Tetrahedron 72 (2016) 7481-7487

Contents lists available at ScienceDirect

Tetrahedron

journal homepage: www.elsevier.com/locate/tet

Limonoids with modified furan rings from root barks of *Toona* sinensis

ABSTRACT

Jun-He Li^a, Yi Li^b, Fa-Liang An^a, Miao-Miao Zhou^a, Jie Luo^a, Kai-Li Jian^a, Jun Luo^{a,*}, Ling-Yi Kong^{a,*}

^a State Key Laboratory of Natural Medicines, Department of Natural Medicinal Chemistry, China Pharmaceutical University, 24 Tong Jia Xiang, Nanjing 210009, People's Republic of China
^b Testing & Analysis Center, Nanjing Normal University, Nanjing 210046, People's Republic of China

macrophages at nontoxic concentration.

ARTICLE INFO

Article history: Received 17 August 2016 Received in revised form 23 September 2016 Accepted 27 September 2016 Available online 28 September 2016

Keywords: Toona sinensis (A. Juss.) Roem. Limonoids Modified furan rings Cytotoxicity anti-Inflammatory activity

1. Introduction

Toona sinensis (A. Juss.) Roem. (Meliaceae), a perennial deciduous arbour, is widely distributed and cultivated in eastern and southeastern Asia, and it has more than 2000 years of cultivation history in China. The leaves of T. sinensis, which contain a distinct flavour, have been the vegetarian cuisine and also used as a traditional Chinese herbal medicine for treatment of itch, dysentery, and enteritis, while the bark was used as astringent and depurative substance.^{1–3} In the previous investigations of *T. sinensis*, a variety of ring A-seco limonoids and ring B-seco limonoids were reported as its main constituents.^{4–6} During an ongoing search for limonoids with novel structure and significant biological activities, the root barks of T. sinensis were chosen as our research subject. As a result, 12 new azadirone- and gedunin-type limonoids with diverse modified furan rings, Toonasinemines A-K (1-12), were isolated from the root barks of T. sinensis. Their structures were elucidated on the basis of spectroscopic methods including high resolutionelectrospray ionization (HR-ESI)-MS, one and two dimensional (1D and 2D)-NMR. The N-containing modification in furan rings of compounds 1–7, known as limonoid-based alkaloids, were rare in limonoids family.^{7–14} Due to the structure of the rare modified furan rings, all new limonoids were evaluated in vitro for their potential biological activities, such as cytotoxicity and antiinflammatory activity. Herein, we describe the isolation and

structural elucidation of these new limonoids as well as their

2. Results and discussion

bioactivities.

Toonasinemines A-K (1–12). 12 new azadirone- and gedunin-type limonoids with eight-classes of

modified furan rings, were isolated from the root barks of Toona sinensis (A. Juss.) Roem. Their structures

were elucidated based on detailed spectroscopic analyses, and the absolute configurations of **1–9** were

established by the electronic circular dichroism exciton chirality method. The lactam-bearing moieties in

compounds 1–7 were rare in limonoids family. Results of bioactivities screening exhibited that 1, 4, and 8 showed moderate cytotoxicities against HepG2, MCF-7, and U2OS human cancer cell lines, and 1, 2, 6, 8,

and 9 exhibited marked inhibitory effects on nitric oxide (NO) production in LPS-activated RAW 264.7

