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# Effect of Elevated CO<sub>2</sub> on the Growth and Macronutrient (N, P and K) Uptake of Annual Wormwood (*Artemisia annua* L.)

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#### ABSTRACT

Annual wormwood (Artemisia annua L.) is the only viable source of artemisinin, an antimalarial drug. There is a pressing need to optimize production per cultivated area of this important medicinal plant; however, the effect of increasing atmospheric carbon dioxide (CO<sub>2</sub>) concentration on its growth is still unclear. Therefore, a pot experiment was conducted in a free-air CO<sub>2</sub> enrichment (FACE) facility in Yangzhou City, China. Two A. annua varieties, one wild and one cultivated, were grown under ambient (374 µmol mol<sup>-1</sup>) and elevated (577 µmol mol<sup>-1</sup>) CO<sub>2</sub> levels to determine the dry matter accumulation and macronutrient uptake of aerial parts. The results showed that stem and leaf yields of both A. annua varieties increased significantly under elevated CO<sub>2</sub> due to the enhanced photosynthesis rate. Although nitrogen (N), phosphorus (P), and potassium (K) concentrations in leaves and stems of both varieties decreased under elevated CO<sub>2</sub>, total shoot N, P, and K uptake of the two varieties were enhanced and the ratios among the concentrations of these nutrients (N:P, N:K, and P:K) were not affected by elevated CO<sub>2</sub>. Overall, our results provided the evidence that elevated CO<sub>2</sub> increased biomass and shoot macronutrient uptake of two A. annua varieties.

Key Words: artemisinin, biomass, free-air CO<sub>2</sub> enrichment (FACE), medicinal plant, photosynthesis

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# INTRODUCTION

The increasing atmospheric carbon dioxide  $(CO_2)$ concentration (IPCC, 2014) has led to many hypotheses and experiments on the possible effects of elevated  $CO_2$  on the growth and development of plants (Ainsworth and Long, 2005). The current ambient level of atmospheric  $CO_2$  is one of the limiting factors for photosynthesis; therefore, any rise in  $CO_2$  above this level would have the potential to increase the rate of photosynthesis, decrease photorespiration (especially in  $C_3$  plants), and generally enhance the levels of carbon (C) fixation and dry matter (DM) accumulation (Poorter, 1993; Ainsworth and Long, 2005; Booker et al., 2007; Högy et al., 2010; Kumari et al. 2013). Because there is no concomitant increase in nutrient uptake with DM accumulation, elevated  $CO_2$  normally has an opposite effect on nutrient concentrations in plants, and reductions in nutrient levels would be observed (Loladze, 2002; Li *et al.*, 2013). These effects of elevated  $CO_2$  have been studied in many crops, such as rice, wheat, soybean, and potato (Rogers *et al.*, 2004; Wu *et al.*, 2004; Long *et al.*, 2006). However, there are also other opinions that  $CO_2$  fertilization effect on plant growth is not universal, especially for trees (Long *et al.*, 2006; Ellsworth *et al.*, 2012; Bader *et al.*, 2013); *e.g.*,  $CO_2$ -induced growth stimulation was not found in deciduous forest trees (Bader *et al.*, 2013), which may be offset by other environmental factors such as nutrient limitation in soils (Norby *et al.*, 2011).

Annual wormwood (*Artemisia annua* L.), a traditional Chinese herb, has been widely cultivated worldwide to meet the increasing demands for artemisinin since the World Health Organization (WHO) recommended artemisinin-based combination therapy (ACT) to treat multi-drug-resistant malaria in 2001 (Wyk and Wink, 2004). Synthetic production is not economically viable, and the only commercially feasible source of

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artemisinin is leaves of *A. annua*. The worldwide cultivated area of *A. annua* has expanded several times (White, 2008) and reached to 12 000 ha (Jessing *et al.*, 2013). Various approaches to conventional breeding and agronomic practices have been tried to increase artemisinin concentration or leaf biomass of *A. annua* (Weathers *et al.*, 2005; Aquil *et al.*, 2009; Zhang *et al.*, 2009).

