



ELSEVIER

Contents lists available at ScienceDirect

Coordination Chemistry Reviews

journal homepage: www.elsevier.com/locate/ccr

Mechanistic studies on versatile metal-assisted hydrogen peroxide activation processes for biomedical and environmental incentives

Maria Oszajca^a, Małgorzata Brindell^a, Łukasz Orzeł^a, Janusz M. Dąbrowski^a,
Klaudyna Śpiewak^a, Przemysław Łabuz^a, Michał Pacia^a, Anna Stochel-Gaudyn^b,
Wojciech Macyk^a, Rudi van Eldik^{a,*}, Grażyna Stochel^{a,*}

^a Faculty of Chemistry, Jagiellonian University, Ingardena 3, 30-060 Kraków, Poland

^b Department of Pediatrics, Gastroenterology and Nutrition, Polish-American Children's Hospital, Jagiellonian University Medical College, Wielicka 265, 30-663 Kraków, Poland

Contents

| | |
|---|----|
| 1. Introduction | 2 |
| 2. Mechanistic studies on model systems – thermal activation | 4 |
| 2.1. Fenton and Fenton-like systems | 4 |
| 2.2. Cytochromes P450 and functional models | 5 |
| 2.2.1. Peroxide shunt and proton-shuttle pathways | 5 |
| 2.2.2. Generation of H ₂ O ₂ by cytochromes P450 | 6 |
| 2.2.3. Activation of H ₂ O ₂ by cytochromes P450 – biomimetic studies | 6 |
| 2.2.4. Activation of H ₂ O ₂ by Fe ^{III} functional models | 7 |
| 2.3. Manganese complexes in bio-related systems | 8 |
| 2.3.1. Activation of H ₂ O ₂ by macrocyclic Mn complexes | 8 |
| 2.3.2. Enzyme-mimicking Mn complexes | 9 |
| 2.3.3. Therapeutic potential of Mn complexes | 10 |
| 2.3.4. Environmental role of Mn complexes | 11 |
| 3. Light and metal compounds in generation, activation and utilization of H ₂ O ₂ | 11 |
| 3.1. Photo-Fenton | 12 |
| 3.2. Macrocyclic photosensitizers and metal ions | 12 |
| 3.2.1. Influence of the central metal ions on the tetrapyrrole properties | 12 |
| 3.2.2. Photodynamic therapy (PDT) | 13 |
| 3.2.3. Mechanisms involving H ₂ O ₂ formation relevant to PDT | 15 |
| 3.2.4. Pro-oxidant and antioxidant effects of ascorbate on PDT | 16 |
| 3.3. Semiconductors as photosensitizers | 17 |
| 3.3.1. Bare TiO ₂ systems | 17 |
| 3.3.2. Photosensitized TiO ₂ systems | 18 |
| 3.3.3. Other photocatalytic systems | 19 |
| 4. Conclusions and perspectives | 19 |
| Acknowledgements | 20 |
| References | 20 |

Abbreviations: ROS, reactive oxygen species; SOD, superoxide dismutase; Por, porphyrin; TAML, tetraamido macrocyclic ligand; DFT, density functional theory; AOPs, advanced oxidation processes; CYPs, cytochromes P450; NAD(P)H, nicotinamide adenine dinucleotide phosphate/nicotinamide adenine dinucleotide; PhIO, iodosylbenzene; TPFPP, *meso*-tetrakis(pentafluorophenyl)porphyrin; TF₄TMAP, *meso*-tetrakis(2,3,5,6-tetrafluoro-N,N,N-trimethyl-4-aniliniumyl)porphyrin; TPPS, *meso*-tetrakis(4-sulphonatophenyl)porphyrin; TOF, turnover frequency; ORII, orange II; NOX, NADPH oxidase; IBD, inflammatory bowel diseases; LMCT, ligand to metal charge transfer; AMD, age-related macular degeneration; PS, photosensitizer; DLI, drug-to-light interval; IL, interleukins; TNF, tumor necrosis factor; IFR, interferon; PET, positron emission tomography; F₂PMet, *meso*-tetrakis(2,6-difluoro-5-N-methylsulfamoylphenyl)porphyrin; WST9 (padaporfin), palladium derivatives of bacteriochlorophyll; PDT, photodynamic therapy; V-PDT, vascular target PDT; C-PDT, cellular target PDT; S^{*}, excited sensitizer; CAT-skl, analog of catalase that enters peroxisomes; CT29, colon carcinoma cell line; A549, human lung cancer cell line; 3AT, 3-amino-1,2,4-triazole; 2ME, 2-methoxyestradiol; BSO, buthionine sulfoximine; CT26, colon carcinoma cell line; AH₂, ascorbic acid; DHS, dehydroascorbate; LED, light-emitting diode; TNSs, titanate nanosheets; GO, graphene oxide; RGO, reduced form of graphene; Pd-BPh, palladium bacteriopheophorbide; Pd-[Bchl], palladium bacteriochlorin; Pd-2Ac-Ph, 2-desvinylnil-2-acetyl-Pd-pheophorbide.

