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The importance of donor-specific anti-HLA antibodies (DSA) identification in renal transplant patients with C4d-negative biopsies



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ABSTRACT

Two index cases of living-related donor renal allografts patients developed C4d-negative rejection. Both cases had negative cytotoxic crossmatch and negative flow crossmatch before transplantation. The serum creatinine levels were tabulated. Both cases experienced augmented anti-T cell therapy (intravenous methyl prednisolone) at the time of rejection, which failed to improve renal function. Meantime, our HLA lab identified circulating anti-class I and/or II HLA antibodies towards donor mismatched antigens by Luminex multiplex bead array. Additional therapy included high-dose IVIg and plasma exchange. The renal function improved significantly. Furthermore, the donor-specific antibody strength decreased after combined plasmapheresis and IVIG therapy. These cases highlight the importance of donor-specific antibody detection by sensitive solid phase assays in the context of C4d-negative ABMR.

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1. Introduction

The presence of C4d sub-endothelial deposition has been considered a major diagnostic element of antibody-mediated rejection (ABMR) after renal transplantation. More than 60% of late antibody mediated rejection (ABMR) is C4d negative. Particularly important for the Indian setting is the practice of not routinely measuring the DSA by single antigen bead (SAB) assay at the time of graft biopsy done

for evaluating graft dysfunction, as is done in western countries.

2. Case studies

Two index cases of living-related donor renal allografts patients developed C4d-negative rejection. Both cases had negative cytotoxic crossmatch and negative flow crossmatch before transplantation. The serum creatinine levels are shown in

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Table	1 – Cases: profile ar	Table 1 – Cases: profile and response to ABMR specific the	cific therapy.					
Case	Creatinine at baseline (mg/dl)	Histology	C4d	Creatinine at rejection mg/dl	Tacrolimus levels at rejection ng/ml	Anti-HLA DSA	Anti-HLA DSA class I + class II	Creatinine after PP + IVIG mg/dl
						At time of rejection	Post PP+ IV IG)
#1	0.92	Acute T-cell mediated	Negative	2.03	3.2 (normal	Anti-A 26 (MFI 2673),	Solid phase DSA negative	1.7
		rejection peritubular			range 4.0–6.0)	anti-DQ06 (MFI 1522),	(class I MFI 141, class	
		capillaritis				anti-DR11 (MFI 1941),	II MFI 645)	
						anti-DR 13 (MFI 1662),		
						Solid phase DSA:		
						(class I MFI 1187,		
						class II MFI 2438)		
#2	1.4	Focal interstitial	Negative	2.7	1.4 (normal	Anti-B 56 (MFI 1598),	Solid phase DSA negative	1.8
		mononuclear infiltrates			range 4.0-6.0)	anti-DQ03 (MFI 15843)	(class I MFI 243, class	
		with peri-tubular				Solid phase DSA:	II MFI 895)	
		capillary dilatation				(class I MFI 1403,		
						class II MFI 12740)		

Table 1. Both cases experienced augmented anti-T cell therapy (Intravenous methyl prednisolone) at the time of rejection, which failed to improve renal function. Meantime, our HLA lab identified circulating anti-class I and/or II HLA antibodies toward donor mismatched antigens by Luminex multiplex bead array. Additional therapy included high-dose IVIg and plasma exchange. The renal function improved significantly. Furthermore, the donor-specific antibody strength decreased after combined plasmapheresis and IVIG therapy. These cases highlight the importance of donor-specific antibody detection by sensitive solid phase assays in the context of C4d-negative ABMR.

3. Discussion

Sensitive solid-phase platform allows for better diagnosis of ABMR after renal transplantation by identifying donor-specific anti-HLA antibodies. This is valuable for the clinical transplant team, especially in cases of inconclusive histopathology and C4d-negative histology. The routine monitoring of DSA in the post-transplant setting is a predictor of progression of antibody mediated rejection independent of C4d status (Case 2). These two cases provide an important message for the Indian population; where DSA (SAB) is not routinely measured at the time of transplant biopsy and where C4d staining alone has until recently been the gold standard for the diagnosis of ABMR.

At our center (Sanjay Gandhi Post Graduate Institute of Medical Sciences), living related renal transplant recipients with negative pre transplant CDC crossmatch were evaluated by solid phase cross match (SPC) after transplantation. 21.5% recipients had DSA by SPC and those who were positive for HLA class II antibodies had significant association with ABMR. Solid phase crossmatch (SPC) testing performed with donor lysate on Luminex platform could sever as an estimate for diagnosis of rejection episodes. SAB is gold standard for the diagnosis of ABMR (Figs. 1 and 2).

DSA with single or multiple antigen beads is costly (particularly for individual patients). Solid phase crossmatch (SPC) testing is less expensive than SAB assay. Donor-specific antibody testing by SPC costs Rs. 2800 per test and by SAB assay costs 25,000 per test.

Solid phase crossmatch assay by Luminex can also be used for post-transplant routine DSA monitoring (economical for Indian population, although this test has limitations for the detection of HLA antibodies).

The presence of pre-transplant antibodies directed against donor antigens (HLA) called donor-specific antibodies (DSA) is strongly associated with hyper acute and acute rejection.^{2–4} Production of de novo DSA after transplant is indicative of the antibody-mediated rejection (AMR).⁵ The HLA-DSA monitoring is an important tool for rejection risk assessment in kidney transplant recipients.^{6,7}

3.1. Complement dependant cytotoxicity (CDC) cross match

This test was first standardized by Terasaki in 1960s.⁸ It is used to detect the donor-specific antibodies directed against the recipient. Patel and Terasaki developed the CDC cross match for pretransplant donor recipient evalua-

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