

Cocaine exposure during the early postnatal period diminishes medial frontal cortex Gs coupling to dopamine D₁-like receptors in adult rat

Ning Zhao^a, Hoau-Yan Wang^b, Diana Dow-Edwards^{a,*}

^a Department of Physiology and Pharmacology, SUNY Downstate Medical Center, Brooklyn, NY, United States

^b Department of Physiology and Pharmacology, Sophie Davis School of Biomedical Education, CUNY Medical School, NY, United States

ARTICLE INFO

Article history:

Received 16 November 2007

Received in revised form 31 March 2008

Accepted 7 April 2008

Keywords:

Dopamine

Cocaine

Receptor

Postnatal

Medial frontal cortex

Caudate/putamen

ABSTRACT

The effect of cocaine exposure during early postnatal ages on coupling of dopamine (DA) D₁- and D₂-like receptors to their respective Gs/olf and Gi was examined in striatum and medial frontal cortex (MFC). Sprague–Dawley rats were subcutaneously injected with either 50 mg/kg cocaine or vehicle during postnatal day (PnD) 11–20 and dopaminergic D₁- and D₂-like receptor signaling was evaluated at PnD 60. Results showed that cocaine exposure did not affect the magnitude of both DA D₁- and D₂-like receptor coupling to their respective Gs/olf and Gi in striatum. However, in the medial frontal cortex, the basal and the DA D₁-like receptor and Gs association were reduced in cocaine-exposed brains. However, there was no change in basal or DA D₂-like receptor–Gi linkage in medial frontal cortex. Since frontal cortex plays a critical role in regulating cognition and working memory, disruption of DA-modulated circuits or alteration of dopaminergic activity resulting from postnatal cocaine exposure may result in abnormal responses to environmental challenges leading to long-term behavioral changes.

© 2008 Elsevier Ireland Ltd. All rights reserved.

Cocaine abuse among childbearing women in the United States remains a public health issue [18]. Evidence from both clinical [23,26] and animal experiments suggests that use of cocaine during pregnancy may cause long-lasting behavioral abnormalities [24,8,2,19]. Even though the exact mechanism through which cocaine exposure produces its long-lasting behavioral effects is largely unknown, dysfunction of dopamine (DA) transmembrane signaling has been implicated [14,5,15,22,7]. While prenatal cocaine exposure reduces Gs/olf–DA D₁ receptor coupling [5,7], effects of cocaine on DA receptor signaling have not been studied following early postnatal exposure in the rat, a period of brain development similar to late third trimester brain development in human. Since this period is characterized by synaptic pruning and functional development of multiple forebrain systems, we hypothesized that cocaine administration during this period may alter DA receptor signaling. In the present study, cocaine was injected subcutaneously during postnatal day (PnD) 11–20 and the coupling of the DA D₁- and D₂-like receptors to their respective Gs/olf and Gi proteins, the key step in signaling, was assessed in striatum and medial frontal cortex (MFC) at adulthood (PnD 60).

All animal protocols were approved by SUNY's Institutional Animal Care and Use Committee. Adult female Sprague–Dawley rats (VAF, Charles River, Wilmington, ME) were mated in our AAALAC-

accredited vivarium (20–22 °C with 12 h light–dark cycles, lights on 7 a.m.) with males of the same strain. Starting from the morning of a sperm-positive smear, referred to as gestation day 1(G1), they were housed individually with ad lib food and water and left undisturbed until day of birth in 44 cm × 24 cm × 20 cm plastic cages with wood chip bedding. On the day of birth (PnD 1), the litter was culled to 12 pups maintaining equivalent gender representation, if possible, and the pups were toe-clipped for identification. Litters were randomly placed into one of two treatment groups: 50 mg/kg cocaine HCl (Sigma, St. Louis, MO) or vehicle (sterile water, 5 μl/g body weight, Baxter). Subcutaneous injections were administered daily from days 11 to 20 between 11:00 and 13:00. On PnD 21 the pups were weaned into same-sex cages, ear-clipped for identification and weighed every 4 days thereafter until they were 60 days of age. At 60 days, rats were weighed, taken to the necropsy room one at a time and placed in a CO₂ chamber until lightly anesthetized and then decapitated. The brains were rapidly removed and put in –50 °C methylbutane for 30 s to solidify. Then the slice containing the striatum (–0.4–1.6 mm relative to Bregma) and medial frontal cortex (5.5–4.2 mm relative to Bregma) was dissected from the remaining piece and then frozen in methylbutane at –20 °C for 1 min. The methylbutane was evaporated and the brain sections were put in labeled bags and frozen at –80 °C until thawed for preparation of membranes.

