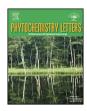
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Mid-infrared spectroscopy and short wave infrared hyperspectral imaging—A novel approach in the qualitative assessment of *Harpagophytum procumbens* and *H. zeyheri* (Devil's Claw)

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ABSTRACT

Harpagophytum procumbens (Burch.) DC. ex Meisn. subsp. *procumbens* (Pedaliaceae) is an important African medicinal plant growing in the Kalahari region of southern Africa. This species, together with its close taxonomic ally *Harpagophytum zeyheri* are collectively referred to as Devil's Claw and are used interchangeably for the treatment of inflammation-related disorders. Although the two taxa are botanically and chemically similar, *H. zeyheri* contains lower levels of harpagoside and these two species have not been proven to exhibit equipotent pharmacological activity. Due to these taxonomic similarities, effective quality control methods are required to distinguish between the two species. Differentiation between the two species was achieved using single point mid-infrared spectroscopy in combination with chemometric data analysis. The orthogonal projections to latent structures discriminant analysis (OPLS-DA) model had good predictive ability, as illustrated by the model statistics: R^2X (cum predictive + orthogonal) = 0.86 and Q^2 (cum) = 0.63. Short wave infrared (SWIR) hyperspectral imaging coupled with chemometric modelling is a reliable and rapid method to determine the authenticity of *Harpagophytum* spp.

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1. Introduction

Harpagophytum procumbens (Burch.) DC. ex Meisn. subsp. procumbens (Pedaliaceae) is an important African medicinal plant found in the Kalahari region of southern Africa which includes Namibia, South Africa, Botswana, Angola, Zimbabwe, Zambia and Mozambique. According to ethnobotanical information, secondary root tubers are used to treat painful inflammation of the joints and muscles in rheumatism and arthritis, back pain, and as a tonic for gastro-enterological disturbances (Mncwangi et al., 2012; van Wyk and Gericke, 2000). A topical ointment prepared with animal fat or petroleum is traditionally applied to treat sores, ulcers and boils (van Wyk and Gericke, 2000). Other traditional uses of *H. procumbens* include the treatment of fever, diabetes, diarrhoea

Abbreviations: ATR, attenuated total reflectance; MIR, mid-infrared; OPLS-DA, orthogonal projections to latent structures discriminant analysis; PCA, principal component analysis; SWIR, short wave infrared; SNV, standard normal variate.

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and blood disease (Stewart and Cole, 2005). Studies have24confirmed that extracts of the secondary tubers of *H. procumbens*25are effective in the treatment of degenerative rheumatoid arthritis,26osteoarthritis, tendonitis, kidney inflammation, heart disease,27dyspepsia and loss of appetite (Stewart and Cole, 2005; Viljoen28et al., 2012).29

H. procumbens and the closely related Harpagophytum zeyheri 30 are collectively known as Devil's Claw and are used interchange-31 ably (Kemper, 1999). The British and European Pharmacopoeias 32 make reference to both H. procumbens and H. zevheri but H. 33 procumbens is the species of choice as it has been shown to contain 34 higher amounts of the biologically active ingredient, harpagoside 35 (Mncwangi et al., 2012). H. zeyheri has been included in 36 formulations as an adulterant where H. procumbens is used as 37 starting material (McGregor et al., 2005), largely due to the 38 declining natural populations of H. procumbens. H. procumbens is 39 extensively commercialised which creates a conducive environ-40 ment for adulteration which may have negative consequences for 41 both the trader and the consumer (Stewart and Cole, 2005). 42

Phytochemical studies have confirmed the presence of iridoid 43 glycosides such as harpagoside, procumbide and harpagide, 44 phenylethanoid glycosides such as acteoside and isoacteoside 45

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46 and other substances including harpagoquinones, amino acids, flavonoids, phytosterols and carbohydrates in Devil's Claw 47 48 (Gruenwald, 2002; Kikuchi et al., 1983; Kurkin, 2003). However, 49 only harpagoside is used as a biomarker to determine the quality of 50 raw material and products. Various chromatographic and spectro-51 scopic methods have been developed for the quality assurance of 52 H. procumbens raw material and products. Using NIR-FT-Raman 53 spectroscopy, Baranska et al. (2005) identified characteristic key 54 bands in *H. procumbens* raw materials. ethanol extracts and tablets. 55 Diagnostic key bands were observed in the frequency range of 1600–1700 cm⁻¹, which is similar to the wavenumber region for 56 57 harpagoside, and is assigned to -C=O, -C=C and benzene ring

stretching vibrations. Schulz and Baranska also developed a calibration model for harpagoside using HPLC as a reference method. Raman mapping was used to visualise the spatial distribution of harpagoside in different samples (Schulz and Baranska, 2007).

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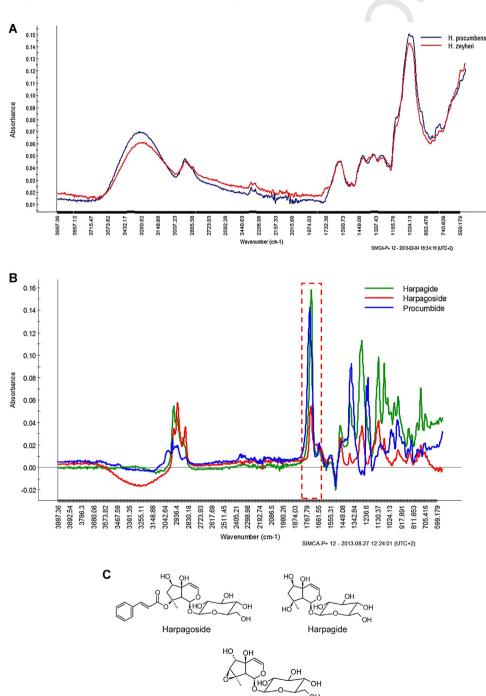
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Quality control (QC) is a major concern in the herbal and phytomedicine industry; vibrational spectroscopy is one of the modern methods used in raw material identification and inprocess monitoring (Reich, 2005). Vibrational spectroscopy offers various advantages over current analytical techniques; these include its robustness, efficiency, non-destructiveness and costeffectiveness. In addition, minimal or no sample preparation is



Procumbide

Q3 Fig. 1. (A) MIR spectra of powdered *H. procumbens* (blue) and *H. zeyheri* (red) secondary root tubers. (B) MIR spectra of three iridoid glycosides (harpagoside, harpagide and procumbide) isolated from *H. procumbens*. (C) Chemical structures of harpagoside, harpagide and procumbide. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of the article.)

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