many as 6% of young children and 3% to 4% of adults. Food-induced allergic reactions are responsible for a variety of symptoms involving the skin, gastrointestinal tract, and respiratory tract and might be caused by IgEmediated and non–IgE-mediated (cellular) mechanisms. Our understanding of how food allergy represents an abrogation of normal oral tolerance is evolving. Although any food can provoke a reaction, relatively few foods are responsible for the vast majority of significant food-induced allergic reactions: milk, egg, peanuts, tree nuts, fish, and shellfish. A systematic approach to diagnosis includes a careful history, followed by laboratory studies, elimination diets, and often food challenges to confirm a diagnosis. Many food allergens have been characterized at a molecular level, which has increased our understanding of the immunopathogenesis of food allergy and might soon lead to novel diagnostic and therapeutic approaches. Currently, management of food allergies consists

of educating the patient to avoid ingesting the responsible allergen and to initiate therapy in case of an unintended ingestion. (J Allergy Clin Immunol 2006;117:S470-5.)

Key words: Food allergy, food hypersensitivity, oral tolerance, gastrointestinal food hypersensitivity, food allergens, anaphylaxis

Approximately 20% of the population alters their diet for a perceived adverse reaction to food, the cause of which might include a verifiable adverse immune response to a food protein (eg, food allergy), a host-specific metabolic disorder (eg, lactose intolerance), a response to a pharmacologically active (eg, caffeine) or toxic (eg, food poisoning) food component, or nonreproducible adverse reactions, such as food aversions (Table I).¹⁻⁴ Foodinduced allergic disorders result from immunologic pathways that include activation of effector cells through food-specific IgE antibodies, cell-mediated reactions resulting in subacute or chronic inflammation, or combined pathways. Approximately 6% of young children and 3.7% of adults in the United States have a food allergy.^{1,5} In young children the most common causal foods are cow's milk (2.5%), egg (1.3%), peanut (0.8%), wheat (approximately 0.4%), soy (approximately 0.4%), tree nuts (0.2%), fish (0.1%), and shellfish (0.1%). Early childhood allergies to milk, egg, soy, and wheat usually resolve by school age (approximately 80%).⁶ Although peanut, tree nut, and seafood allergies are generally considered permanent, 20% of young children with peanut allergy experience resolution by age 5 years (recurrence is also possible).^{7,8} Adults are therefore more likely to have allergies to shell fish (2%), peanut (0.6%), tree nuts (0.5%), and fish (0.4%). Reactions to fruits and vegetables are common (approximately 5%) but usually not severe. Allergy to seeds (eg, sesame) is being increasingly reported.⁹ Genetic risk factors include a family history of atopic disorders, but environmental factors modulate the expression of food allergy, as evidenced by a recent doubling of the rate of peanut allergy in children.¹⁰

PATHOGENESIS

Food allergy might result from a breach in oral tolerance to foods while they are being ingested (class 1 food allergy) or might result from sensitization to allergens apart from their exposure to the gastrointestinal tract, recognized instead during respiratory exposure (class 2 food allergy).^{11,12} Class 1 food allergy typically occurs to food proteins that are generally stable to digestion that are encountered by infants or children during a presumed

doi:10.1016/j.jaci.2005.05.048

Abbreviation used SPT: Skin prick test

window of immunologic immaturity. In contrast, class 2 food allergy is typically the result of sensitization to labile proteins encountered through the respiratory route, such as pollens resulting in IgE antibodies that recognize homologous epitopes on food proteins of plant origin (eg, pollenfood related syndrome). Murine studies¹³ and evidence from human epidemiologic studies¹⁴ indicate that class 1 allergens, such as egg and peanut, might evade oral tolerance by initial sensitizing exposure through the skin.

Gut barrier

The gastrointestinal mucosal barrier is a complex physical (mucus, epithelial cell tight junctions, acid, and enzymes) and immunologic structure.¹² Abrogation of the barrier might promote food allergy; studies neutralizing stomach pH showed increased ability to promote allergic sensitization.¹⁵ Similarly, developmental immaturity of components of the gut barrier (enzymatic activity and sIgA) might account for the increased prevalence of food allergy in infancy. However, a small amount of ingested food antigens is normally absorbed and transported throughout the body in an immunologically intact form, and oral tolerance prevails.^{1,12}

Oral tolerance induction

Antigen-presenting cells, especially intestinal epithelial cells and dendritic cells, and regulatory T cells play a central role in oral tolerance.^{12,16} Five regulatory T cells have been identified in conjunction with intestinal immunity: T_H3 cells, a population of CD4⁺ cells that secrete TGF- β ; T_R1 cells, CD4⁺ cells that secrete IL-10; CD4⁺CD25⁺ regulatory T cells; CD8⁺ suppressor T cells; and $\gamma\delta$ T cells. Intestinal epithelial cells process luminal antigen and present it to T cells on an MHC class II complex but lack a second signal, thus suggesting their potential to play a role in tolerance induction.¹² Dendritic cells residing within the lamina propria and noninflammatory environment of Peyer's patches express IL-10 and IL-4, which favor the generation of tolerance.^{12,17} Properties of antigens, dose, and frequency of exposure influence tolerance induction. High-dose tolerance involves deletion of effector T cells, whereas low-dose tolerance is mediated by activation of regulatory T cells with suppressor functions.12

Commensal gut flora might also influence the mucosal immune response. Gut flora is largely established in the first 24 hours after birth and is dependent on maternal flora and local environment. Studies feeding lactating mothers and their offspring *Lactobacillus GG* suggest that probiotics might be of benefit in preventing atopic dermatitis,¹⁸ possibly by enhancing a T_H1 cytokine response (IFN- γ),¹⁹ but whether they will be useful for preventing food allergy remains to be demonstrated.

The Elliot and Roslyn Jaffe Food Allergy Institute, Division of Allergy and Immunology, Department of Pediatrics, Mount Sinai School of Medicine, New York.

Reprint requests: Scott H. Sicherer, MD, Division of Allergy/Immunology, Mount Sinai Hospital, Box 1198, One Gustave L. Levy Place, New York, NY 10029-6574. E-mail: scott.sicherer@mssm.edu.

^{0091-6749/\$32.00}

^{© 2006} American Academy of Allergy, Asthma and Immunology

Download English Version:

https://daneshyari.com/en/article/3202649

Download Persian Version:

https://daneshyari.com/article/3202649

Daneshyari.com