

Cytotoxic tiglane-type diterpenoids from *Croton tiglim*

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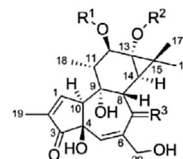
ABSTRACT

Chemical study on the twigs and leaves of *Croton tiglim* has led to the isolation of 11 new tiglane-type diterpenoids, crotnignoids A–K (**1**–**11**), along with eight known analogues (**12**–**19**). The structures of these compounds were elucidated on the basis of detailed spectroscopic analysis. In vitro cytotoxic assays revealed moderate inhibition of compounds **1**–**10** toward human HL-60 and A549 tumor cells, with **1** exhibiting the strongest activity against HL-60 cell line with an IC₅₀ value of 1.61 μM.

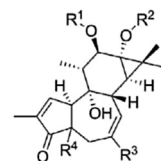
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1. Introduction

The genus *Croton* (Euphorbiaceae) comprises more than 1300 species mainly distributed in the tropical and subtropical regions of the world.¹ Many species of this genus are used as folk medicines for the treatment of stomachache, abscesses, malaria, inflammation, and so on.^{2–4} The *Croton* plants are rich sources of terpenoids displaying a broad spectrum of biological activities, such as acetylcholinesterase inhibitory,² neurite outgrowth-promoting,⁵ antimycobacterial,⁶ antifungal,⁷ antiplasmodial,⁸ cytotoxic,^{9,10} antiviral,¹¹ and antiinflammatory activities.^{12,13} *Croton tiglim* L. is a shrub or small tree that grows in southeast of China such as Zhejiang and Fujian Provinces.¹⁴ Phorbol esters, the typical metabolites of *C. tiglim*, were found to possess tumor-promoting function by Van Duuren and co-workers in 1963.¹⁵ Fifteen years later, Castagna and co-workers discovered the functional mechanism that the protein kinase C (PKC) is the receptor of phorbol esters.¹⁶ In continuing our efforts to identify structurally diverse and biologically important compounds from medicinal plants, 11 new diterpenoids crotnignoids A–K (**1**–**11**) and eight known analogues were isolated from *C. tiglim* L. Subsequent in vitro cytotoxic evaluation revealed that most new compounds displayed cytotoxic activities against two human tumor cell lines A549 and HL-60. Of these compounds, crotnignoid A (**1**) showed the best activity against HL-60 cell line with an IC₅₀ value of 1.61 μM. We herein present the isolation, structural elucidation, and biological tests of these diterpenoids.



	R ¹	R ²	R ³
1	tiglyl	2-methylbutyryl	H,β-OOH
2	tiglyl	isobutyryl	H,β-OOH
3	tiglyl	2-methylbutyryl	H,β-OH
4	tiglyl	isobutyryl	H,β-OH
5	tiglyl	isobutyryl	O
6	tiglyl	propionyl	O
7	H	decanoyl	O



	R ¹	R ²	R ³	R ⁴
8	tiglyl	isobutyryl	CHO	β-OH
9	2-methylbutyryl	2-methylbutyryl	CH ₂ OH	β-OH
10	benzoyl	isobutyryl	CH ₂ OH	β-OH
11	benzoyl	acetyl	CH ₂ OH	α-H
12	tiglyl	2-methylbutyryl	CH ₂ OH	β-OH
13	isobutyryl	acetyl	CH ₂ OH	β-OH
14	tiglyl	propionyl	CH ₂ OH	β-OH
15	tiglyl	isobutyryl	CH ₂ OH	β-OH
16	2-methylbutyryl	acetyl	CH ₂ OH	β-OH
17	tiglyl	acetyl	CH ₂ OH	β-OH
18	2-methylbutyryl	isobutyryl	CH ₂ OH	β-OH
19	tiglyl	acetyl	CH ₂ OH	α-H

