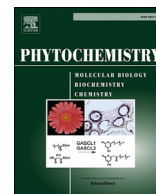




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journal homepage: www.elsevier.com/locate/phytochemCytotoxic triterpenoid saponins from *Lysimachia foenum-graecum*Lu-Mei Dai^{a,1}, Ri-Zhen Huang^{a,b,1}, Bin Zhang^a, Jing Hua^a, Heng-Shan Wang^{a,**}, Dong Liang^{a,*}^a State Key Laboratory for Chemistry and Molecular Engineering of Medicinal Resources, School of Chemistry and Pharmaceutical Sciences, Guangxi Normal University, Guilin 541004, People's Republic of China^b Department of Pharmaceutical Engineering, School of Chemistry and Chemical Engineering, Southeast University, Nanjing 211189, People's Republic of China

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ABSTRACT

Eleven oleanane-type triterpenoid saponins, foegraecumosides A–K, and eight known ones, were isolated from the aerial parts of *Lysimachia foenum-graecum*. Their structures were elucidated by spectroscopic data analyses and chemical methods. All isolated saponins were evaluated for their cytotoxicity against four human cancer cell lines (NCI-H460, MGC-803, HepG2, and T24). Seven saponins containing the aglycone cyclamiretin A exhibited moderate cytotoxicity against all tested human cancer cell lines, with IC₅₀ values of 9.3–24.5 μM. Simultaneously, the cytotoxic activities of foegraecumosides A and B, lysichriside A, ardisiacrispines A and B, cyclaminorin, and 3-O-α-L-rhamnopyranosyl-(1 → 2)-β-D-glucopyranosyl-(1 → 4)-α-L-arabinopyranosyl-cyclamiretin A were tested on drug-resistant lung cancer cell lines (A549 and A549/CDDP, respectively). Ardisiacrispin B displayed moderate cytotoxicity against A549/CDDP, with an IC₅₀ value of 8.7 μM and a resistant factor (RF) of 0.9.

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1. Introduction

The genus *Lysimachia* of the family Primulaceae, comprising about 180 species, is widely distributed throughout the world. However, data from recent phylogenetic analyses suggested its relocation to the family Myrsinaceae (Podolak et al., 2013). *L. foenum-graecum* is one of the best-known plants from this genus, distributed mainly in the Guangxi and Yunnan Provinces of China. The aerial parts of this plant have been used not only as a perfumery plant and an insect repellent, but also for the treatment of colds and headaches in traditional Chinese medicine (The Health Administration of Beijing, 1998). Previous phytochemical investigation of this species led to isolation of several triterpenoid

saponins and flavonoids (Shen et al., 2005; Li et al., 2007, 2009a,b; 2010). In a continuing search for bioactive constituents from medicinal plants in the Guangxi Zhuang Autonomous Region, the aerial parts of *L. foenum-graecum* were investigated and eleven new oleanane-type triterpenoid saponins, named foegraecumosides A–K (1–11) (Fig. 1) were isolated, together with eight known ones (12–19) (Fig. 1). The isolated saponins were evaluated for their cytotoxic activities against a panel of drug-sensitive and drug-resistant human cancer cell lines. Herein, described are the isolation, structural elucidation, and biological assays of these compounds.

2. Results and discussion

The aerial parts of *L. foenum-graecum* were collected from the Jinxiu County of Guangxi Province and extracted with 95% aqueous EtOH. The EtOH extract was suspended in H₂O and partitioned successively with EtOAc and *n*-BuOH. The *n*-BuOH-soluble extract was subjected to macroporous resin column chromatography to give a crude saponin fraction, which showed cytotoxicity against three human cancer cell lines (HepG2, MGC-803, and T24) with IC₅₀ values of 2.6–7.1 μg/mL. The crude saponin fraction was subjected to further column chromatography and purified by preparative HPLC, to afford eleven new (1–11) and eight known (12–19)

