

# Distribution of innate efflux-mediated aminoglycoside resistance among different *Achromobacter* species

J. Bador, C. Neuwirth, P. Liszczynski, M.-C. Mézier, M. Chrétiennot, E. Grenot, A. Chapuis, C. de Curraize and L. Amoureux

Department of Bacteriology, University Hospital of Dijon, Dijon, France

## Abstract

*Achromobacter* spp. are emerging respiratory pathogens in cystic fibrosis patients. Since 2013 the genus *Achromobacter* includes 15 species for which innate antibiotic resistance is unknown. Previously the AxyXY-OprZ efflux system has been described to confer aminoglycoside (AG) resistance in *A. xylosoxidans*. Nevertheless, some *Achromobacter* spp. strains are susceptible to AG. This study including 49 *Achromobacter* isolates reveals that AG resistance is correlated with different *Achromobacter* spp. It is noteworthy that the *axyXY-oprZ* operon is detected only in AG-resistant species, including the most frequently encountered in cystic fibrosis patients: *A. xylosoxidans*, *A. ruhlandii*, *A. dolens* and *A. insuavis*.

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**Keywords:** *Achromobacter*, aminoglycoside resistance, AxyXY-OprZ, cystic fibrosis, *nrdA*, selection pressure

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**Corresponding author:** C. Neuwirth, Laboratoire de Bactériologie, Hôpital Universitaire, Plateau technique de Biologie, BP 37013, 21070 Dijon, Cedex, France

**E-mail:** [catherine.neuwirth@chu-dijon.fr](mailto:catherine.neuwirth@chu-dijon.fr)

J. Bador and C. Neuwirth contributed equally to this article, and both should be considered first author.

*Achromobacter* spp. are nonfermenting Gram-negative bacilli considered as emerging pathogens in cystic fibrosis (CF)

patients [1,2]. Since the description of the type species, *A. xylosoxidans* [3], 14 other species have been ranked into the genus *Achromobacter*: *A. piechaudii* and *A. ruhlandii* [4], *A. denitrificans* [5], *A. spanius* and *A. insolitus* [6], *A. marplatensis* [7], *A. animicus*, *A. mucicolens*, *A. pulmonis* and *A. spiritinus* [8], *A. insuavis*, *A. aegrifaciens*, *A. anxifer* and *A. dolens* [9], and 6 other genogroups. These 21 species and genogroups can be distinguished by the multilocus sequence typing (MLST) scheme proposed by Spilker et al. [10]. The study demonstrated that sequencing a 765 bp internal fragment of the only *nrdA* gene is sufficient for correct identification [11]. Because of the actual difficulty in performing accurate species identification, most isolates are still referred by default as *A. xylosoxidans*, preventing the evaluation of the real epidemiology and clinical impact of each species. Moreover, the data about the mechanisms of innate antibiotic resistance are scarce [12,13]. The AxyXY-OprZ RND efflux system confers resistance to aminoglycosides (AG) in *A. xylosoxidans* AXX-A since reclassified as *A. insuavis* (accession number [AFRQ01000000](https://ncbi.nlm.nih.gov/nucl/AFRQ01000000)). Nevertheless, AG, which take an important part in CF antimicrobial therapy, remain active against some isolates of *Achromobacter* spp. [14,15].

We sought to describe the distribution of AG-resistant isolates among the different species of the genus *Achromobacter* and to search for the *axyXY-oprZ* efflux operon in AG-resistant and -susceptible isolates to assess if AG resistance is correlated with the presence of the operon.

Forty-nine *Achromobacter* isolates harbouring various AG resistance patterns were included in this study: 21 from CF patients' sputum, 20 from non-CF clinical samples and eight from environmental samples (Table 1). Most of them ( $n = 35$ ) were collected in our laboratory; nine collection strains were purchased from the Institut Pasteur, France, including six type strains, and five were kindly provided by J. J. LiPuma (Department of Pediatrics and Communicable Diseases, University of Michigan Medical School). Isolates were identified at the genus level either by using the conventional biochemical method API 20NE (bioMérieux) or by sequencing the 16S rRNA gene. The identification to the species level was performed by sequencing the 765 bp internal *nrdA* fragment followed by *Achromobacter* PubMLST database query (<http://pubmlst.org/achromobacter/>). Minimal inhibitory concentrations (MICs) of tobramycin, amikacin, gentamicin and netilmicin were measured by the Etest method (bioMérieux). Mueller-Hinton agar plates were inoculated by swabbing from a 0.5 McFarland turbidity bacterial suspension, and MICs were recorded after overnight incubation at 37°C by two persons independently. The phenotype "AG-susceptible" (AG-S) was attributed to isolates susceptible to all

