

Proximal tubule sphingosine kinase-1 has a critical role in A₁ adenosine receptor-mediated renal protection from ischemia

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Renal ischemia–reperfusion injury is a major cause of acute kidney injury. We previously found that renal A₁ adenosine receptor (A₁AR) activation attenuated multiple cell death pathways including necrosis, apoptosis, and inflammation. Here, we tested whether induction of cytoprotective sphingosine kinase (SK)-1 and sphingosine-1-phosphate (S1P) synthesis might be the mechanism of protection. A selective A₁AR agonist (CCPA) increased the synthesis of S1P and selectively induced SK1 in mouse kidney and HK-2 cells. This agonist failed to protect SK1-knockout but protected SK2-knockout mice against renal ischemia–reperfusion injury indicating a critical role of SK1 in A₁AR-mediated renal protection. Inhibition of SK prevented A₁AR-mediated defense against necrosis and apoptosis in HK-2 cells. A selective S1P₁R antagonist (W146) and global *in vivo* gene knockdown of S1P₁Rs with small interfering RNA completely abolished the renal protection provided by CCPA. Mice selectively deficient in renal proximal tubule S1P₁Rs (S1P₁R^{f/f} PEPCK^{Cre/+}) were not protected against renal ischemia–reperfusion injury by CCPA. Mechanistically, CCPA increased nuclear translocation of hypoxia-inducible factor-1 α in HK-2 cells and selective hypoxia-inducible factor-1 α inhibition blocked A₁AR-mediated induction of SK1. Thus, proximal tubule SK1 has a critical role in A₁AR-mediated protection against renal ischemia–reperfusion injury.

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Renal ischemia–reperfusion (IR) injury is a frequent cause of acute kidney injury (AKI).¹ AKI a major clinical problem often leading to multi-organ dysfunction and systemic inflammation with extremely high mortality.² Unfortunately, the severity and incidence of AKI has been increasing without any improvements in therapy or patient survival over the past 50 years.³ Currently, the incidence of renal dysfunction after major surgery in high-risk patients has been reported to be as high as 70–80%.^{4–6} Despite continued research on renal protective strategies, there are no proven therapies to reduce AKI in the perioperative setting.

We have previously demonstrated that A₁ adenosine receptor (A₁AR) activation reduced necrosis and apoptosis in cultured proximal tubule cells.^{7,8} Moreover, A₁AR activation immediately before renal ischemia protected against renal IR injury in rats and mice *in vivo*.^{9,10} In addition to reduction in renal necrosis and apoptosis,^{9,11} we also observed a surprising reduction in renal inflammation (reduced leukocyte influx and pro-inflammatory cytokine upregulation) as an important component of renal protection with A₁AR activation. Unlike the better characterized and traditionally understood anti-inflammatory mechanisms of A_{2a}ARs, the anti-inflammatory mechanisms of A₁ARs remain unclear. Therefore, we hypothesized that A₁AR activation may induce additional mediator(s) that produce direct anti-inflammatory effects to protect against renal IR.

Phosphorylation of sphingosine by two subtypes of sphingosine kinase (SK1 and SK2) leads to the formation of sphingosine-1-phosphate (S1P), a lysophospholipid targeting G-protein coupled receptors with diverse extracellular as well as intracellular effects.^{12–14} In particular, renal tubular SK1 activation has been shown to produce renoprotection after IR.^{15–17} Moreover, of five G-protein coupled S1P receptors (S1PRs), S1P₁R activation has been shown to counteract against cardiac^{18,19} and renal IR injury^{17,20} and attenuates T-lymphocyte-mediated tissue inflammation.^{13,21,22} Furthermore, S1P₁R agonists produce renal protection via direct activation of S1P₁Rs in renal proximal tubules.²³

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In this study, we tested the hypothesis that renal tubular A₁AR activation produces anti-inflammatory and cytoprotective S1P via activation of renal proximal tubule SK. We utilized genetically modified strains of mice lacking SK1 (SK1^{-/-}) and SK2 (SK2^{-/-}) enzyme. In addition, we tested that activation of S1P₁R via A₁AR-mediated S1P generation is critical in renal protection utilizing mice treated with a selective S1P₁R antagonist (W146) as well as mice treated with small interfering RNA (siRNA) designed for *in vivo* targeting of S1P₁R. To investigate the direct involvement of proximal tubule S1P₁R in A₁AR agonist-mediated renal protection *in vivo*, we generated mice with proximal tubule-specific deletion of S1P₁R by crossing mice carrying the floxed S1P₁R gene¹² with mice expressing Cre recombinase under the control of the phosphoenolpyruvate carboxykinase promoter.²⁴ Finally, we examined the mechanisms of A₁AR-mediated renal proximal tubule SK activation. Utilizing cultured human proximal tubule cells *in vitro*, we tested the hypothesis that hypoxia-inducible factor-1 α (HIF-1 α) has a critical role in mediating A₁AR-mediated induction of SK1.

