



The effect of NOM to TiO₂: interactions and photocatalytic behavior



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ABSTRACT

Natural organic matter (NOM) is ubiquitous in aquatic environment, which plays a predominant role in the sorption of pharmaceuticals onto the TiO₂ nanoparticles. It is a matter of concern whether NOM could act as a surface sensitizer of TiO₂ or not. In this context, the role of NOM is investigated for the photocatalytic degradation of carbamazepine (CBZ) using TiO₂. Four different ratios of NOM:TiO₂ were used varying from 400 μg g⁻¹ to 400 mg g⁻¹. The findings reveal that small amounts of NOM could enhance the TiO₂ efficiency up to 8%. Electron paramagnetic spectroscopy (EPR), along with size exclusion chromatography with dissolved organic carbon detection (SEC-OCD) and nuclear magnetic resonance spectroscopy (NMR) reveal the occurring mechanism. TiO₂ binds small molecular size fractions of NOM and breaks aromatic bonds of adsorbed NOM transforming it to stranded alkyl groups. This modified TiO₂ bears a significant amount of electrons (e⁻) and lesser holes (h⁺) than the purified TiO₂ and when irradiated, produces hydroxyl radicals which degrade CBZ.

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1. Introduction

Titanium oxide is used in a wide variety of technological applications where surface properties play a role. It is used in heterogeneous catalysis, as photocatalyst; in solar cells for the production of hydrogen and electric energy, as white pigment, as corrosion-protective or optical coating, as spacer material in magnetic spin-value systems, and it is important in earth sciences [1].

By far, the most actively pursued applied research on TiO₂ is its use for photo-assisted degradation of organic molecules. TiO₂ is a semiconductor where the electron–hole pair, created upon irradiation with sunlight, became separated and the resulting charge carriers migrate to the surface. These react with adsorbed water and oxygen to produce radical species, which attack any adsorbed organic molecule and can, ultimately, lead to their complete decomposition into CO₂ and H₂O [2].

Photocatalysis based on TiO₂ has been intensively investigated in order to improve photocatalytic efficiency for different applications such as decomposition of various organic pollutants in aqueous media [3]. Studies on the doping of non-metal and transition-metal ions into TiO₂ have become attractive in the area of photocatalysis [4].

Natural organic matter (NOM) is ubiquitous in aquatic environment and plays a predominant role in the sorption of toxic substances onto the TiO₂ nanoparticles (NPs) [5]. Once released in the environment, NPs will inevitably interact with abundant NOM [6]. Fulvic acid (FA) and humic acid (HA) are the main constituents of NOM from water resources responsible for water color and odor [7], and also responsible for membrane fouling in water treatment [8]. It is reported that TiO₂ has a strong affinity for HA [9–11], especially at a low pH value. Previous investigations showed that certain groups of organic compounds, such as carboxylic acids, strongly adsorb onto the TiO₂ surface at low pH values whilst others, such as alcohols and saturated long-chain aliphatic compounds, do not [12].

The research on the NOM/TiO₂ interactions was so far limited to the photodegradation of NOM by TiO₂ [7,9–11,13–16]. However, it is reported that the removal of Cr(IV), by using TiO₂ photocatalysis, was greatly enhanced when the system contained both HA and Cr(VI) [16] and also that HA coating enhanced the adsorption of Cd(II) to nano-TiO₂ [6].

It is a matter of concern whether NOM could act as a TiO₂ surface sensitizer or not. So far, only activated carbon has been used as a catalyst support for the photocatalytic degradation [17]. Carbon doping has been shown to be more active than doping with nitrogen [18]. For example, Khan et al. [19] first demonstrated the high photoactivity of carbon-doped TiO₂ in the photoelectrochemical context by measuring the rate of water splitting to H₂ and O₂. Previously, it was reported by Doll et al. that low concentrations of NOM could accelerate the degradation of carbamazepine

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due to the photochemical formation of reactive species by NOM, but at higher concentrations of NOM, the acceleration of the degradation decreased [20]. It is suggested that for the best evaluation of the photocatalytic degradation of NOM by TiO₂, NOM should be characterized in detail [13]. Therefore, the aim of this work was to evaluate the photodegradation efficiency of NOM-modified TiO₂ to CBZ, by using a well-characterized lignite humic acid, as a model NOM, to elucidate the effect of NOM to the photocatalytic behavior of TiO₂.

2. Experimental

2.1. Reagents

Millipore Milli-Q Water (resistivity = 18 MΩ cm) and P25 titanium oxide (nano-TiO₂) (Evonik Degussa GmbH, Düsseldorf, Germany) with purity >99.5% were used for all the experiments. The pH adjustments were carried out using NaOH and HNO₃ from Fluka. The fairly homogeneous humic acid, used as NOM model, is a lignite humic acid (LHA2) previously characterized by Drosos et al. [21] and also named L_{parental}HA in [22]. Carbamazepine (99%) was obtained from Sigma Aldrich.

2.2. Purification

In order to obtain a completely DOC-free TiO₂, a purification step was required by baking P25 TiO₂ at 500 °C for 60 min. An aqueous solution (Milli-Q water) of the baked TiO₂ (20 g L⁻¹) was irradiated for 60 min, under a Solar UV Simulator (Oriel Corp., Stratford, CT, USA) with additional WG 295 filters (6 mm, Schott Glaswerke, Germany). The scheme and detailed description of the solar simulator used for the irradiation experiments was given by Doll et al. [20]. The radiation source was a 1000-W Xe short-arc lamp, which was turned on at least 30 min prior to irradiation in order to insure a constant photon flux.