* Corresponding authors. Tel.: +48 12 663 2248 (R. van Eldik); +48 12 663 2243 (G. Stochel); fax: +48 12 634 0515.

E-mail addresses: rudi.vaneldik@fau.de (R. van Eldik), stochel@chemia.uj.edu.pl (G. Stochel).

<http://dx.doi.org/10.1016/j.ccr.2016.05.013>

0010-8545/© 2016 Published by Elsevier B.V.

ARTICLE INFO

Article history:

Received 18 February 2016

Accepted 26 May 2016

Available online

Keywords:

Hydrogen peroxide activation

Metal complexes

Photosensitizers

Semiconductors

Redox reactions

Photocatalysis

ABSTRACT

This article focuses on the versatile ways in which hydrogen peroxide can be activated by metal ions and metal compounds, in terms of possible biomedical and environmental applications and future incentives in these areas. Mechanistic studies performed in our group are discussed in reference to related studies reported in the literature. It covers various thermal activation pathways, including Fenton and Fenton-like reactions, the application of iron, manganese and other metals complexes, as well as metalloenzymes. Furthermore, light-induced generation, activation and utilization of hydrogen peroxide are presented for the photo-Fenton reactions, the application of macrocyclic photosensitizers and the use of semiconductors such as TiO₂ as heterogeneous photosensitizers. Mechanistic aspects of several thermal and photochemical pathways in metal-assisted hydrogen peroxide applications for biomedical or environmental purposes are discussed, and some obstacles to be overcome are highlighted.

© 2016 Published by Elsevier B.V.

1. Introduction

The activation of small redox-active inorganic molecules has a significant impact in biology, medicine, environmental protection and industrial processes [1–5]. Metal ions and their compounds play a very important role not only in thermal, but also in photochemical processes of small molecule activation. They can mediate the actual active form of small molecules, as well as control their spatial concentration dynamics. In homogenous systems, metal species can operate as a coordination and/or energy, electron, hydrogen atom transfer center. In heterogeneous systems, small molecules can undergo adsorption followed by electron or energy transfer processes involving solid surface participation. Coordination compounds or solid surface participating in the activation process point to an appropriate reaction site where the geometry plays a key role. The metal ion and its ligands control the redox properties and therefore the nature of coordinated or chemisorbed small molecules.

This review focuses on the versatility of transition metal-assisted activation processes of hydrogen peroxide (H₂O₂), a neutral reactive oxygen species (ROS) with significant biomedical and environmental impact.

Even though H₂O₂ is a strong two-electron oxidant (standard reduction potential $E^{\text{pH}=7}$ is 1.32 V at pH 7.0) (Fig. 1), it exhibits poor or no reactivity at all toward most pollutants or biological molecules. This behavior is a consequence of a high activation energy barrier indicating that the oxidation reaction is a kinetically-controlled process. One-electron reduction makes H₂O₂ a rather weak oxidant ($E^{\text{pH}=7}$ is 0.38 V); however, the resulting product, the hydroxyl radical (HO•), is one of the strongest oxidants known ($E^{\text{pH}=7}$ is 2.31 V). The reactions of HO• have a very low activation energy barrier and proceed very fast at rates close to diffusion-controlled. Therefore, the activation of H₂O₂ by facilitating one-electron reduction plays a crucial role in the application of this molecule as an efficient oxidant. One of the most explored ways to enable H₂O₂ utilization is its transition metal-assisted activation.

