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Data Article

# Data on the positive synergic action of dimethylacetamide and trehalose on quality of cryopreserved chicken sperm



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#### ARTICLE INFO

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# ABSTRACT

This data article contains supporting information regarding the research article entitled "Combined effect of permeant and nonpermeant cryoprotectants on the quality of frozen/thawed chicken sperm" (Mosca et. al., 2016) [1]. The combined effect of the permeant cryoprotectants agent dimethylacetamide and the non-permeant cryoprotectants agent trehalose on the quality of frozen-thawed chicken semen was assessed. In particular, the quantitative dimethylacetamide/trehalose ratio was investigated freezing semen samples according to the following treatments: trehalose 0.1 M+0% dimethylacetamide (DMA-0), trehalose 0.1 M+3% dimethylacetamide (DMA-3), trehalose 0.1 M+6% dimethylacetamide (DMA-6). © 2016 The Authors. Published by Elsevier Inc. This is an open access

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## Specifications Table

Subject areaBiology, Animal ScienceMore specific<br/>subject areaCryoconservation of chicken semen

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Type of data How data was acquired	Table Fluorescence microscopy, SCA (Sperm Class Analyzer)
Data format	Analyzed
Experimental factors	The natural osmoprotectant trehalose $(0.1 \text{ M})$ was combined with different level $(0-6\%)$ of the permeant cryoprotectant dimethylacetamide to prevent cryoda- mages in chicken semen.
Experimental features	Sperm quality was assessed before and after freezing/thawing in chicken semen processed for cryopreservation using a range of quantitative dimethylacetamide/ trehalose ratios to identify the most effective cryoprotective combination.
Data source location	Milano, Lodi (Italy)
Data accessibility	Data is available with this article

## Value of the data

- Data presented in this paper confirm a positive synergic action of dimethylacetamide and trehalose on quality of frozen-thawed chicken sperm.
- These data encourage the investigation on the interaction between permeating cryoprotectants, like dimethylacetamide, and natural osmoprotectants, such as trehalose, to improve the success of sperm cryopreservation in birds.
- These data contribute for designing further experiments aiming to identify a chicken semen cryopreservation reference procedure.

#### 1. Data

Data include all sperm quality parameters recorded in fresh and cryopreserved chicken semen (Table 1) and the recovery rates of viable and motile sperm after freezing-thawing (Table 2). The most effective cryoprotectant combination includes both trehalose and DMA; in contrast, the absence of DMA (DMA-0) is responsible for more severe loss in sperm quality.

#### Table 1

Sperm quality parameters (LSMeans  $\pm$  SE) measured in fresh semen and in semen frozen according the following treatments: 0.1 M trehalose+0% dimethylacetamide (DMA-0), 0.1 M trehalose+3% dimethylacetamide (DMA-3), 0.1 M trehalose+6% dimethylacetamide (DMA-6).

Sperm parameters <sup>a</sup>	Fresh	DMA-0	DMA-3	DMA-6	S.E.
Viability (%)	87.9 <sup>A</sup>	4.3 <sup>B</sup>	31.8 <sup>C</sup>	37.1 <sup>C</sup>	2.0
Motility (%)	81.7 <sup>A</sup>	8.0 <sup>B</sup>	24.2 <sup>C</sup>	29.1 <sup>C</sup>	2.1
Progressive motility (%)	14.1 <sup>A</sup>	0.1 <sup>B</sup>	1.5 <sup>B</sup>	1.2 <sup>B</sup>	1.3
VCL (µm/s)	47.4 <sup>A</sup>	25.7 <sup>B</sup>	35.6 <sup>C</sup>	33.7 <sup>C</sup>	1.5
VSL (µm/s)	17.0 <sup>A</sup>	4.6 <sup>B</sup>	10.1 <sup>C</sup>	9.3 <sup>C</sup>	0.8
VAP $(\mu m/s)$	28.3 <sup>A</sup>	10.2 <sup>B</sup>	18.4 <sup>C</sup>	17.8 <sup>C</sup>	1.0
LIN (%)	35.7 <sup>A</sup>	17.9 <sup>B</sup>	28.1 <sup>C</sup>	27.7 <sup>C</sup>	1.0
STR (%)	59.8 <sup>A</sup>	45.2 <sup>B</sup>	54.4 <sup>C</sup>	52.5 <sup>C</sup>	1.1
WOB (%)	59.6 <sup>A</sup>	39.3 <sup>B</sup>	51.6 <sup>C</sup>	52.7 <sup>C</sup>	0.9
ALH (µm)	2.8 <sup>A</sup>	0.9 <sup>B</sup>	2.5 <sup>C</sup>	2.7 <sup>A</sup>	0.1
BCF (Hz)	7.9 <sup>A</sup>	0.7 <sup>B</sup>	6.1 <sup>C</sup>	5.4 <sup>C</sup>	0.4

<sup>A,B</sup> Values within each row with different superscript letters are significantly different (p < 0.001).

<sup>a</sup> Viability, the percentage of viable spermatozoa; motility, the percentage of motile spermatozoa; progressive motility, spermatozoa swim forward fast in a straight line; VCL, curvilinear velocity; VSL, straight-line velocity; VAP, average path velocity; LIN (VSL/VCL  $\times$  100), linearity; STR (VSL/VAP  $\times$  100) straightness; WOB (VAP/VCL  $\times$  100); ALH, amplitude of lateral head displacement; BCF, beat cross frequency.

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