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Temperature-dependent models of Zannichellia palustris seed germination for application in aquatic systems

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A B S T R A C T

The temperature-dependent germination rates of Zannichellia palustris, a native submersed aquatic plant from the spring-fed Fall River in Northern California, were examined with linear models and with the non-linear Logan Type III biophysical model. The Logan Type III model, which previously had not been applied to predicting seed germination, provided superior fit for estimating germination rates across eight constant temperatures (5–40 \degree C) and across a range of subpopulations representing the entire examined seed population. The Logan model was particularly successful at estimating seed germination rates at sub-optimal temperatures (5–25 ◦C) that occur under field conditions and at temperatures near the base and maximum germination temperatures. Based on the Logan Type III model, a computer simulation was developed using Time Varying Distributed Delays as implemented in the USDA-ARS Hermes (Hierarchical Environment for Research Modeling of Ecological Systems) simulation modeling system, tested across fluctuating temperatures in the laboratory, and applied to predict seed germination of Z. palustris under actual field conditions over an entire year in differing sections of the Fall River. The Logan Type III biophysical model and the Hermes simulation system are shown to be useful for predicting germination under fluctuating temperature conditions in the field and in seed populations that do not exhibit a linear germination response.

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1. Introduction

Assessment and prediction of key biological processes can be of extreme importance to land and waterway managers as they work to restore natural systems ([Frissell](#page--1-0) [and](#page--1-0) [Bayles,](#page--1-0) [1996;](#page--1-0) [Palmer](#page--1-0) et [al.,](#page--1-0) [1997\).](#page--1-0) Such is the case with the on-going restoration of the Fall River, California's largest spring-fed wild trout fishery (see [http://fallriverconservancy.org/programs/restoration/\)](http://fallriverconservancy.org/programs/restoration/). Threats to the river include a combination of sedimentation events [\(Spencer](#page--1-0) [and](#page--1-0) [Ksander,](#page--1-0) [2002\)](#page--1-0) and invasion by Myriophyllum spicatum (Eurasian watermilfoil) ([Spencer](#page--1-0) [and](#page--1-0) [Carruthers,](#page--1-0) [2012\).](#page--1-0) Central to the restoration effort is Zannichellia palustris (horned pondweed), a globally distributed species ([van](#page--1-0) [Vierssen,](#page--1-0) [1982;](#page--1-0) [Lombardi](#page--1-0) et [al.,](#page--1-0) [1996;](#page--1-0) [Greenwood](#page--1-0) [and](#page--1-0) [DuBowy,](#page--1-0) [2005\)](#page--1-0) and the most abundant native submersed macrophyte in the Fall River [\(Spencer](#page--1-0) [and](#page--1-0) [Ksander,](#page--1-0) [1998;](#page--1-0) [USDA,](#page--1-0) [NRCS,](#page--1-0) [2013\).](#page--1-0)

Understanding the germination dynamics of Z. palustris is particularly important, as this annual plant regenerates each season from newly produced seeds and from those within the seed-bank [\(van](#page--1-0) [Vierssen,](#page--1-0) [1982\).](#page--1-0) To aid waterway managers, it is important to understand seed production and the timing of germination in spe-cific waterways, such as the Fall River. [Bytnerowicz](#page--1-0) [and](#page--1-0) [Carruthers](#page--1-0) [\(in](#page--1-0) [review\)](#page--1-0) conducted germination studies for Z. palustris from the Fall River under variable light and temperature conditions. Results denote a primarily temperature-dependent germination response, an extensive variability of the population response within and between test temperatures, and an absence of a dormancy period in the tested seed population. These results are supported by the observation that temperature is the most important environmental factor controlling the rate and overall germination percentage of non-dormant seeds across plant species [\(Bewley](#page--1-0) [and](#page--1-0) [Black,](#page--1-0) [1982;](#page--1-0) [Garcia-Huidobro](#page--1-0) et [al.,](#page--1-0) [1982a;](#page--1-0) [Steinmaus](#page--1-0) et [al.,](#page--1-0) [2000\).](#page--1-0) The work presented here, was designed to further address the impact of temperature on Z. palustris seed germination and to develop dynamic means of assessing and forecasting seed germination timing in the natural environment.