Toonasinemine A (1) was obtained as a white, amorphous powder. Its molecular formula was determined to be C₂₈H₃₅NO₆ from the $[M+H]^+$ peak at m/z 482.2536 (calcd 482.2537). The ¹H NMR data of 1 (Table 1) displayed resonances characteristic of five tertiary methyl groups ($\delta_{\rm H}$ 1.06, 1.07, 1.07, 1.20 and 1.22, each 3H, s) and one acetate methyl group ($\delta_{\rm H}$ 2.02, 3H, s). The ¹³C NMR (Table 1) spectrum indicated the presence of six methyls, four methylenes, eight methines, and 10 quaternary carbons with the help of HSQC data. ¹H and ¹³C NMR data combined with 2D NMR spectra suggested 1 as being a azadirone-type limonoid with intact A-D rings. The ¹³C NMR signals of C-1 (δ_{C} 157.7), C-2 (δ_{C} 125.9), and C-3 ($\delta_{\rm C}$ 204.3) and the ¹H NMR signals of a pair of AB doublets at $\delta_{\rm H}$ 5.87 and 7.16 (*J*=10.5 Hz) suggested that the A-ring of **1** possessed a 1-en-3-one system. The signals of C-14 (δ_C 72.4), C-15 (δ_C 57.4), and H-15 ($\delta_{\rm H}$ 3.92) indicated the presence of 14,15-epoxide. In HMBC spectrum, the correlations (Fig. 2) from H-15 ($\delta_{\rm H}$ 3.39, s)/H-



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^{*} Corresponding authors. Fax: +86 25 8327 1405; e-mail addresses: luojun1981ly@ 163.com (J. Luo), cpu_lykong@126.com (L-Y. Kong).

Table 1			
¹ H NMR (500 MHz) a	and ¹³ C NMR (125 MHz)) spectroscopic data of com	pounds 1–4 in CDCl ₃

Position	1		2		3	3		4	
	$\delta_{\rm H}$ (multi, J in Hz)	δ_{C}	$\delta_{\rm H}$ (multi, J in Hz)	δ_{C}	$\delta_{\rm H}$ (multi, J in Hz)	δ_{C}	$\delta_{\rm H}$ (multi, J in Hz)	δ_{C}	
1	7.16, d (10.2)	157.7	7.16, d (10.2)	158.4	7.11, d (10.2)	157.2	7.09, d (10.2)	156.7	
2	5.87, d (10.2)	125.9	5.84, d (10.2)	125.6	5.86, d (10.2)	126.1	5.88, d (10.2)	126.3	
3		204.3		204.7		204.1		203.9	
4		44.4		44.3		44.2		44.2	
5	2.17, dd (12.7, 3.0)	47.0	2.19, m	46.4	2.16, d (13.5)	46.3	2.17, d (13.3)	46.3	
6α	1.88, m	24.4	1.96, m	24.0	1.94, m	23.4	1.96, m	23.4	
6β	1.88, m		1.79, m		1.81, m		1.80, m		
7	4.71, br s	74.0	5.26 (t-like, 2.6)	74.8	4.53, br s	73.5	4.54, br s	73.4	
8		43.5		43.1		42.8		42.9	
9	2.59, dd (12.5, 4.1)	40.1	2.20, m	38.7	2.45, dd (12.8, 6.3)	39.6	2.54, dd (12.8, 5.8)	39.6	
10		39.9		40.2		40.3		40.3	
11α	1.96, m	16.3	1.92, m	16.7	1.98, m	15.1	2.01, m	15.1	
11β	1.85, m		1.74, m		1.89, m		1.90, m		
12α	1.80, m	29.4	1.81, m	33.1	1.50, m	25.6	1.41, m	26.0	
12β	2.41, m		2.02, m		2.10, m		2.02, m		
13		42.9		47.4		39.7		39.9	
14		72.4		159.0		69.9		69.7	
15	3.39, s	57.4	5.35, d (1.9)	118.9	3.49, s	57.0	3.54, s	56.9	
16α		208.2	2.50, m	34.2		167.7		166.5	
16β			2.25, m						
17	3.93, s	50.3	2.89, m	51.0	5.63, s	76.7	5.67, s	75.5	
18	1.06, s	25.1	0.85, s	21.2	1.23, s	17.4	1.26, s	18.5	
19	1.22, s	20.0	1.19, s	19.2	1.22, s	19.9	1.23, s	19.9	
20		132.1		139.6		135.6		145.2	
21		174.3		174.8		172.5		169.3	
22	7.20, br s	144.5	6.86, s	140.0	7.22, br s	144.6	6.72, s	133.3	
23α	4.07, d (20.1)	47.3	3.99, s	46.5	4.06, s	47.1		168.4	
23β	4.02, d (20.1)		3.99, s		4.06, s				
28	1.07, s	27.2	1.08, s	27.2	1.06, s	27.3	1.06, s	27.3	
29	1.07, s	21.4	1.08, s	21.3	1.07, s	21.4	1.07, s	21.3	
30	1.22, s	19.7	1.22, s	27.6	1.16, s	18.5	1.17, s	17.7	
7-OAc	2.02, s	169.8	1.96, s	170.2	2.10, s	169.9	2.10, s	169.9	
		21.2		21.5		21.2		21.2	
N-H	6.31, br s		6.18, br s		6.35, br s		7.50, br s		