RESULTS

A₁AR activation or overexpression increases SK1 synthesis and induces SK activity in mouse kidney

We initially tested whether A₁AR activation increased SK1 expression and activity in mouse kidney (cortex and corticomedullary junction). Figure 1a and b show that a selective A₁AR agonist 2-chloro-N6-cyclopentyladenosine (CCPA) (0.1 mg/kg, intraperitoneal (i.p.)) increased SK1 mRNA (measured at 6 h) and protein expression (measured at 16 h) and upregulated SK activity (measured at 6 h) in mouse kidney. In contrast, the A₁AR agonist CCPA failed to

increase SK2 synthesis in mouse kidney. We measured preferentially SK1 activity by adding Triton x-100 in our SK activity assay as described by Vessey *et al.*²⁵

Next, we aimed to determine whether overexpression of renal A₁ARs increased SK1 expression in mouse kidney. We previously demonstrated that intrarenal A₁AR-enhanced green fluorescent protein (EGFP) lentivirus transduction resulted in a selective and robust A₁AR overexpression in the injected kidneys within 48 h.²⁶ In this study, we show that kidneys injected with A₁AR-EGFP lentivirus 48 h prior showed increased SK1 mRNA and protein expression compared with EGFP lentivirus-injected kidneys (Figure 1c). SK2 mRNA expression did not change with A₁AR-EGFP lentivirus injection.

SK1 activation is critical for A₁AR-mediated protection against renal IR

Next, we tested whether SK1 activation is critical in A₁AR-mediated protection against renal IR. Plasma creatinine values were similar between sham-operated (anesthesia, laparotomy, right nephrectomy and recovery) wild-type (WT) creatinine (Cr) = 0.4 $\pm$ 0.03 mg/dl, N = 3), SK1^{-/-} (SK1 knockout, Cr = 0.4 $\pm$ 0.06 mg/dl, N = 3), and SK2^{-/-} (SK2 knockout, Cr = 0.4 $\pm$ 0.09 mg/dl, N = 3) mice. Plasma creatinine increased in WT, SK1^{-/-}, and SK2^{-/-} mice subjected to 30-min renal IR compared with sham-operated mice (Figure 2a). As we described previously, SK1^{-/-} and SK2^{-/-} mice had slightly increased and decreased renal injury, respectively, compared with WT mice.¹⁷ Treatment with CCPA significantly attenuated the increases in plasma creatinine in WT and SK2^{-/-} mice but not in SK1^{-/-} mice (Figure 2a). Figure 2b demonstrates increased histological

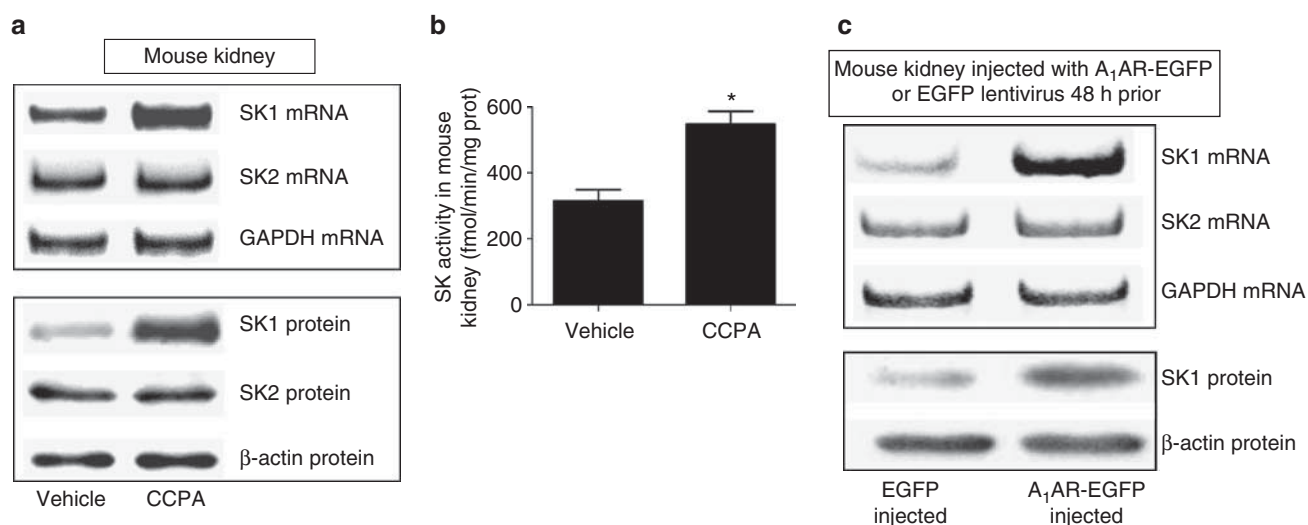


Figure 1 | A₁ adenosine receptor (A₁AR) activation or overexpression increases sphingosine kinase (SK)-1 expression and activity in mouse kidney. (a) Representative bands for SK1 and SK2 mRNA (reverse transcription (RT)-PCR, 6 h) and protein (immunoblotting, 16 h) expression in mouse kidney treated with vehicle (0.4% dimethyl sulfoxide (DMSO) in saline) or 2-chloro-N6-cyclopentyladenosine (CCPA) (0.1 mg/kg, intraperitoneal (i.p.)) (N = 4 for each group). (b) Mouse kidney SK activity 6 h after vehicle (0.4% DMSO) or CCPA (0.1 mg/kg, i.p.) treatment (N = 4 for each group). Data are presented as means $\pm$ s.e.m. *P < 0.05 vs. vehicle-treated mice. (c) Representative bands for SK1 and SK2 mRNA (RT-PCR) and SK1 protein (immunoblotting) expression in mouse kidney injected with enhanced green fluorescent protein (EGFP) or A₁AR-EGFP lentivirus 48 h prior (N = 4 for each group). GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

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