

University School of Medicine, NYU Langone Medical Center, New York, New York, USA; ⁵Division of Epidemiology and Community Health, University of Minnesota, Minnesota, USA; ⁶Division of Molecular Genetic Epidemiology, German Cancer Research Center, Heidelberg, Germany; ⁷Department of Internal Medicine and Medical Specialties, University of Genoa, Italy; ⁸IRCCS AOU San Martino-IST, Genoa, Italy; ⁹Department of Chronic Disease Epidemiology, Yale School of Public Health, Yale Cancer Center, New Haven, Connecticut, USA; ¹⁰Department of Dermatology, Leiden University Medical Center, Leiden, The Netherlands; ¹¹Melanoma Unit, Dermatology Department, Hospital Clinic Barcelona, University of Barcelona, CIBER de Enfermedades Raras, Spain; ¹²Department of Cancer Epidemiology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, Florida, USA; ¹³Skin Research Department, POLA Chemical Industries, Yokohama, Japan; ¹⁴Department of medical oncology and hematology, Fundación Investigación Clínico de Valencia Instituto de Investigación Sanitaria- INCLIVA, Valencia, Spain; ¹⁵Division of Cancer Epidemiology and Genetics, National Cancer Institute, NIH, Bethesda, Maryland, USA; ¹⁶Department of Dermatology, University of L'Aquila, L'Aquila, Italy; ¹⁷NHS Forth Valley, UK; ¹⁸First Department of Dermatology, Andreas Sygros Hospital, Medical School, National and Kapodistrian University of Athens, Athens, Greece; ¹⁹Department of Pathology, Oslo University Hospital, Oslo, Norway; ²⁰Department of Basic Medical Sciences, Neuroscience and Sense Organs, University of Bari, Bari, Italy; ²¹International Prevention Research Institute, Lyon, France; ²²Department of Epidemiology, Richard M. Fairbanks School of Public Health, Melvin & Bren Simon Cancer Center, Indiana University, Indianapolis, Indiana, USA; ²³School of Epidemiology, Public Health and Preventive Medicine, University of Ottawa, Ottawa, Canada; and ²⁴Department of Social and Environmental

Health Research, London School of Hygiene & Tropical Medicine, London, UK

*Corresponding author e-mail: sara.raimondi@ieo.it

SUPPLEMENTARY MATERIAL

Supplementary material is linked to the online version of the paper at www.jidonline.org, and at <http://dx.doi.org/10.1016/j.jid.2016.05.099>.

REFERENCES

- Beaumont KA, Shekar SN, Newton RA, James MR, Stow JL, Duffy DL, et al. Receptor function, dominant negative activity and phenotype correlations for MC1R variant alleles. *Hum Mol Genet* 2007;16:2249–60.
- Garcia-Borrón JC, Sanchez-Laorden BL, Jimenez-Cervantes C. Melanocortin-1 receptor structure and functional regulation. *Pigment Cell Res* 2005;18:393–410.
- Garcia-Borrón JC, Abdel-Malek Z, Jimenez-Cervantes C. MC1R, the cAMP pathway, and the response to solar UV: extending the horizon beyond pigmentation. *Pigment Cell Melanoma Res* 2014;27:699–720.
- Gerstenblith MR, Goldstein AM, Fargnoli MC, Peris K, Landi MT. Comprehensive evaluation of allele frequency differences of MC1R variants across populations. *Hum Mutat* 2007;28:495–505.
- Han J, Kraft P, Colditz GA, Wong J, Hunter DJ. Melanocortin 1 receptor variants and skin cancer risk. *Int J Cancer* 2006;119:1976–84.
- Herraz C, Journe F, Ghanem G, Jimenez-Cervantes C, Garcia-Borrón JC. Functional status and relationships of melanocortin 1 receptor signaling to the cAMP and extracellular signal-regulated protein kinases 1 and 2 pathways in human melanoma cells. *Int J Biochem Cell Biol* 2012;44:2244–52.
- Kadekaro AL, Leachman S, Kavanagh RJ, Swope V, Cassidy P, Supp D, et al. Melanocortin 1 receptor genotype: an important determinant of the damage response of melanocytes to ultraviolet radiation. *FASEB J* 2010;24:3850–60.
- Pasquali E, Garcia-Borrón JC, Fargnoli MC, Gandini S, Maisonneuve P, Bagnardi V, et al. MC1R variants increased the risk of sporadic cutaneous melanoma in darker-pigmented Caucasians: a pooled-analysis from the M-SKIP project. *Int J Cancer* 2015;136:618–31.
- Perez Oliva AB, Fernandez LP, Detorre C, Herraiz C, Martinez-Escribano JA, Benitez J, et al. Identification and functional analysis of novel variants of the human melanocortin 1 receptor found in melanoma patients. *Hum Mutat* 2009;30:811–22.
- Raimondi S, Sera F, Gandini S, Iodice S, Caimi S, Maisonneuve P, et al. MC1R variants, melanoma and red hair color phenotype: a meta-analysis. *Int J Cancer* 2008;122:2753–60.
- Raimondi S, Gandini S, Fargnoli MC, Bagnardi V, Maisonneuve P, Specchia C, et al. Melanocortin-1 receptor, skin cancer and phenotypic characteristics (M-SKIP) project: study design and methods for pooling results of genetic epidemiological studies. *BMC Med Res Methodol* 2012;12:116.
- Scherer D, Bermejo JL, Rudnai P, Gurzau E, Koppova K, Hemminki K, et al. MC1R variants associated susceptibility to basal cell carcinoma of skin: interaction with host factors and XRCC3 polymorphism. *Int J Cancer* 2008;122:1787–93.
- Scott MC, Wakamatsu K, Ito S, Kadekaro AL, Kobayashi N, Groden J, et al. Human melanocortin 1 receptor variants, receptor function and melanocyte response to UV radiation. *J Cell Sci* 2002;115(Pt 11):2349–55.
- Tagliabue E, Fargnoli MC, Gandini S, Maisonneuve P, Liu F, Kayser M, et al. MC1R gene variants and non-melanoma skin cancer: a pooled-analysis from the M-SKIP project. *Br J Cancer* 2015;113:354–63.
- Valverde P, Healy E, Jackson I, Rees JL, Thody AJ. Variants of the melanocyte-stimulating hormone receptor gene are associated with red hair and fair skin in humans. *Nat Genet* 1995;11:328–30.

