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Research Report
Characterization of *Gpr101* expression and G-protein coupling selectivity

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ABSTRACT

This report describes the identification and characterization of the murine orphan GPCR, *Gpr101*. Both human and murine genes were localized to chromosome X. Similar to its human ortholog, murine *Gpr101* mRNA was detected predominantly in the brain within discrete nuclei. A knowledge-restricted hidden Markov model-based algorithm, capable of accurately predicting G-protein coupling selectivity, indicated that both human and murine GPR101 were likely coupled to Gs. This prediction was supported by the elevation of cyclic AMP levels and the activation of a cyclic AMP response element-luciferase reporter gene in HEK293 cells over-expressing human GPR101. Consistent with this, over-expression of human GPR101 in a yeast-based system yielded an elevated, agonist-independent reporter gene response in the presence of a yeast chimeric G α s protein. These results indicate that GPR101 participates in a potentially wide range of activities in the CNS via modulation of cAMP levels.

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1. Introduction

The recent identification of numerous G-protein-coupled receptors (GPCRs) in the mammalian genome has created a large catalog of uncharacterized receptors (Vassilatis et al., 2003). This presents the significant challenge of identifying cognate ligands for these numerous orphan GPCRs (oGPCRs). Investigators have used a diversity of approaches to identify endogenous ligands for oGPCRs including bioinformatic methods applied to oGPCR sequences, searching sequence databases for likely peptide ligands, testing panels of biologically active compounds and screening fractionated tissue extracts in a process referred to as reverse pharma-

cology (Civelli et al., 2001; Lee et al., 2003; Wise et al., 2004). In reverse pharmacology approaches, oGPCRs are first expressed in cell lines that permit efficient functional expression followed by detection of G-protein- and agonist-dependent activation by various methods. These methods take advantage of well-characterized signal transduction pathways with robust downstream readouts, such as ³⁵S-GTP γ S binding, second messenger production (e.g. cAMP, inositol phosphates, intracellular Ca²⁺) and second messenger-dependent, G-protein-coupled reporter genes. Using these systems, oGPCR ligands have been identified by screening known compound panels prioritized based on the similarity of oGPCRs to GPCRs with known ligands (Briscoe et

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