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Germination response across temperatures has most frequently been described with thermal-time models that rely on linear relationships between germination rate and temperature ([Garcia-](#page--1-0)Huidobro et [al.,](#page--1-0) [1982a;](#page--1-0) [Scott](#page--1-0) et [al.,](#page--1-0) [1984;](#page--1-0) [Washitani,](#page--1-0) [1985;](#page--1-0) [Covell](#page--1-0) et [al.,](#page--1-0) [1986;](#page--1-0) [Ellis](#page--1-0) et [al.,](#page--1-0) [1986;](#page--1-0) [Ellis](#page--1-0) [and](#page--1-0) [Butcher,](#page--1-0) [1988\).](#page--1-0) Thermal-time models assume that germination rate increases linearly from a base temperature (T_b) to an optimal temperature (T_0) and decreases linearly from T_0 to a maximum temperature (T_m) . They have been widely used to compare seeds from different species and climates ([Garcia-Huidobro](#page--1-0) et [al.,](#page--1-0) [1982a;](#page--1-0) [Covell](#page--1-0) et [al.,](#page--1-0) [1986;](#page--1-0) [Ellis](#page--1-0) et [al.,](#page--1-0) [1986;](#page--1-0) [Ellis](#page--1-0) [and](#page--1-0) [Butcher,](#page--1-0) [1988;](#page--1-0) [Hardegree,](#page--1-0) [2006\)](#page--1-0) and to make predictions on the timing of seed germination in nature [\(Alvarado](#page--1-0) [and](#page--1-0) [Bradford,](#page--1-0) [2002\).](#page--1-0)

The use of thermal-time models for describing temperaturedependent germination rates is a powerful tool for germination response that is linear. However, other modeling schemes may be equally or more appropriate for seed populations that do not fit the linear shape constraints ([McDonald,](#page--1-0) [2002;](#page--1-0) [Hardegree,](#page--1-0) [2006;](#page--1-0) [Bytnerowicz](#page--1-0) [and](#page--1-0) [Carruthers,](#page--1-0) [in](#page--1-0) [review\).](#page--1-0) In such seed populations, linear regression can be especially ill-suited for fitting lower and upper developmental thresholds that are typically determined by extrapolating a straight line through the temperature axis. It is in these extreme regions of these response curves where development is often underestimated at lower temperatures and overestimated at higher temperatures [\(Hilbert](#page--1-0) [and](#page--1-0) [Logan,](#page--1-0) [1983;](#page--1-0) [Wagner](#page--1-0) et [al.,](#page--1-0) [1984\).](#page--1-0) We suggest the use of the Logan Type III biophysical model as an alternative for such seed populations. Logan models have been frequently used to estimate temperaturedependent developmental rates, and lower and upper thresholds of development in the discipline of entomology [\(Logan](#page--1-0) et [al.,](#page--1-0) [1976;](#page--1-0) [Sharpe](#page--1-0) [and](#page--1-0) [DeMichele,](#page--1-0) [1977;](#page--1-0) [Hilbert](#page--1-0) [and](#page--1-0) [Logan,](#page--1-0) [1983;](#page--1-0) [Wagner](#page--1-0) et [al.,](#page--1-0) [1984;](#page--1-0) [Wermelinger](#page--1-0) [and](#page--1-0) [Seifert,](#page--1-0) [1988;](#page--1-0) [López](#page--1-0) et [al.,](#page--1-0) [2001;](#page--1-0) [Roy](#page--1-0) et [al.,](#page--1-0) [2002;](#page--1-0) [Herrera](#page--1-0) et [al.,](#page--1-0) [2005\).](#page--1-0) The model, originally developed by [Logan](#page--1-0) et [al.\(1976\),](#page--1-0) and improved by [Hilbert](#page--1-0) [and](#page--1-0) [Logan](#page--1-0) [\(1983\),](#page--1-0) is a hybrid function composed of two subcomponents, each describing different phases of a temperature-dependent developmental rate curve. The ascending phase of the curve is represented by a sigmoid function, while the descending phase is represented by an exponential decay function. Combined, the two processes form the non-linear Logan Type III equation, as follows:

$$
r(T) = \psi \left\{ \left(\frac{(T - T_{b'})^2}{(T - T_{b'})^2 + D^2} \right) - \exp \left(\frac{-(T_{m'} - (T - T_{b'}))}{\Delta T} \right) \right\}, \quad (1)
$$

where $r(T)$ is the germination rate (1/d) at T, T is temperature (\degree C), $T_{b'}$ is the arbitrary base temperature, $T_{m'}$ is the arbitrary lethal maximum temperature, ΔT is the width of high temperature breakdown, and ψ and D are estimated constants that alter the shape of their fitted "biophysical" function.