Given the public health importance of this medicinal plant and the pressing need to optimize its production (Ferreira, 2007), the impact of projected  $CO_2$  increases on the growth of A. annua must be known. Meanwhile, in view of down-regulation of plant growth responses induced by nutrient limitation found in some experiments (Oren et al., 2001; Reich and Hobbie, 2013), it is also important to understand the macronutrient demands of A. annua under elevated  $CO_2$ . However, few studies have answered these questions. Thus, we conducted an experiment to determine whether elevated CO<sub>2</sub> could increase the leaf biomass and nutrient accumulation of a wild variety and a cultivated variety of A. annua. This study would be helpful to predict future yields and fertilizer needs of A. annua.

#### MATERIALS AND METHODS

## Experiment description

This study was conducted at the free-air  $CO_2$  enrichment (FACE) facility located at Zhongcun Village (119°42'0" E, 32°35'5" N), Yangzhou City, Jiangsu Province, China. Details of the design, operation, and performance of the FACE facility used in this study can be found in Liu et al. (2002). The experiment was laid out in a split plot design with elevated and ambient  $CO_2$  levels as the main treatments, which were split into subplots of two cultivars. Three rings (replications) were for elevated  $CO_2$  and three rings (replications) for ambient  $CO_2$ . Three pots per variety were placed within each of the three elevated CO<sub>2</sub> and ambient  $CO_2$  rings. The target  $CO_2$  concentration of the elevated  $CO_2$  rings was controlled to approximately 200  $\mu$ mol mol<sup>-1</sup> above ambient during day and night by a computer system that factored in ambient  $CO_2$  concentration, wind direction, wind speed, and canopy height during day changes. Average daytime  $CO_2$  concentrations at canopy height during the experiment were 374 and 577  $\mu$ mol mol<sup>-1</sup> for the ambient and elevated CO<sub>2</sub> rings, respectively. Elevated CO<sub>2</sub> concentrations were within 80% of the set point, > 90% of those in each C. W. ZHU et al.

year.

Seeds of cultivated A. annua cv. Youyang were obtained from the Institute of Chinese Medicine in Chongqing City, China. Seeds of a wild A. annua variety were collected from populations in Wulong (107°45′42″ E, 29°21′17″ N), Chongqing City, China. The seeds were soaked in 2% (volume/volume) sodium hypochlorite for 15 min and then washed with distilled water. After germination in vermiculite-peat medium, two or three uniform seedlings (about 5-cm height) of each variety were transplanted into pots (25cm diameter and 25-cm height) containing 4 kg soil on June 15, 2013 and then thinned to one plant per pot on June 30, 2013. The soil was locally obtained and classified as Shajiang Aquic Cambisol according to Chinese Soil Taxonomy, with 1.16 g cm<sup>-3</sup> bulk density, 54% total porosity,  $18.4 \text{ g kg}^{-1}$  organic C,  $1.45 \text{ g kg}^{-1}$  total nitrogen (N), and 0.63 g kg<sup>-1</sup> total phosphorus (P) (as  $P_2O_5$ ). Before the transplanting, the soils in the pots were mixed with 0.43 g urea, 0.39 g  $KH_2PO_4$ , and  $0.13 \text{ g K}_2\text{SO}_4$  per kg soil as basal fertilizers. The plants were watered with tap water to maintain soil moisture at about 70%–80%. The wild and cultivated varieties were harvested at initial anthesis on September 14 and September 19, 2013, respectively.

## Measurement of plant growth

Plant heights were measured from plant base to top of stem prior to harvest and branch numbers were counted. At harvest, shoots were separated into stems and leaves, washed with distilled water, and dried in an oven at 75 °C for 72 h to determine the dry weight.

#### Measurement of photosynthesis parameters

Photosynthesis rate, stomatal conductance, and transpiration rate of leaves were measured with an LI-6400 portable photosynthesis system (LI-COR, Lincoln, USA) at 1800 µmol m<sup>-2</sup> s<sup>-1</sup> light intensity at initial anthesis of plants. Leaf temperature in the leaf chamber was set to 30 °C, with humidity in the leaf chamber set to that in the field. Leaf chamber CO<sub>2</sub> concentration was set to 580 µmol mol<sup>-1</sup> for the elevated CO<sub>2</sub> level and 380 µmol mol<sup>-1</sup> for the ambient CO<sub>2</sub> level, with the flow rate set to 500 µmol s<sup>-1</sup>. All measurements were taken during the periods of 10:00– 11:30 a.m.

#### Analysis of plant macronutrients

After being ground and passed through a 5-mm mesh, plant samples were digested in concentrated

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