The irradiated TiO₂ solution was centrifuged at 4000 rpm for 15 min. The filtered TiO₂ was placed into an oven at 60 °C for 24 h, in order to remove moisture completely. The purified TiO₂ was then grained to obtain fine powder. The supernatant was collected and measured by total organic carbon detection (TOC) and size exclusion chromatography with organic carbon detector (SEC-OCD) after filtration using 0.45 μm filter supor 450.

2.3. Samples preparation

One gram of purified TiO₂ was placed in 50 ml of an LHA2 solution at pH 5, stirred overnight and centrifuged at 4000 rpm for 15 min. The supernatant was collected, filtered using 0.45 μm filter supor 450 and measured by total organic carbon detection (TOC) and size exclusion chromatography with organic carbon detector (SEC-OCD). The different LHA2 solutions used were 8000, 800, 80, and 8 mg L⁻¹ respectively, giving four TiO₂-LHA2 ratios. By using 50 ml of Milli-Q instead of the LHA2 solution, the TiO₂ obtained was used as a blank (Blank TiO₂). The materials were placed into an oven at 60 °C for 24 h, in order to remove moisture completely and were grained to obtain a fine powder. The four LHA2-modified TiO₂ samples are named here as it follows: (1) LT04 for the LHA2:TiO₂ ratio of 400 μg g⁻¹, (2) LT4 for the 4 mg g⁻¹ ratio, (3) LT40 for the 40 mg g⁻¹ ratio and (4) LT400 for the 400 mg g⁻¹ ratio.

All CBZ samples were prepared by dissolving CBZ in Milli-Q water, stirred for 6 h and placed in an ultrasonic bath for 15 min to ensure complete dissolution.

The catalysis samples were prepared by adding 1 g L⁻¹ LHA2-modified TiO₂ material in 10 mg L⁻¹ pharmaceutical solution at pH 7. The sample volume was 50 ml. Samples were irradiated by the Solar UV Simulator introduced by Doll et al. [20].

All samples were stirred and irradiated simultaneously from above by a homogenous light field. During photoreaction, samples were collected at different time intervals for analysis. Before irradiation, adsorption was allowed to take place for 30 min in the dark.

2.4. EPR spectroscopy

Electron paramagnetic resonance (EPR) spectra were recorded at liquid-nitrogen temperature (77 K) with a Bruker ER200D spectrometer equipped with an Agilent 5310A frequency counter. Typically, adequate signal-to-noise ratio was obtained after 20–40 scans. Samples were cooled to 77 K and illuminated in the cavity while spectra were recorded. A 450 W lamp (Oriel model 66929) was used as illumination source. EPR samples were prepared by pasting the wet pellet of each material into 5 mm suprasil (Willmad Co.) quartz tube with no further treatment. *g*-values were calibrated versus DPPH, *g* = 2.0036, as described earlier [23].

2.5. SEC-OCD chromatography and TOC analysis

Size exclusion chromatography coupled with organic carbon and UV (254 nm) detector used a TSK HW 50S Column. The mobile phase was 1.5 g L⁻¹ Na₂HPO₄·2H₂O and 2.5 g L⁻¹ KH₂PO₄ with 1 mL min⁻¹ flow rate, followed by acidification using 5 mL L⁻¹ phosphoric acid 85% with a flow rate of 0.5 mL min⁻¹ under nitrogen 5.0 (28 L h⁻¹) with 0.5–1 bar pressure. The contact time in the reactor was 80 s, 60 s UV radiation and rotation time of the reactor 400 rpm. For the determination of the apparent molecular size, calibration using PEGs was performed. In detail, V_o was determined by blue dextran (2 × 10⁶ Da), V_p by methanol and the calibration PEG standards used varied in molecular weight (200 kDa, 100 kDa, 40 kDa, 20 kDa, 10 kDa, 6 kDa, 4 kDa, 1550 Da, 1000 Da, 400 Da, 200 Da). DOC concentrations were measured both by SEC-OCD and a TOC 5000 Carbon Analyzer (Shimadzu).

2.6. NMR spectroscopy

Solid state ¹³C NMR spectra of powder samples were recorded on a Bruker NMR spectrometer at a resonance frequency of 400 MHz, using a Ramped-Cross Polarization MAS with a spinning speed of 6.8 kHz [24]. A contact time of (1 ms) and a pulse delay of (400 ms) were used. A ramped ¹H-pulse decreasing the power from 100 to 50% was used to circumvent spin modulation of Hartmann–Hahn conditions [24]. At least 50,000 single scans were collected for each sample.

2.7. HPLC analysis

The concentration of CBZ was measured by high-performance liquid chromatography (HPLC) using an HP 1100 HPLC system (column RP 18e, 5 μm; mobile-phase acetonitrile: water 60:40; 1 mL min⁻¹) with detection at 280 nm by a UV detector.

3. Results and discussion

3.1. TOC and samples

LHA2 was proved to contain 44% of carbon both by elemental analysis in [22] and TOC analysis. More information about LHA2 is summarized in Table 1. The adsorbed carbon on 1 g of purified TiO₂ was 160 μg L⁻¹ for LT04 and 670 μg L⁻¹ for LT4, close to the concentration found in drinking water. The adsorbed carbon for LT40 was 8 mg L⁻¹, and for LT400 was 44 mg L⁻¹, which are values,

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