



# Prediction of FMN-binding residues with three-dimensional probability distributions of interacting atoms on protein surfaces

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## HIGHLIGHTS

- First structure-based approach for prediction of protein–FMN interaction.
- Does not require evolutionary information for the prediction.
- Useful in annotating proteins structures of unknown function and computational protein models.

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## ABSTRACT

Flavin mono-nucleotide (FMN) is a cofactor which is involved in many biological reactions. The insights on protein–FMN interactions aid the protein functional annotation and also facilitate in drug design. In this study, we have established a new method, making use of an encoding scheme of the three-dimensional probability density maps that describe the distributions of 40 non-covalent interacting atom types around protein surfaces, to predict FMN-binding sites on protein surfaces. One machine learning model was trained for each of the 30 protein atom types to predict tentative FMN-binding sites on protein structures. The method's capability was evaluated by five-fold cross-validation on a dataset containing 81 non-redundant FMN-binding protein structures and further tested on independent datasets of 30 and 15 non-redundant protein structures respectively. These predictions achieved an accuracy of 0.94, 0.94 and 0.96 with the Matthews correlation coefficient (MCC) of 0.53, 0.53 and 0.65 respectively for the three protein structure sets. The prediction capability is superior to the existing method. This is the first structure-based approach that does not rely on evolutionary information for predicting FMN-interacting residues. The webserver for the prediction is available at <http://ismlab.genomics.sinica.edu.tw/>.

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## 1. Introduction

FMN is an essential cofactor in flavoproteins which are involved in (i) redox reactions in the energy producing metabolic pathways and (ii) non-redox reactions in which FMN acts as acid or base in the covalent-intermediate formation (Mansoorabadi et al., 2007; Serrano et al., 2012). Flavodoxin, which is one of the flavoproteins, is considered as one of the potential drug targets against microbial infections because it plays a critical role in the electron transfer

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system of pathogenic bacteria but not in mammals. *Helicobacter pylori* flavodoxin acts as an electron acceptor in pyruvate metabolic pathway, and thus inhibition of this protein can affect the bacterial survival (Cremades et al., 2005). Chorismate synthase is another FMN-binding protein involved in shikimate pathway (Macheroux et al., 1999) and is considered as a primary target in developing antibacterial therapeutics against tuberculosis (Fernandes et al., 2007). Hence, identification of FMN-binding proteins and binding site residues can aid in the drug discovery processes for antimicrobial therapeutics.

A computational method for predicting the FMN-binding residues on proteins would greatly facilitate defining FMN-binding sites on protein structures. Computational methods have been developed to predict FMN (Wang et al., 2012), flavin adenine dinucleotide (FAD) (Mishra and Raghava, 2010) and nicotinamide adenine dinucleotide (NAD) (Ansari and Raghava, 2010) binding

sites. These computational methods are reasonably successful in their respective predictions. Nevertheless, they are all sequence-based predictors relying on evolutionary information. Consequently, these methods may have difficulty in predicting binding site in orphan proteins, which shares very low sequence similarity with existing proteins. A structure-based method which does not rely on evolutionary information has yet to be developed. In this study, we have developed such a structure-based method for the prediction of FMN-interacting residues on protein surfaces.

This method uses machine learning approach to predict FMN-binding sites on protein surfaces by recognizing characteristic interacting atom distribution patterns associated with the FMN binding. The basic principle has been already applied to predict the protein–protein (Chen et al., 2012) and protein–carbohydrate (Tsai et al., 2012) interactions successfully. Here we extend this approach to predicting FMN-interacting residues. In the prediction, protein surface atoms were first categorized into 30 atom types and one machine learning model was trained for each of the atom types. The input attributes for the machine learning algorithm were normalized distance-weighted sum of three-dimensional probability density maps (PDMs) of 40 interacting atom types (30 atom types from protein, one from water and nine from FMN) on the protein surfaces. The PDMs around the query protein atoms for the protein interacting atom types and water have been described in previous publications (Chen et al., 2012; Tsai et al., 2012); the PDMs for the nine FMN interacting atom types were constructed with the protein–FMN interacting atom pairs from the dataset of 192 FMN–protein complex structures. The machine learning algorithm learned the patterns of the attributes to distinguish the binding atoms from the non-binding atoms on the protein surfaces. We evaluated our predictor performance on the training dataset P81 as well independent test sets P30 and P15 (Wang et al., 2012). The results indicate that our approach is the best method for predicting the FMN-binding sites on protein structures.

## 2. Materials and methods

### 2.1. Dataset

The training and test datasets except P15 were obtained from Wang et al. (2012). The author obtained 111 protein chains from PDB (Berman et al., 2002). Then they randomly selected 30 proteins (P30) for the independent test and the remaining 81 protein chains (P81) were used as a training set. For the P15, we extracted protein–FMN complexes from PDB and then used PISCES program (Wang and Dunbrack, 2003) to remove structures which has the sequence identity more than 10% with P81 and P30 datasets that finally yielded 15 protein chains.

### 2.2. Construction of three-dimensional probability density maps on protein surfaces

The detailed method for the PDMs construction has been discussed previously (Chen et al., 2012; Tsai et al., 2012; Yu et al., 2012). In brief, the interacting atom types from protein, water, and FMN are shown in Table 1. The PDMs for these interacting atom types were constructed with interacting atoms retrieved from the interacting atom database described previously (Chen et al., 2012; Tsai et al., 2012; Yu et al., 2012). The interacting atom database for protein–FMN interacting atom pairs was constructed with the dataset of 192 protein–FMN complexes.