Independent of the method used to estimate the temperaturedependent developmental curves in the laboratory, relating such functions to actual event prediction in the field may be more complicated due to the non-linearity of the relationship and the time varying nature of temperature patterns in the environment. That is, temperatures are rarely constant, fluctuating both diurnally and seasonally in most habitats and their combined effects on development are not necessary additive as suggested in simple linear models. Thus more dynamic methods of prediction such as non-linear thermal accumulations and/or event simulations are often required to estimate complex temperature-dependent processes where the stimulus variable changes continuously with time ([Legaspi](#page--1-0) et [al.,](#page--1-0) [1997\).](#page--1-0)

Here we propose to test aggregate simulation methods ([Forrester,](#page--1-0) [1961;](#page--1-0) [Manetsch,](#page--1-0) [1976\)](#page--1-0) that are especially appropriate for linking dynamic and non-linear environmental variables, such as temperature, with functional response equations (such as the Logan Type III biophysical model). We propose to simulate these rate processes using "Time Varying Distributed Delays", TVDDs [\(Manetsch,](#page--1-0) [1976;](#page--1-0) [Carruthers](#page--1-0) et [al.,](#page--1-0) [1986,](#page--1-0) [1992;](#page--1-0) [Larkin](#page--1-0) et [al.,](#page--1-0) [2000\).](#page--1-0) TVDDs are commonly used simulation components that allow complex biological processes such as phenological development and overall population response levels to be predicted accurately under changing environmental conditions, using well known and highly tested simulation routines developed by system scientists and engineers. By altering just two parameters, DEL (the mean delay time) and k (a distribution shape parameter), TVDDs can simulate a wide range of delay processes like seed germination or age-specific organism development with realistic means and variances, as those observed under both constant and cycling temperature conditions [\(Abkin](#page--1-0) [and](#page--1-0) [Wolf,](#page--1-0) [1976\).](#page--1-0) The temperature-dependent maturation times (DEL, duration of germination period for Z. palustris seeds) were calculated as:

$$
DEL = \frac{1.0}{\psi\{(T - T_{b'})^2/[(T - T_{b'})^2 + D^2] - \exp[-(T_{m'} - (T - T_{b'})]/\Delta T]\}},
$$
\n(2)

that is, the inverse of the Logan Type III biophysical model. The second parameter value (k) is a characterization of the variance of the seed germination response at each experimental temperature (converting developmental rate to germination time).

In a TVDD, the simulation of the full population response is achieved by calculating output that follows the Erlang family of probability density functions using kth-order differential equations (see [Larkin](#page--1-0) et [al.,](#page--1-0) [2000](#page--1-0) for an in-depth discussion of TVDDs and their application in simulating biological processes). This approach produces distributed output according to the Erlang law, which is used to calculate a series of response curves that range from an exponential decay curve $(k = 1)$, to highly skewed distributions $(k=2-15)$, to near to normal distributions $(k>20)$, on to a discrete spike or pulse as k approaches infinity. In each situation the value of k describes the distribution of the response data that can be controlled through changing the order of the differential equation involved in the simulation [\(Manetsch,](#page--1-0) [1976;](#page--1-0) [Larkin](#page--1-0) et [al.,](#page--1-0) [2000\).](#page--1-0) Values of k are directly calculable from observed data using the following relationship:

$$
k = \frac{DEL^2}{variance} \tag{3}
$$

Values of k tend to be fairly constant for a population, as plants (as well as insects and fungi) often have a relatively stable relationship between the mean developmental time and the variance observed across treatment conditions, as reported by [Garcia-Huidobro](#page--1-0) et [al.](#page--1-0) [\(1982a\)](#page--1-0) specifically for seed germination.

In this study, we were interested in using and comparing a variety of statistical seed germination models for our experimental system (Z. palustris in the Fall River of California), based on laboratory implemented constant temperature experiments [\(Bytnerowicz](#page--1-0) [and](#page--1-0) [Carruthers,](#page--1-0) [in](#page--1-0) [review\).](#page--1-0) First we examine the most appropriate means to describe temperature-dependent seed germination rates for Z. palustris, and then we apply appropriate statistical models as the bases for constructing a computer simulation to estimate germination times under highly controlled fluctuating temperature conditions and then under actual field conditions where temperature changes dynamically within individual days and across the entire growth season. We are further interested in assessing changes in germination response through time and at different geographical locations along the course of the river.

2. Materials and methods

2.1. Seed collection

Mature Z. palustris plants, from which newly produced seeds were harvested, were collected from the Fall River in the autumn

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