Hydrogen peroxide is an important biological reactive oxygen species; it is generated as a by-product of mitochondrial electron transport of aerobic respiration, or as an end product of various metabolic reactions, and is suggested to be the transmitter of cellular signaling [6]. Overproduction of H₂O₂ is well known to lead to oxidative damage of macromolecules and to be the cause of various diseases and aging, whereby the role of H₂O₂ as a messenger molecule in redox signaling is emerging. Although H₂O₂ is less reactive

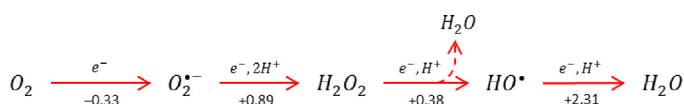


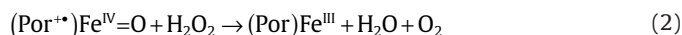
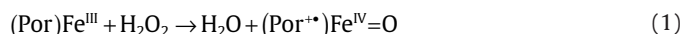
Fig. 1. ROS standard reduction potentials adapted from Reference 6.

than superoxide, O₂^{•-}, control of H₂O₂ levels is critical, given that H₂O₂ can readily diffuse across cellular membranes, and thus have an extended sphere of influence. Active transport is also possible through membranes of the aquaporin family [7]. Since one-electron reduction of O₂ is thermodynamically unfavorable in contrast to one-electron reduction of O₂^{•-}, disproportionation of O₂^{•-} leading to O₂ and H₂O₂ proceeds spontaneously or is significantly accelerated by superoxide dismutases (SODs). In mitochondria, SOD is a manganese (Mn(III)/Mn(II)) enzyme and in cytoplasm a Cu(II)/Cu(I) Zn(II) enzyme.

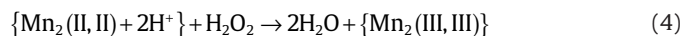
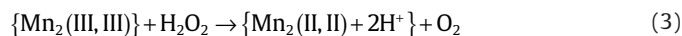
On the assumption that all diseases have the perturbation of the cellular redox environment in common, the development of SOD mimics seems to be an appropriate strategy for the design of potent redox active therapeutics.

There are three major groups of enzymes, viz. catalases, glutathione peroxidases and peroxiredoxins, that are involved in cell defense systems against H₂O₂. Among them, only catalase possesses the metal cofactor, either non-heme manganese or heme (Fig. 2A and B).

Heme catalases possess four heme groups with phenolate from tyrosine as an axial ligand (Fig. 2B) that enable the enzyme to react with hydrogen peroxide. There is a continuing debate in the literature on the mechanism of catalase activity [8]; among others, it is postulated that the oxoferryl porphyrin π-cation radical ((Por^{•+})Fe^{IV}=O, compound I), formed by heterolytic O–O bond cleavage, is a major reactive intermediate as outlined in the following reactions:



In some revised mechanisms, the formation of the oxoferryl porphyrin ((Por)Fe^{IV}=O or (Por)Fe^{IV}OH, compound II) is also proposed to be involved in the catalytic cycle of catalase. Manganese catalases contain a binuclear manganese complex (Fig. 2A) as their catalytic active site and perform a two-electron catalytic cycle in two distinguished steps [9,10] as outlined in the following reactions:



The H₂O₂ molecule coordinates to Mn1 in the {Mn₂(III,III)} cluster by displacing the terminally bound solvent molecule [9,10]. Proton-coupled, two-electron oxidation of the terminally bound hydroperoxide results in formation of the enzyme in the reduced {Mn₂(II,II)} state, with two protons bound to solvent bridges and dioxygen. The release of dioxygen is accompanied by the coordination of solvent in the axial position of Mn1. Subsequently, a second

Download English Version:

<https://daneshyari.com/en/article/5150870>

Download Persian Version:

<https://daneshyari.com/article/5150870>

[Daneshyari.com](https://daneshyari.com)