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2. Results and discussion

Compound **1** was assigned a molecular formula of $C_{30}H_{42}O_{10}$ based on ^{13}C NMR data and the (+)-HRESIMS ion peak at m/z 585.2660 $[M+Na]^+$ (calcd 585.2670), requiring 10 indices of hydrogen deficiency. The IR spectrum showed absorption bands for hydroxyl (3389 cm^{-1}), carbonyl (1713 cm^{-1}) and vinyl (1651 cm^{-1}) groups. Analysis of the NMR data (Tables 1 and 2) revealed characteristic signals for a 2-methylbutyryl, a tiglyl, and an α,β -unsaturated carbonyl groups, as well as a trisubstituted double bond, six methines (two oxygen bonded), an oxygenated methylene, and four methyls. These observations accounted for six out of 10 indices of hydrogen deficiency supportive of a tetracyclic carbon skeleton for **1**. Further interpretation of 2D NMR especially HMBC data (Fig. 1A) established that compound **1** was a diterpenoid of the tiglane family as briefly characterized below. The HMBC correlations of H-19/C-1, C-2 and C-3; H-5/C-3, C-4 and C-10; H-10/C-1; and H-1/C-4 confirmed the presence of the methylsubstituted five-membered ring-A incorporating an α,β -conjugated ketone moiety. Similarly, the cyclopropyl ring-D with gem-dimethyl substitution was also established via the mutual correlations between the two methyl groups both also showing correlations with C-13, C-14 and C-15. Subsequent observations of the correlations of H-10/C-9; H₃-18/C-9, C-11 and C-12; H-12/C-13; H-8/C-9 and C-14; H-7/C-8; and H₂-20/C-5, C-6, and C-7, facilitated the construction of ring-B and ring-C as shown. The 12-O-tiglyl group was evidenced by the HMBC correlation from H-12 (δ_H 5.52) to its carbonyl carbon (δ_C 167.8), while the presence of 4-OH, 9-OH, 20-OH and 13-O-(2-methylbutyryl) groups was indicated by the chemical shifts of C-4, C-9, C-20 and C-13 (δ_C 72.6, 75.1, 66.6 and 65.9, respectively) and comparison with known tiglane analogues possessing the same substitution patterns.¹⁷ The structural characterization presented above explained all but the elements of O_2H which was identified to be a hydroperoxyl group at C-7 based on the chemical shift of the latter (δ_C 83.6) and excellent comparison to 12-O-decanoyl-7-hydroperoxy-phorbol-5-ene-13-acetate having the same 7- O_2H functionality.¹⁸

The relative configuration of **1** was characterized mainly by NMR comparison to 12-O-decanoyl-7-hydroperoxy-phorbol-5-ene-13-acetate¹⁸ and examination of NOESY data (Fig. 1B). The

only differences between the two compounds were attributable to the O-substituents at C-12 and C-13 and the remaining structural parts displayed highly comparable NMR data, which indicated that the two molecules possessed the same relative configuration. Moreover, the NOESY correlations of H-10/H-7, H-7/9-OH, and 9-OH/H-11 suggested that they were *pseudo*-axially bonded and α -oriented, while those of H-8/H-11, H-8/H₃-17, and H-7/H-14 supported the β -direction of H-8, H-11 and the cyclopropyl moiety. The structure of **1** was thus unequivocally characterized and was named crotignoid A.

Compound **2** displayed a sodiated molecular ion peak at m/z 571.2507 $[M+Na]^+$ in the (+)-HRESIMS spectrum, consistent with a molecular formula of $C_{29}H_{40}O_{10}$ (calcd 571.2514). Analysis of the NMR data (Tables 1 and 2) revealed a tiglane scaffold for **2** showing high similarity to **1**. Compared with **1**, the absence of NMR signals for the 2-methylbutyryl group and the presence of those for an isobutyryl residue (δ_H 2.59 (m), 1.19 (d, 7.0) and 1.17 (d, 7.0)) supported a 13-O-isobutyl substitution in **2**. The HMBC correlation from H-12 (δ_H 5.50) to the tiglyl carbonyl carbon (δ_C 167.9) further supported the above-mentioned assignment. The remaining NMR resonances for the two compounds were almost superimposable indicating a common relative stereochemistry. Thus the structure of **2** was elucidated as shown. The presence of the hydroperoxyl group in **1** and **2** is interesting, and this functionality has also been found in other types of natural products.^{19–21} However, the possibility that **1** and **2** might be handling artifacts could not be totally excluded, since the formation of hydroperoxyl derivative from phorbol esters was once reported.^{22,23}

