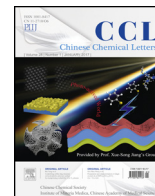




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Original article

Triterpenoids and phenolics from the fruiting bodies of *Inonotus hispidus* and their activations of melanogenesis and tyrosinase

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ABSTRACT

Two new 24-methyl lanostane triterpenoids, hispindic acids A and B (**1** and **2**), and a new phenolic compound, hispinine (**7**), along with nine known compounds (**3–6**, and **8–12**), were isolated from the fruiting bodies of *Inonotus hispidus*. Their structures were elucidated based on the extensive analysis of spectroscopic data (NMR and HRMS). Hispindic acid A (**1**) possesses an unusual formyl group at C-30. Compounds **1**, **3–4**, and **8** showed stronger activate abilities of melanogenesis and tyrosinase in B16 melanoma cells than those of positive control, 8-methoxypsoralen, at 50 $\mu\text{mol/L}$.

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1. Introduction

Inonotus hispidus (Bull. Ex Fr.) Karst is a parasitic fungus in the family Hymenochaetaceae. It is preferably living on a variety of deciduous trees such as malus, fraxinus, sorbus and quercus. *I. hispidus* mainly distributed in the Northeast regions and Xinjiang province of China, and was used as a traditional medicine for the treatment of dyspepsia, cancer, diabetes and stomach problems in these regions [1]. Previous chemical investigations of this species have reported the presence of a considerable quantity of yellow-brown pigments, e.g. hispidin, hispolon, and some hispidin derivatives dimmers, which exhibited antimicrobial, antioxidant, and anti-inflammatory activities [2–7]. In our interest in discovering bioactive compounds from Xinjiang (China) indigenous medicinal fungus, three new compounds including two 24-methyl lanostane triterpenoids (**1** and **2**) and a phenolic compound (**7**), together with nine known compounds (**3–6**, and **8–12**) (Fig. 1), were isolated and identified from the methanolic extract of the fruiting bodies of *I. hispidus*. Moreover, all of the isolates were evaluated for their activations of melanogenesis and tyrosinase, the related targets of vitiligo. Herein, the isolation and structural elucidation, as well as the evaluation of activating melanogenesis and tyrosinase, were present.

2. Results and discussion

The total of 12 compounds (**1–12**) including three new ones, were isolated from the fruiting bodies of *I. hispidus*. Herein, the structural elucidation of the new compounds is presented.

Compound **1** was obtained as white amorphous powder, its molecular formula was determined as $\text{C}_{31}\text{H}_{48}\text{O}_4$ by HRESIMS at m/z 483.3489 $[\text{M}-\text{H}]^-$ (calcd. for $\text{C}_{31}\text{H}_{47}\text{O}_4$, 483.3474). The IR absorptions showed the presence of hydroxyl (3445 cm^{-1}) and carbonyl (1706 cm^{-1}) functionalities. The ^1H NMR spectrum of **1** (Table 1) showed six methyl groups signals at δ_{H} 0.71, 0.73, 0.87, 0.98 (each 3H, s), 0.95 (d, 3H, $J=7.5\text{ Hz}$), 0.96 (d, 3H, $J=6.6\text{ Hz}$), a terminal double bond signals at δ_{H} 4.72, 4.63 (s, each 1H), a formyl group signal at δ_{H} 9.36 (s, 1H) and an oxygen-bearing methine signal at δ_{H} 3.00 (m, 1H). The ^{13}C NMR (Table 1) and HSQC spectra revealed the presence of 31 carbon resonances comprising six methyls, 11 methylenes, five methines and nine quaternary carbons (two of which are carbonyl and three olefinic carbons). The aforementioned data implied compound **1** to be a 24-methyl lanostane triterpenoid. Comparison of the ^1H NMR and ^{13}C NMR data of **1** with those of compound **3** (eburicoic acid) [8], indicated that they were structurally similar, the only difference being the group at C-30. The methyl group of position-30 was replaced by a formyl group in **1**, which was supported by the NMR data of CHO-30 (δ_{H} 9.36, δ_{C} 199.8), and verified by the HMBC correlations (Fig. 2a) from H-30 to C-14 (δ_{C} 67.2) and C-15 (δ_{C} 22.3). The overall structure of compound **1** was further confirmed by $^1\text{H}-^1\text{H}$ COSY

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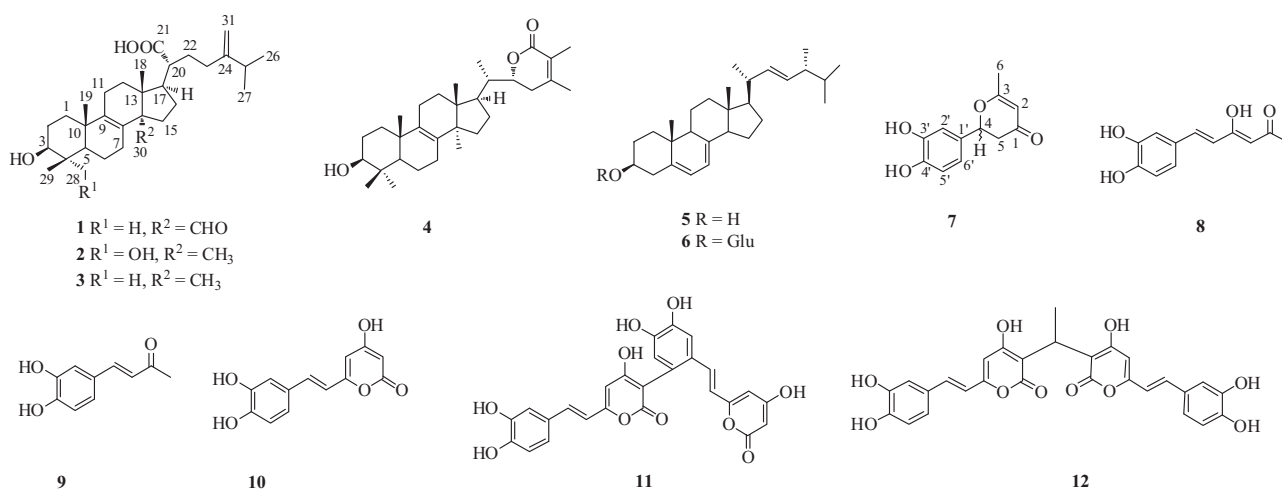