To assess the effect of cocaine exposure on the linkage between DA D₁ receptors and Gs/olf proteins as well as the coupling between DA D₂ receptors and Gi proteins, crude neuronal membranes

* Corresponding author.

E-mail address: diana.dow-edwards@downstate.edu (D. Dow-Edwards).

were prepared from rat brain striata and medial frontal cortices as described previously [9,28]. Tissues were thawed on ice and then homogenized in 10 volumes of 25 mM HEPES (pH 7.5) buffer containing 2 mM $MgCl_2$, 1 mM EDTA, 0.2% 2-mercaptoethanol, 50 μ g/ml leupeptin, 25 μ g/ml pepstatin A, 0.01 U/ml soybean trypsin inhibitor, 0.04 mM phenylmethylsulfonyl fluoride (PMSF) using glass/glass homogenizer. Homogenate was centrifuged at 800 g for 5 min and the supernatant obtained was then centrifuged for 10 min at 48,200 g. Membranes were washed twice, resuspended in 500 μ l of oxygenated Krebs–Ringer solution: 25 mM HEPES, pH 7.4; 118 mM NaCl, 4.8 mM KCl, 25 mM $NaHCO_3$, 1.3 mM $CaCl_2$, 1.2 mM $MgSO_4$, 1.2 mM KH_2PO_4 , 10 mM glucose, 100 μ M ascorbic acid, 50 μ g/ml leupeptin, 10 μ g/ml aprotinin, 2 μ g/ml soybean trypsin inhibitor, 0.04 mM PMSF. The concentration of membrane proteins was determined by the method of Bradford according to manufacturer's instruction. Membrane proteins (200 μ g) were incubated in Krebs's–Ringer solution with or without 1 μ M of dopamine for 5 min (total incubation volume of 500 μ l). The reaction was terminated by addition of Ca^{2+} -, Mg^{2+} -free Krebs's–Ringer solution containing 1 mM EDTA and centrifuged for 10 min at 48,200 g (at 4 °C). Tissues were then resuspended by sonicating for 10 s on ice in 0.25 ml of the immunoprecipitation buffer containing 100 mM HEPES, pH 7.5, 200 mM NaCl, 2 mM $MgCl_2$, 1 mM EDTA, 0.02% 2-mercaptoethanol, 50 μ g/ml leupeptin, 25 μ g/ml pepstatin A, 0.01 U/ml soybean trypsin inhibitor and 0.04 mM PMSF and solubilized by 0.5% digitonin, 0.2% Sodium cholate, 0.5% (v/v) Nonidet P-40 at 4 °C with end-over-end rotation for 1 h. Following dilution with 0.75 ml immunoprecipitation buffer and then clearing by centrifugation at 48,200 g for 10 min, the solubilized membrane proteins were immunoprecipitated with antibodies directed against $G\alpha$ s/olf (SC-383) or $G\alpha$ i (SC-7276) proteins (Santa Cruz Biotechnology, Santa Cruz, CA) using the procedure described previously [9,28]. The specificities of the anti- $G\alpha$ antibodies were extensively characterized and described previously [28]. While anti- $G\alpha$ i antibody cross-reacts mildly with $G\alpha$ o, anti- $G\alpha$ o did not precipitate appreciable D_1 - and D_2 receptors [9]. Solubilized tissues were incubated with 2 μ g anti- $G\alpha$ s/olf or - $G\alpha$ i for 2 h at 4 °C followed by an 1 h incubation with 25 μ l of Agarose-conjugated protein A/G (Santa Cruz Biotechnology). The suspension was centrifuged and washed twice with 1 ml immunoprecipitation buffer, the pellet obtained from each tube is suspended in 500 μ l of binding buffer (50 mM Tris–HCl, pH 7.5; 5 mM $MgCl_2$ and 1 μ M mesulergine, a 5-HT_{2C} receptor antagonist) since SCH23390 could also label the Gq-coupled 5-HT_{2C} receptors and incubated for 30 min at 30 °C with 1 nM [³H]SCH23390 (70.3 Ci/mmol, PerkinElmer, Boston, MA) or 2 nM

[³H]raclopride (60.5 Ci/mmol, PerkinElmer) for determination of $G\alpha$ s/olf-coupled DA D_1 -like receptors and $G\alpha$ i-coupled DA D_2 -like receptors, respectively. Nonspecific binding was defined by the addition of 1 μ M of unlabeled cis-(Z)-flupenthixol (for determination of the D_1 -like DA receptors) or l-sulpiride (for measurement of D_2 -like DA receptors). The reaction was terminated by addition of 9 ml of ice-cold binding buffer and immediately vacuum filtered over Whatman GF/C filters. The amount of radioactivity on filter was assessed by liquid scintillation spectrometry and specific [³H]SCH23390 or [³H]raclopride binding was determined.