**Table 1**  
Protein and FMN atom types.

| ID # | Atom type | Radius (Å) | Description                                   |
|------|-----------|------------|-----------------------------------------------|
| 1    | NH1       | 1.65       | Backbone NH                                   |
| 2    | C         | 1.76       | Backbone C                                    |
| 3    | CH1E      | 1.87       | Backbone CA (exc. Gly)                        |
| 4    | O         | 1.40       | Backbone O                                    |
| 5    | CH0       | 1.76       | Arg CZ, Asn CG, Asp CG, Gln CD, Glu CD        |
| 6    | CH1S      | 1.87       | Sidechain CH1: Ile CB, Leu CG, Thr CB, Val CB |
| 7    | CH2E      | 1.87       | Tetrahedral CH2 (except CH2P,CH2G) All CB     |
| 8    | CH3E      | 1.87       | Tetrahedral CH3                               |
| 9    | CR1E      | 1.76       | Aromatic CH (except CR1W, CRHH, CR1H)         |
| 10   | OH1       | 1.40       | Alcohol OH (Ser OG, Thr OG1, Tyr OH)          |
| 11   | OC        | 1.40       | Carboxyl O (Asp OD1, OD2, Glu OE1, OE2)       |
| 12   | OS        | 1.40       | Sidechain O: Asn OD1, Gln OE1                 |
| 13   | CH2G      | 1.87       | Gly CA                                        |
| 14   | CH2P      | 1.87       | Pro CB, CG, CD                                |
| 15   | NH1S      | 1.65       | Sidechain NH: Arg NE, His ND1, NE1, Trp NE1   |
| 16   | NC2       | 1.65       | Arg NH1, NH2                                  |
| 17   | NH2       | 1.65       | Asn ND2, Gln NE2                              |
| 18   | CR1W      | 1.76       | Trp CZ2, CH2                                  |
| 19   | CY2       | 1.76       | Tyr CZ                                        |
| 20   | SC        | 1.85       | Cys S                                         |
| 21   | CF        | 1.76       | Phe CG                                        |
| 22   | SM        | 1.85       | Met S                                         |
| 23   | CY        | 1.76       | Tyr CG                                        |
| 24   | CW        | 1.76       | Trp CD2, CE2                                  |
| 25   | CRHH      | 1.76       | His CE1                                       |
| 26   | NH3       | 1.50       | Lys NZ                                        |
| 27   | CR1H      | 1.76       | His CD2                                       |
| 28   | C5        | 1.76       | His CG                                        |
| 29   | N         | 1.65       | Pro N                                         |
| 30   | C5W       | 1.76       | Trp CG                                        |
| 31   | HOH       | 1.40       | Water                                         |
| 32   | P         | 2.10       | Phosphate                                     |
| 33   | O1O2      | 1.68       | Phosphate oxygen                              |
| 34   | N1N9      | 1.82       | Base                                          |
| 35   | N         | 1.82       | ring                                          |
| 36   | C         | 1.90       | ring                                          |
| 37   | N         | 1.82       | link                                          |
| 38   | O         | 1.68       | link                                          |
| 39   | C.3       | 1.90       | Sp3 carbon                                    |
| 40   | O.3       | 1.68       | Sp3 oxygen                                    |

The protein atom types 1–31 have been previously defined by Laskowski et al. (1996) with minor modifications. The atom types 32–40 were defined in this work for FMN molecule.

### 2.3. PDM-based attributes as inputs for machine learning algorithms

Protein surface atoms were categorized into 30 protein atom types (Table 1, 1–30), and one machine learning model was trained for each of the atom types. The input attributes for each of the protein atom  $i$  ( $a_{i,j}$  ( $j=1,41$ )): 40 attributes from the 40 interacting atom type PDMs plus one attribute from geometry) for each of the machine learning models were calculated from the PDMs on the protein surface and from the geometry of the protein surface as the following: for each atom  $i$  on the surface of the query protein (solvent accessible surface area of atom  $i > 0$ ), the PDM values associated with the grids within 5 Å radius centered at the atom are summed in the following equation:

$$S_{ij} = \sum_k^{r_{ik} \leq 5 \text{ Å}} g_{kj} \quad (1)$$

where  $S_{ij}$  is the PDM sum for interacting atom type  $j$  at atom  $i$ ;  $r_{ik}$  is the distance between atom  $i$  to a grid point  $k$ ; and  $g_{kj}$  is the PDM value of interacting atom type  $j$  at grid point  $k$ .  $A_{ij}$  ( $j=1,40$ ) associated with each atom  $i$  was calculated with the following equation:

$$A_{ij} = S_{ij} + \sum_k^{d_{ik} \leq 10 \text{ Å}} S_{kj} \times d_{ik}^{-2} / \sum_n^{d_{in} \leq 10 \text{ Å}} d_{in}^{-2} \quad (2)$$

where  $S_{ij}$  is defined in Eq. (1);  $d_{ik}$  is the distance between atom  $i$  and atom  $k$ . The attribute set ( $a_{i,j}$  ( $j=1,40$ )) for the machine learning

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