

available at www.sciencedirect.comwww.elsevier.com/locate/brainres**BRAIN
RESEARCH****Research Report****Pathway and gene ontology based analysis of gene expression in a rat model of cerebral ischemic tolerance**Zheng Feng^{a,b}, Daniel P. Davis^c, Roman Šášik^d, Hemal H. Patel^a, John C. Drummond^{a,b}, Piyush M. Patel^{a,b,*}^aDepartment of Anesthesiology, University of California, San Diego, USA^bVeterans Affairs San Diego Health Care System, San Diego, USA^cDepartment of Emergency Medicine, University of California, San Diego, USA^dUCSD Cancer Center and Biomedical Genomics Microarray Facility, University of California, San Diego, USA

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ABSTRACT

Ischemic tolerance is a phenomenon whereby a sublethal ischemic insult [ischemic preconditioning (IPC)] provides robust protection against subsequent lethal ischemia. Activation of N-methyl-D-aspartate (NMDA) receptors and subsequent new gene transcription are required for tolerance. We utilized the NMDA antagonist, MK801, prior to the IPC stimulus to separate candidate genes from epiphenomena. Rats were divided into four groups: vehicle/IPC (preconditioned), MK801/IPC (attenuated preconditioning), vehicle/sham (non-preconditioned), and MK801/sham (non-preconditioned). Hippocampi (5/group/time point) were harvested immediately after ischemia as well as 1, 4, and 24 h post-ischemia to profile gene expression patterns using microarray analyses. Extracted mRNAs were pooled and subsequently hybridized to Affymetrix arrays. In addition, groups of rats were sacrificed for Western blot analysis and histological studies. Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway and gene ontology (GO) analyses were used to identify functionally related groups of genes whose modulation was statistically significant, while hierarchical cluster analysis was used to visualize the fold expression within these groups. Significantly modulated pathways included: MAP kinase signaling pathway, Toll receptor pathway, TGF- β signaling pathways, and pathways associated with ribosome function and oxidative phosphorylation. Our data suggest that the tolerant brain responds to subsequent ischemic stress by partially downregulating inflammatory and upregulating protein synthesis and energy metabolism pathways.

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1. Introduction

Ischemic tolerance (IT) is a phenomenon whereby a brief exposure to ischemic episodes results in ischemic precondition-

ing (IPC), providing neuroprotection against subsequent lethal ischemic insults (Dinaghi et al., 2003; Kirino, 2002; Schaller, 2005). Although the precise mechanisms by which tolerance is induced are not clear, the available evidence indicates that IPC

* Corresponding author. Department of Anesthesiology, Veterans Affairs San Diego Health Care System and University of California, San Diego, CA 92161, USA. Fax: +1 858 534 0104.

E-mail address: ppatel@ucsd.edu (P.M. Patel).

involves new gene transcription and *de novo* protein synthesis (Truettner et al., 2002). Previous studies have shown that IPC modulates the transcription of genes associated with apopto-

sis, growth factors, free radical metabolism, and inflammation. These studies have revealed modulations in expression for genes involved in synthesis of proteins, cytokines, and ATP in