17 ($\delta_{\rm C}$ 3.93, s) to C-16 ($\delta_{\rm C}$ 208.2) suggested a carbonyl group at C-16. The cross-peak between $\delta_{\rm C}$ 169.8 and $\delta_{\rm H}$ 4.71 (H-7) demonstrated that the acetoxy group was attached to C-7. Except for the aforementioned data of rings A–D, the molecular formula C₄H₄NO was left. It suggested the presence of a lactam located at C-17, which was confirmed by the HMBC correlations from H-17 ($\delta_{\rm H}$ 3.93) to C-20 ($\delta_{\rm C}$ 132.1), C-21 ($\delta_{\rm C}$ 174.3), C-22 ($\delta_{\rm C}$ 144.5). The NMR resonances associated with the C-17 substituent [C-20 ($\delta_{\rm C}$ 132.1), C-21 ($\delta_{\rm C}$ 174.3); H-22 ($\delta_{\rm H}$ 7.20, br s), H-23 α ($\delta_{\rm H}$ 4.07, d, *J*=20.0 Hz), H-23 β ($\delta_{\rm H}$ 4.02, d, *J*=20.0 Hz)] were similar to that of the 3-substituted 1,5-dihydro-2*H*-pyrrol-2-one unit in amooramide A.¹⁴ Moreover, the ¹H NMR spectrum of **1** exhibited a resonance characteristic of an amino proton at $\delta_{\rm H}$ 6.31 (br s), which was correlated with C-20, C-21, C-22, and C-23 in the HMBC spectrum. Thus, the planar structure of **1** was determined as depicted in Fig. 1.

The relative configuration of **1** was determined by a ROESY experiment (Fig. 2). The ROESY correlations between H-9/H₃-18, H-5/H-9, H-5/H₃-28 and H-15/H₃-18 showed that H-5, H-9, H-15, Me-18, and Me-28 were on the same face and arbitrarily assigned as α -oriented, whereas the 14,15-epoxide group was β -oriented. The ROESY interactions between H₃-19/H₃-29, H β -12/H₃-19, H β -12/H-17, H β -12/H₃-30, and H-7/H₃-30 suggested the β -orientation of H-7, H-17, Me-19, Me-29, and Me-30. Thus, the structure of **1** was unambiguously determined as depicted.

Toonasinemine B (**2**), an amorphous powder, gave the molecular formula $C_{28}H_{37}NO_4$ on the basis of its $[M+Na]^+$ HRESIMS ion peak at m/z 474.2619 (calcd for $C_{28}H_{37}NNaO_4$, 474.2615). The ¹H NMR and ¹C NMR data (Table 1) of **2** were similar to those of **1**. The



Fig. 1. Chemical structures of compounds 1-12.

differences noted between these two limonoids were that **2** had no signals due to 14,15-epoxide and carbonyl group at C-16, but a 14-double bond was indicated by resonances at 159.0 and 118.9 for C-14 and C-15, respectively, and a resonance ascribed to H-15 at 5.64.

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