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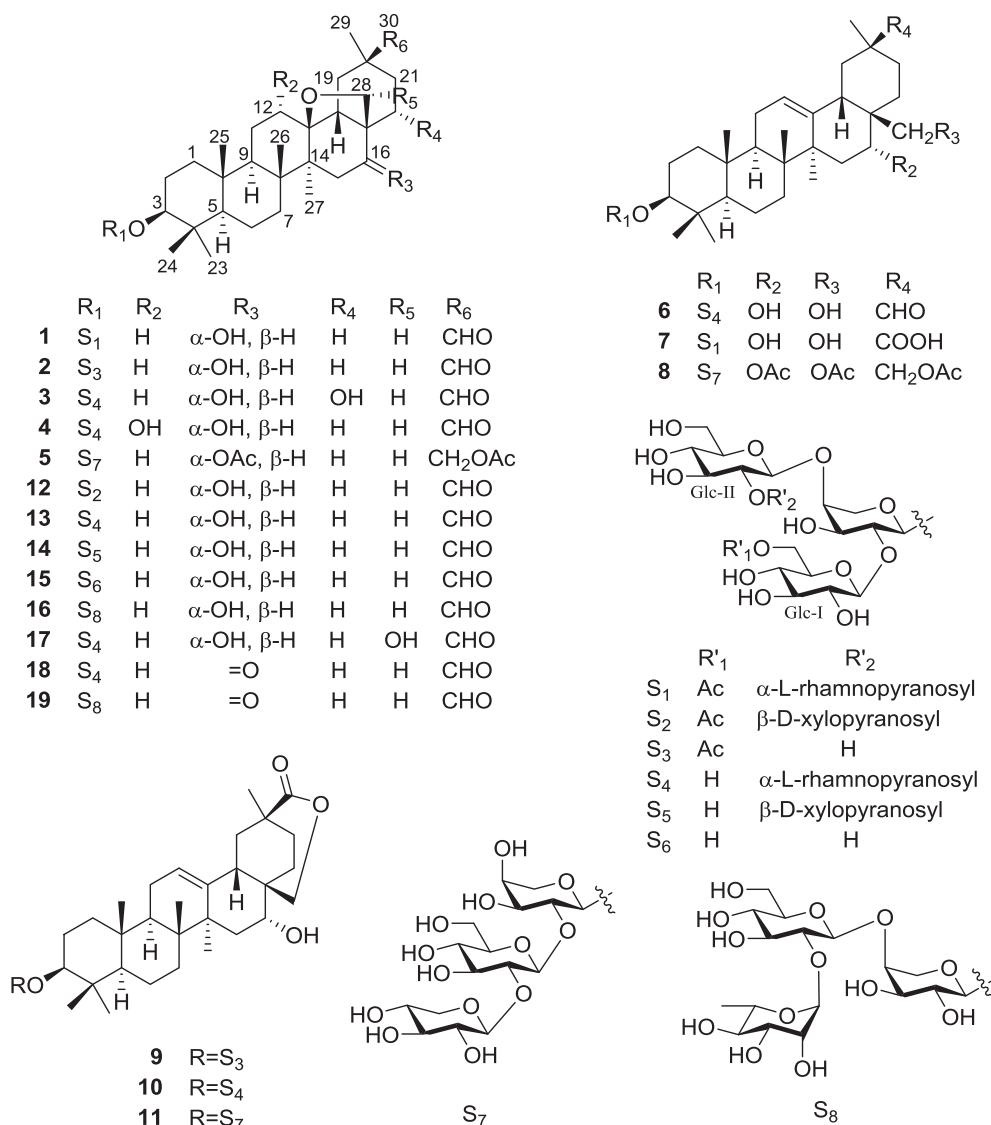


Fig. 1. Structures of compounds 1–19.

oleanane-type triterpenoid saponins. All of the new compounds (1–11) were obtained as amorphous powders. Acid hydrolysis of each compound (1–11) and subsequent HPLC analysis of the sugars using an optical detector (Yoshikawa et al., 2003, 2007) allowed the characterisation of D-glucose, L-arabinose, and L-rhamnose for compounds 1, 3, 4, 6, 7, and 10, D-glucose and L-arabinose for compounds 2 and 9, and D-glucose, L-arabinose, and D-xylose for compounds 5, 8, and 11, respectively.

Foegraecumoside A (1) possessed the molecular formula of C₅₅H₈₈O₂₃ on the basis of its positive-ion HRESIMS (*m/z* 1139.5608 [M + Na]⁺, calcd 1139.5609) and ¹³C NMR data (Table 2). Its ¹H NMR spectrum (Table 1) showed six methyl singlets at δ_H 0.83, 0.99, 1.02, 1.21, 1.27, and 1.51, one oxygenated methylene at δ_H 3.14 and 3.52 (each 1H, d, *J* = 7.5 Hz), and one aldehyde proton at δ 9.60 (1H, s), which were unambiguously designated by HSQC experiment. Its ¹³C NMR (Table 2) and DEPT spectra displayed six *sp*³ carbon resonances at δ_C 16.38, 16.42, 18.5, 19.7, 24.1, and 28.0, an oxygenated methylene at δ_C 77.6, a quaternary carbon resonance at δ_C 86.4, and an aldehyde carbon at δ_C 207.5. The NOESY correlation of H-30 (δ_H 9.60)/H-18 (δ_H 1.35) confirmed the β-orientation of the aldehyde group. These data indicated that 1 was based on a 13,28-epoxy-

oleanane skeleton. On the basis of its 2D NMR (¹H–¹H COSY, HSQC, and HMBC) analyses and comparison with literature data, the aglycone of 1 was identified as 3β,16α-dihydroxy-13β,28-epoxy-oleanan-30-al (cyclamiretin A) (Dong et al., 2011).

The ¹H NMR spectrum of 1 exhibited signals of four sugar anomeric protons at δ_H 4.87 (1H, d, *J* = 5.0 Hz), 5.12 (1H, d, *J* = 8.0 Hz), 5.38 (1H, d, *J* = 7.5 Hz), and 6.35 (1H, br s), which correlated to four anomeric carbons at δ_C 104.4, 103.5, 105.0, and 101.6, respectively, according to the HSQC spectrum. The extensive 2D NMR spectroscopic analyses together with results of acid hydrolysis of 1 allowed the characterisation of two β-D-glucopyranosyls (Glc), one α-L-rhamnopyranosyl (Rha), and one α-L-arabinopyranosyl (Ara), respectively. In the same way, these sugars were identified in compounds 3, 4, 6, 7, and 10. The coupling constants also confirmed the β-glycosidic linkages for two glucopyranosyl units. The arabinopyranosyl unit was determined to be the α-anomer on the basis of: the ³_J_{H1,H2} value (5.0 Hz) and the correlations of Ara-H-1/Ara-H-3 and Ara-H-1/Ara-H-5 in the NOESY spectrum (Chang et al., 2007; Liang et al., 2011), as well as the ¹H non-splitting pattern (br s) and the large *J*_{C1-H1} coupling constants (171 Hz) which confirmed the α-anomeric orientation of the

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