Low Levels of Genetic Heterogeneity in Matched Lymph Node Metastases from Patients with Melanoma

Journal of Investigative Dermatology (2016) **136**, 1917–1920; doi:10.1016/j.jid.2016.05.103

TO THE EDITOR

In our previous experience, a high consistency of *BRAF* and *NRAS* mutation patterns was observed between primary tumors and lymph node metastases in patients with advanced

melanoma (Colombino et al., 2012). Conversely, increasing rates of discrepancies in *BRAF*/*NRAS* mutation patterns were found between primary melanomas and metastases in other sites (brain or, mostly, skin) (Colombino

et al., 2012). When the distribution of *BRAF*/*NRAS* mutations was evaluated in a larger cohort, the high rate of consistency in sequence variations of these two genes was further confirmed between primary melanomas and lymph node metastases (142/156; 91%) (Colombino et al., 2013; unpublished data). However, intraindividual heterogeneity of *BRAF* mutations has been



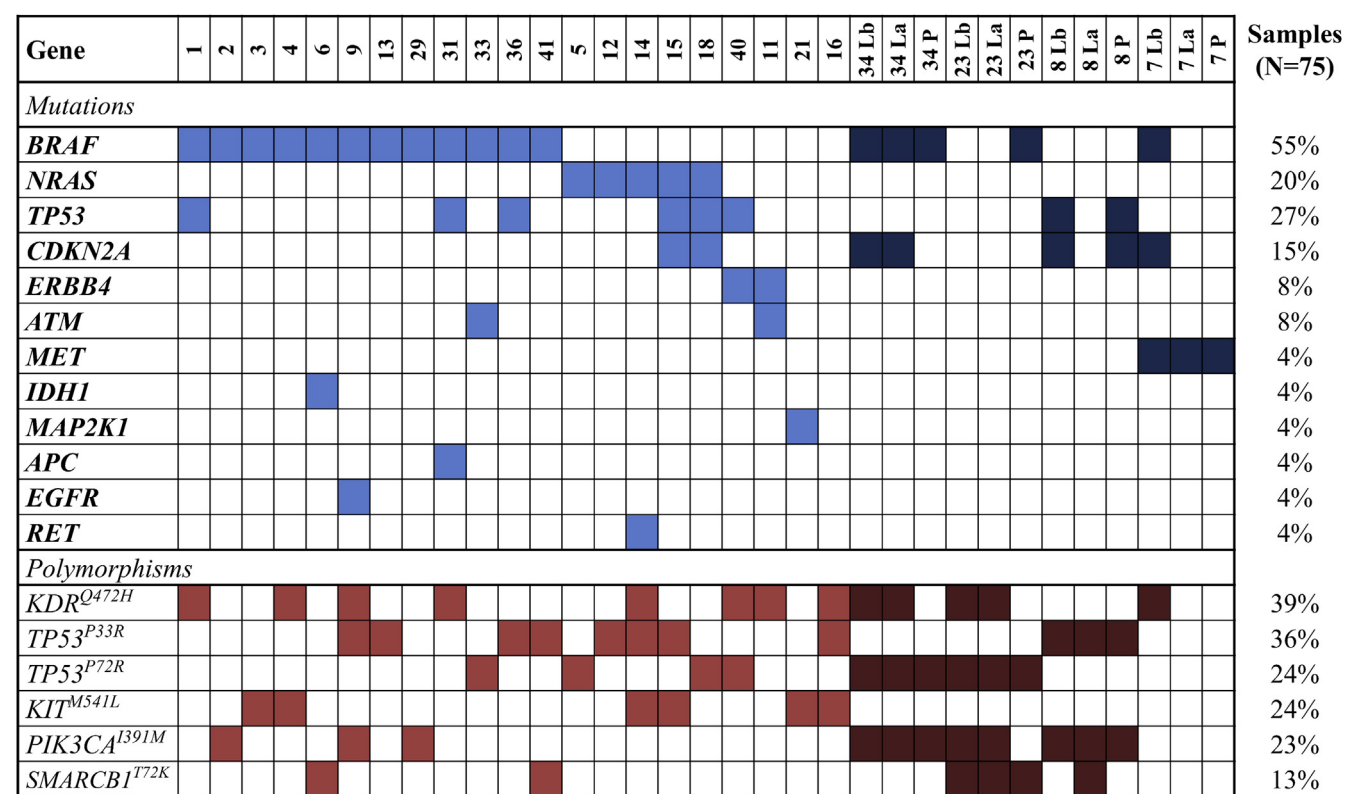


Figure 1. Distribution of nonsilent sequence variations in melanoma samples. Individual gene mutations (blue squares) and polymorphic sequence variants with unknown functional significance (red squares) are shown across all cases. Sample results for the four discrepant cases are indicated with darker colors into the right part of the diagram. La and Lb, the two paired lymph node samples from each patient; P, primary melanoma.

reported in primary and secondary melanomas (Lin et al., 2011; Shi et al., 2014; Yancovitz et al., 2012).

Recently, recurrent or driver mutations have been searched for comparison in different types of primary tumors and matched metastases from the same patients—including melanoma—through next-generation sequencing approaches (The Cancer Genome Atlas Network, 2015; Vakiani et al., 2012; Vignot et al., 2013). Using a next-generation sequencing methodology, we here tried to definitely assess whether clonal expansion to lymph node sites may be associated with consistently low levels of genetic discrepancies in multiple lesions from patients with melanoma.

Genomic DNA was isolated from a total of 75 paired specimens of primary melanomas and synchronous or asynchronous lymph node metastases among consecutively collected patients (N = 25) (see Supplementary Table S1 online), after obtaining their written informed consent for tissue sampling; the study protocol was reviewed and approved by the