To assess the expression levels of the $D_{1A}R$, D_2R and various $G\alpha$ proteins, Western blotting was conducted as described previously [9,28] using 25 μ g MFC or striatal lysate with antibodies specific for $D_{1A}R$ (SC-33660), D_2R (SC-5303), $G\alpha$ s/olf (SC-383), $G\alpha$ i (SC-7276), $G\alpha$ o (SC-387) and $G\alpha$ q/11 (SC-392), respectively.

Data were analyzed with mixed linear models in SPSS with D_1 -like striatum, D_2 -like striatum, D_1 -like MFC, D_2 -like MFC as separate dependent variables, sex and treatment as fixed factors, litter as random factor. All variables failed to show any differences in striatum (Fig. 1). In MFC, however, basal SCH 23,390 binding showed a significant decrease in adults treated with cocaine postnatally [$F(1,30) = 446, p < 0.001$] (Fig. 2). In addition, control males showed a lower level of SCH binding than did the control females [$F(1,14) = 78.4, p < 0.001$]. There was also a main effect for treatment for the stimulated D_1 -like effect [$F(1,30) = 107.24, p < 0.0001$] within both sexes with the cocaine-exposed rats showing a lower D_1 -like effect. This cocaine-mediated effect was not the result of altered total receptor populations or G protein levels since our Western blot data showed comparable D_{1A} and D_2 -like receptor concentrations as well as various $G\alpha$ protein levels in both striatum (data not shown) and medial frontal cortex (Table 1).

The reduced coupling of Gs to DA D_1 -like receptors in MFC observed in current study is compatible with that from our previous studies on prodynorphin mRNA expression [4] and rat brain glucose metabolism after postnatal cocaine exposure [3] as well as the observation of abnormal differentiation of cerebral cortical neurons in rabbit following cocaine exposure [10]. Since D_2 -like receptor coupling to Gi proteins in both striatum and MFC following cocaine exposure was unaffected, our data also match the observations on rabbit in which prenatal cocaine exposure impaired receptor-G protein coupling only in the D_1 - but not D_2 -like receptors [5]. Similar to these results [5], we also did not find changes in density of the D_{1A} and D_2 -like receptors as well as various $G\alpha$ proteins (Data not shown). These methods used enabled us to differentiate the coupling status of a specific receptor system, in this case Gs/olf-

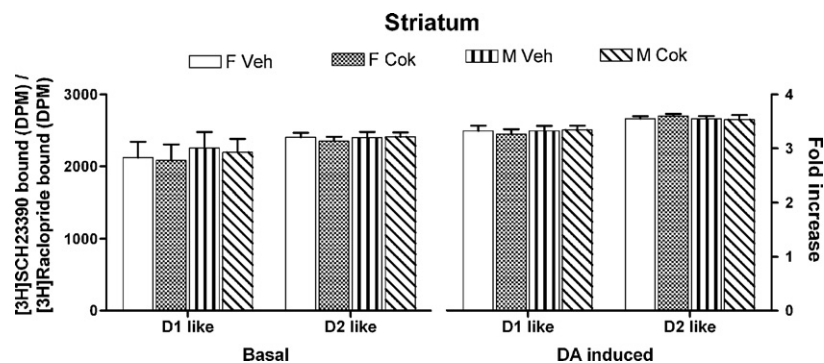


Fig. 1. Comparable association of the Gs/olf–DA D_1 -like receptors and Gi– D_2 -like receptors in striatum of PnD 60 male and female rats that have been exposed to cocaine during PnD 11–20. The effect of cocaine exposure during Pn 11–20 on Gs/olf– D_1 -like receptor and Gi– D_2 -like receptor coupling in striatum from PnD 60 male and female rats was assessed under basal (left) and 1 μ M DA-stimulated (right) conditions. The Gs/olf-coupled D_1 -like receptors and Gi-linked D_2 -like receptors were isolated together with Gs/olf and Gi, respectively, by anti- $G\alpha$ s/olf and - $G\alpha$ i and then assessed using [³H]SCH23390 and [³H]raclopride binding. No significant differences were observed in the levels of Gs/olf-coupled D_1 -like receptors and Gi-associated D_2 -like receptors under either basal or DA-stimulated conditions (mean + sem).

Download English Version:

<https://daneshyari.com/en/article/4348266>

Download Persian Version:

<https://daneshyari.com/article/4348266>

[Daneshyari.com](https://daneshyari.com)