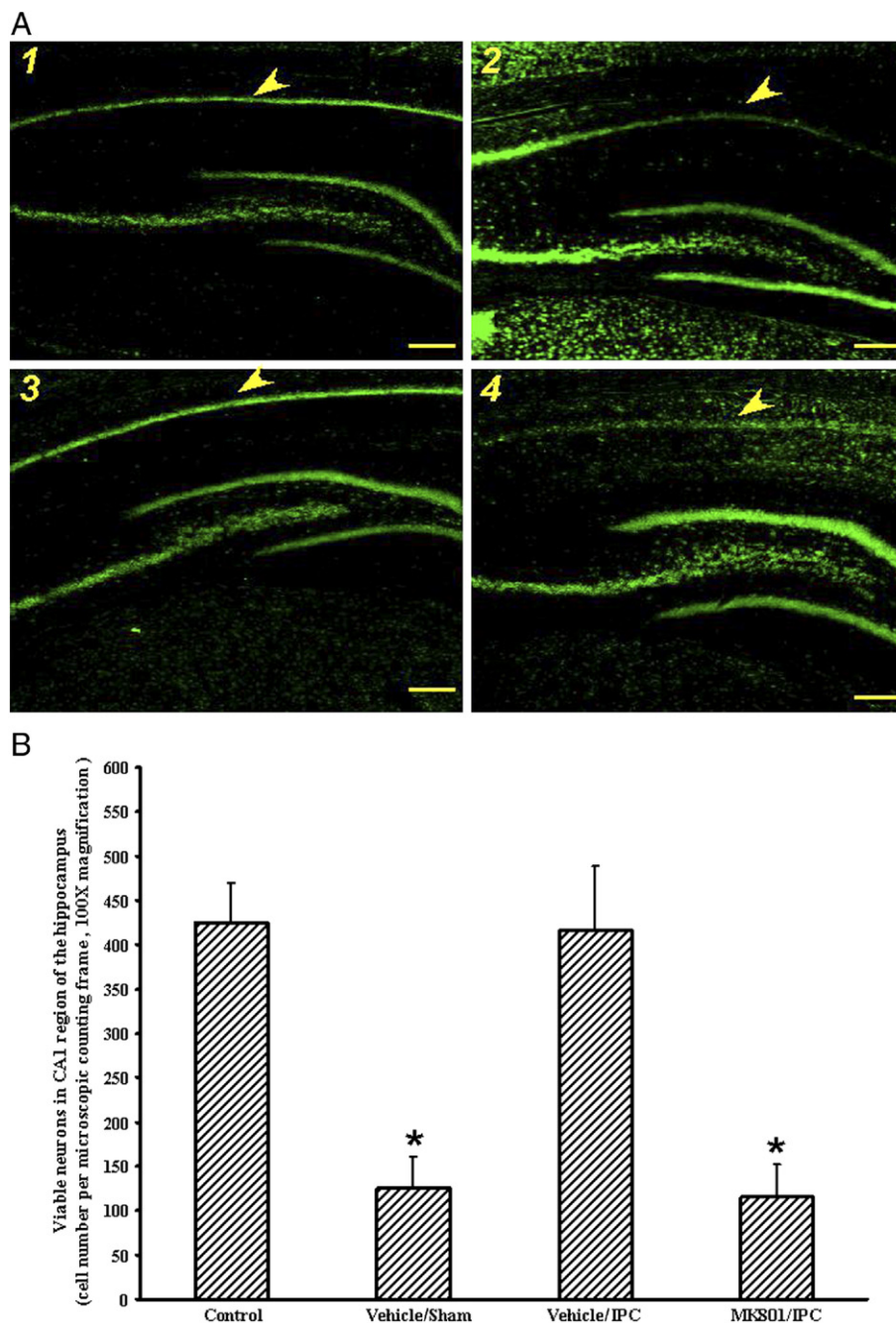


Fig. 1 – Validation of in vivo model. (A) Representative images demonstrating neuronal viability in the hippocampal CA1 sub-region. Coronal sections of the rat hippocampi were stained by NeuroTrace Nissl and observed under 50× magnification. 1. Nissl-stained section from a non-lesioned naive control showing broad and even distribution of CA1 pyramidal cells (arrows). 2. Lethal ischemia-induced marked loss of pyramidal neurons in CA1 region (arrows) in the vehicle/sham group. 3. IPC afforded nearly complete protection against subsequent lethal ischemia (arrows). 4. MK801 (3 mg/kg) pretreatment attenuated the protective effects of IPC (scale bar = 100 μm; magnification: 50×). **(B)** Quantitative analysis of neuronal loss in hippocampus CA1 region at baseline (control) and 7 days following lethal ischemia in vehicle/IPC, vehicle/sham, and MK801/IPC rats. The Nissl-stained sections were observed under an Olympus fluorescence microscope at 200× magnification. The number of surviving neurons in mediolateral segment of CA1 region was counted in three representative sections for each rat ($n=3$). Neurons were counted only if their nuclei were completely within the margins of the counting frame. Data are expressed as mean ± SD ($n=3$ per group). Statistical significance ($P<0.05$) versus naive animals is indicated (*).

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