Bioethics Committee of the Sassari Healthcare District. In particular, the following formalin-fixed, paraffin-embedded tumor tissues from pathological archives were collected: (i) two asynchronous metastatic lymph nodes sequentially excised from the same patients with melanoma (N = 9) (median time period between first and second nodal excision, 7 months; range, 3–15); (ii) two metastatic lymph nodes synchronously excised from the same patients with melanoma (N = 16); and (iii) corresponding primary melanoma tissues.

Genomic DNA was obtained from macrodissected tumor tissues containing at least 80% neoplastic cells and analyzed for approximately 2,800 mutations in 50 most common oncogenes and tumor suppressor genes, using the AmpliSeq Cancer Panel HotSpot V2/CHPv2 on the Ion Torrent platform (Life Technologies-Thermo Fisher Scientific, Waltham, MA). Data from the runs on such a platform were processed to generate multiple sequence reads, by which redundant mutated alleles were identified (Supplementary Materials

and Methods and Table S2 online). Significantly recurrent gene mutations were validated by Sanger sequencing (Supplementary Materials and Methods); all detected variants have already been reported in both the Human Gene Mutation Database at <http://www.hgmd.cf.ac.uk/ac/index.php> and the Catalogue of Somatic Mutations in Cancer at <http://www.sanger.ac.uk/genetics/CGP/cosmic/>.

Overall, 4 of 25 (16%) analyzed patients presented discrepant mutation patterns between primary melanomas and correspondent lymph nodal metastases (Figure 1; Table 1). Discrepancies were markedly higher among patients with asynchronous (2 of 9; 22%) than among those with synchronous (2 of 16; 12.5%) metastases (Table 1). In nearly all (3 of 4; 75%) discrepant cases, the main differences were observed in driver mutations of the BRAF gene: a mutated metastasis in a patient with wild-type primary tumor (case 7 in Table 1), a mutated primary tumor and wild-type metastases (case 23), or two distinct mutations in the two lesion types (case 34).

Download English Version:

<https://daneshyari.com/en/article/3214893>

Download Persian Version:

<https://daneshyari.com/article/3214893>

[Daneshyari.com](https://daneshyari.com)