TABLE 1. *Achromobacter* isolates and main results

nrdA identification	Isolate	Origin	MIC (mg/L)				AG S/R	PCR axyY	PCR oprZ
			TOB	AMK	GEN	NET			
<i>A. aegrifaciens</i>	ACH-CF-D59	CF sputum <sup>a</sup>	>256	>256	>256	>256	R	+	+
<i>A. aegrifaciens</i>	ACH-CF-802	CF sputum <sup>a</sup>	64	48	16	32	R	+	+
<i>A. aegrifaciens</i>	ACH-ENV-2	Hospital hand-washing sink <sup>a</sup>	12	8	3	8	R	+	+
<i>A. aegrifaciens</i>	ACH-CF-766	CF sputum <sup>a</sup>	192	48	12	64	R	+	+
<i>A. animicus</i>	ACH-CF-864	CF sputum <sup>a</sup>	1.5	4	1	1	S	–	–
<i>A. animicus</i>	ACH-NCF-33	Catheter <sup>a</sup>	1.5	6	2	2	S	–	–
<i>A. animicus</i>	ACH-CF-D63	CF sputum <sup>a</sup>	1.5	4	1.5	1	S	–	–
<i>A. animicus</i>	ACH-CF-D64	CF sputum <sup>a</sup>	2	8	2	1.5	S	–	–
<i>A. animicus</i>	ACH-CF-D65	CF sputum <sup>a</sup>	1	3	1.5	1.5	S	–	–
<i>A. animicus</i>	ACH-CF-711	CF sputum <sup>a</sup>	2	4	1.5	1.5	S	–	–
<i>A. denitrificans</i>	CIP-77.15T	Soil	32	256	64	64	R	+	+
<i>A. dolens</i>	AUI8822	CF sputum	>256	64	>256	12	R	+	+
<i>A. dolens</i>	AU20310	CF sputum	>256	>256	>256	128	R	+	+
<i>A. genogroup 12</i>	ACH-ENV-3	Dialysis water <sup>a</sup>	3	32	8	8	R	+	+
Novel species	ACH-CF-583	CF sputum <sup>a</sup>	24	>256	96	>256	R	+	+
<i>A. insolitus</i>	CIP-108202T	Leg wound	256	32	48	>256	R	+	+
<i>A. insuavis</i>	ACH-CF-476	CF sputum <sup>a</sup>	>256	>256	>256	>256	R	+	+
<i>A. insuavis</i>	ACH-CF-777	CF sputum <sup>a</sup>	96	>256	>256	>256	R	+	+
<i>A. insuavis</i>	AXX-A	Ear swab <sup>a</sup>	16	256	24	64	R	+	+
<i>A. insuavis</i>	CIP-102062	Blood	12	256	16	24	R	+	+
<i>A. marplatensis</i>	ACH-ENV-4	Lake <sup>a</sup>	12	256	24	32	R	+	+
<i>A. mucicolens</i>	ACH-NCF-34	Tracheal aspirate <sup>a</sup>	1.5	6	1.5	1.5	S	–	–
<i>A. mucicolens</i>	ACH-NCF-35	Tracheal aspirate <sup>a</sup>	2	8	2	1.5	S	–	–
<i>A. mucicolens</i>	ACH-NCF-36	Blood <sup>a</sup>	2	6	2	2	S	–	–
<i>A. mucicolens</i>	ACH-CF-510	CF sputum <sup>a</sup>	2	8	2	2	S	–	–
<i>A. piechaudii</i>	CIP-60.75T	Pharynx	1.5	6	3	3	S	–	–
<i>A. ruhlandii</i>	CIP-77.26T	Soil	8	24	12	16	R	+	+
<i>A. ruhlandii</i>	AUI9877	CF sputum	16	>256	48	64	R	+	+
<i>A. ruhlandii</i>	AUI9911	CF sputum	3	48	6	12	R	+	+
<i>A. ruhlandii</i>	AUI9929	CF sputum	>256	>256	>256	>256	R	+	+
<i>A. spanius</i>	CIP-108199T	Blood	1.5	6	4	4	S	–	–
<i>A. spanius</i>	ACH-NCF-37	Foot wound <sup>a</sup>	1	4	1	1.5	S	–	–
<i>A. spanius</i>	ACH-CF-746	CF sputum <sup>a</sup>	2	6	2	2	S	–	–
<i>A. xylosoxidans</i>	ACH-CF-809	CF sputum <sup>a</sup>	128	>256	256	>256	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-39	Insertion-site skin swab <sup>a</sup>	24	>256	64	128	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-18	Tracheal aspirate <sup>a</sup>	64	>256	128	256	R	+	+
<i>A. xylosoxidans</i>	CIP-71.32T	Ear discharge	192	>256	>256	>256	R	+	+
<i>A. xylosoxidans</i>	CIP-101902	Pleural fluid	>256	>256	>256	>256	R	+	+
<i>A. xylosoxidans</i>	CIP-102236	Sputum	48	>256	256	>256	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-41	Sputum <sup>a</sup>	32	>256	64	128	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-42	Tracheal aspirate <sup>a</sup>	8	256	24	64	R	+	+
<i>A. xylosoxidans</i>	ACH-ENV-1	Dental instrument <sup>a</sup>	192	>256	>256	>256	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-13	Bronchial aspirate <sup>a</sup>	64	>256	128	>256	R	+	+
<i>A. xylosoxidans</i>	ACH-CF-805	CF sputum <sup>a</sup>	24	>256	48	192	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-11	Sputum <sup>a</sup>	16	256	32	48	R	+	+
<i>A. xylosoxidans</i>	ACH-CF-842	CF sputum <sup>a</sup>	8	192	24	32	R	+	+
<i>A. xylosoxidans</i>	ACH-NCF-40	Blood <sup>a</sup>	32	>256	96	192	R	+	+
<i>A. xylosoxidans</i>	ACH-ENV-5	River <sup>a</sup>	16	>256	24	64	R	+	+
<i>A. xylosoxidans</i>	ACH-ENV-6	Domestic hand-washing sink <sup>a</sup>	32	>256	48	128	R	+	+