Fig. 1. Structures of compounds 1–12.

and HMBC data. The relative configurations of compound **1** were established by examination of the NOESY experiment (Fig. 2b) and coupling constant. The β -orientation of 3-OH was determined on the basis of coupling constant of H-3 ($J = 10.3, 5.0$ Hz) [9]. NOESY correlations from: H-3 to H-1 α , H-5 and H₃-28; H-30 to H-12 α ; H-12 α /H-17, indicated that they were co-facial, and in an α -orientation. Consequently, the NOESY correlations from: H₃-19 to H₃-29, H-12 β and H₃-18; H₃-18 to H-20, showed they were β -oriented. The above NOESY data indicated that compound **1** possessed the

same relative configurations with eburicoic acid. Therefore, the structure of compound **1** was elucidated as 24-exomethylene-3 β -hydroxy-30-oxo-lanost-8-en-21-oic acid, assigned the trivial name hispidic acid A.

Compound **2** was obtained as a white, amorphous powder with a molecular formula of C₃₁H₅₀O₄, as determined by HR-ESIMS at m/z 485.3661 [M-H]⁻ (calcd. for C₃₁H₄₉O₄, 485.3631). Analysis of ¹H NMR and ¹³C NMR data of compound **2** suggested that the structure of **2** was also similar to that of compound **3** (eburicoic

Table 1
¹H NMR and ¹³C NMR Spectroscopic data of compounds **1**, **2** and **7**.^a

Position	1 ^b		2 ^b		7 ^c	
	δ_H (mult, J, Hz)	δ_C	δ_H (mult, J, Hz)	δ_C	δ_H (mult, J, Hz)	δ_C
1	1.72 (m), 1.22 (m)	36.5	1.61 (m), 1.09 (m)	35.4		194.4
2	1.52 (m, 2H)	28.7	1.51 (m, 2H)	27.6	5.40 (s)	103.7
3	3.00 (dd, 10.3, 5.0)	77.7	3.44 (m)	70.7		176.3
4		39.8		42.4	5.29 (dd, 13.9, 3.6)	80.9
5	0.93 (m)	50.9	1.38 (m)	42.3	2.82 (dd, 16.6, 13.9)	41.3
					2.47 (dd, 16.6, 3.6)	
6	1.59 (m), 1.45 (m)	19.0	1.55 (m), 1.40 (m)	17.8	2.05 (3H, s)	19.7
7 (1')	1.87 (m), 1.46 (m)	27.3	1.92 (m), 1.31 (m)	26.9		129.6
8 (2')		122.7		133.8	6.86 (brs)	113.3
9 (3')		145.1		134.8		145.1
10 (4')		38.7		36.7		145.6
11 (5')	2.27 (m), 2.13 (m)	22.4	2.01 (m), 1.91 (m)	20.8	6.77 (d, 8.5)	114.8
12 (6')	1.90 (m), 1.51 (m)	29.0	1.55 (m), 1.37 (m)	28.9	6.74 (brd, 8.5)	117.9
13		46.2		44.3		
14		67.2		49.5		
15	1.98 (m), 1.35 (m)	22.3	1.55 (m), 1.15 (m)	30.5		
16	1.75 (m), 1.35 (m)	27.2	2.03 (m), 1.93 (m)	26.0		
17	1.38 (m)	49.8	1.93 (m)	47.0		
18	0.73 (s, 3H)	17.7	0.69 (3H, s)	16.1		
19	0.98 (s, 3H)	20.4	0.93 (3H, s)	19.8		
20	2.13 (m)	48.0	2.11 (m)	48.0		
21		177.8		177.4		
22	1.53 (m), 1.42 (m)	31.3	1.60 (m), 1.49 (m)	31.1		
23	1.88 (m, 2H)	32.5	1.91 (2H, m)	32.0		
24		155.8		155.2		
25	2.15 (m)	34.4	2.19 (m)	33.7		
26	0.95 (d, 3H, 7.5)	22.8	0.96 (d, 3H, 6.5)	22.1		
27	0.96 (d, 3H, 6.6)	22.7	0.97 (3H, d, 7.0)	22.0		
28	0.87 (s, 3H)	29.2	3.33 (overlapped)	64.9		
			3.10 (dd, 10.3, 5.1)			
29	0.71 (3H, s)	17.0	0.56 (3H, s)	12.9		
30	9.36 (s)	199.8	0.84 (3H, s)	24.5		
31	4.72 (s), 4.63 (s)	108.0	4.72 (s), 4.64 (s)	107.2		

^a Recorded at 600 or 150 MHz for ¹H and ¹³C, resp.

^b In DMSO-*d*₆.

^c In CD₃OD.

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