AG, aminoglycoside; AMK, amikacin; CF, cystic fibrosis; GEN, gentamicin; MIC, minimum inhibitory concentration; NET, netilmicin; R, resistant; S, susceptible; TOB, tobramycin.

<sup>a</sup>Isolates collected in our laboratory.

AG and the phenotype “AG-resistant” (AG-R) to the other by using the European Committee on Antimicrobial Susceptibility Testing clinical breakpoints for *Pseudomonas* spp. ([http://www.eucast.org/clinical\\_breakpoints/](http://www.eucast.org/clinical_breakpoints/); version 5.0). Detection of the *axyXY-oprZ* operon was performed by 2 PCRs targeting the genes (a) *axyY*, encoding the RND transporter, and (b) *oprZ*, encoding the outer membrane factor. PCRs were carried out in reaction mixtures containing dNTP (0.2 mM), forward and reverse primers (0.25 µM each), Taq polymerase (Fermentas) (2.5 U) with the supplied buffer, MgCl<sub>2</sub> (1.5 mM), dimethyl sulfoxide (5% volume) and template DNA (1 µL), adjusted with water to a final volume of 50 µL. The cycling parameters were 94°C for 10 minutes, 30 cycles of 94°C for 90 seconds, annealing primers temperature for 90 sections, 72°C for 60

seconds, and 72°C for 10 minutes. The results are summarized in Table 1.

The *nrdA* sequences analysis allowed identification of 48 of the 49 isolates. *nrdA* sequence of ACH-CF-583 harboured 39 nucleotide differences, with its closest match in database (genogroup 19) (Fig. 1) indicating that this isolate belonged to a novel genogroup or a novel species. Fourteen of the 49 studied isolates were categorized as AG-S. Interestingly, all isolates belonging to a same species harboured the same AG resistance pattern. In the resistant species, the level of resistance was sometimes variable among the isolates. Nevertheless, none of these isolates had been categorized as susceptible for all four AG molecules. A variable expression level of the efflux operon might account for these differences as already observed for

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