Compounds **3** and **4** had molecular formulae of $C_{30}H_{42}O_9$ and $C_{29}H_{40}O_9$ as deduced from the HRESIMS ions at m/z 591.2792 $[M+HCO_2]^-$, calcd 591.2811) and 555.2568 $[M+Na]^+$, calcd 555.2565), respectively. The NMR data (Tables 1 and 2) of **3** exhibited high similarity to those of **1** with major differences occurring to signals around C-7. Compared with **1**, the markedly up-field shifted C-7 resonance ($\Delta\delta_C$ 13.3) of **3** suggested the presence of a 7-OH instead of the hydroperoxyl group in the former. The relative stereochemistry of **3** was assigned to be identical with that of **1** based on the same proton coupling patterns and close coupling constants, and this was also confirmed by acquisition of NOESY data (Fig. S20, Supplementary data). The NMR data of **4** (Tables 1 and 2)

Table 1
 1H NMR data of compounds **1–6** (500 MHz, $CDCl_3$)

No.	1	2	3	4	5	6
1	7.65 brs	7.64 brs	7.61 brs	7.61 brs	7.67 brs	7.67 brs
5	6.39 d (2.4)	6.39 d (2.4)	6.13 brs	6.09 d (1.6)	6.90 brs	6.91 brs
7	4.78 dd (9.2, 2.4)	4.78 dd (9.0, 2.4)	4.78 brd (9.7)	4.82 dd (9.6, 1.6)		
8	2.88 dd (9.2, 5.5)	2.88 dd (9.0, 5.5)	2.61 dd (9.7, 5.3)	2.55 dd (9.6, 5.4)	3.75 d (5.4)	3.76 d (5.4)
10	2.99 m	2.99 m	3.05 m	3.08 m	3.32 brs	3.31 brs
11	2.22 dq (10.2, 6.6)	2.23 dq (10.2, 6.5)	2.21 dq (10.2, 6.4)	2.18 dq (10.3, 6.5)	2.24 dq (10.2, 6.5)	2.24 dq (10.3, 6.4)
12	5.52 d (10.2)	5.50 d (10.2)	5.47 d (10.2)	5.47 d (10.3)	5.49 d (10.2)	5.49 d (10.3)
14	1.62 d (5.5)	1.62 d (5.5)	1.50 d (5.3)	1.54 d (5.4)	1.83 d (5.4)	1.84 d (5.4)
16	1.29 s	1.28 s	1.19 s	1.23 s	1.20 s	1.19 s
17	1.26 s	1.25 s	1.22 s	1.24 s	1.24 s	1.23 s
18	0.91 d (6.6)	0.91 d (6.5)	0.89 d (6.4)	0.90 d (6.5)	0.94 d (6.5)	0.94 d (6.4)
19	1.79 brs	1.79 brs	1.75 brs	1.79 brs	1.84 brs	1.84 brs
20	4.22 d (11.2)	4.23 d (11.3)	4.32 s	4.31 d (12.4)	4.40 brd (14.0)	4.39 brd (14.0)
	4.46 d (11.2)	4.45 d (11.3)		4.37 d (12.4)	4.29 brd (14.0)	4.29 brd (14.0)
9-OH	6.15 s	6.14 s	6.02 s	6.00 s	6.29 s	6.22 s
3'	6.84 m	6.84 m	6.82 m	6.83 m	6.83 m	6.84 m
4'	1.83 brs	1.83 brs	1.82 brs	1.83 brs	1.84 brs	1.84 brs
5'	1.80 brd (7.0)	1.80 brd (7.1)	1.78 brd (7.1)	1.80 brd (7.1)	1.80 brd (7.2)	1.80 brd (7.3)
2''	2.39 m	2.59 m	2.37 m	2.58 m	2.60 m	2.39 m
3''	1.46 m	1.17 d (7.0)	1.44 m	1.17 d (7.0)	1.18 d (6.9)	1.16 t (7.5)
	1.74 m		1.71 m			
4''	1.14 d (7.0)	1.19 d (7.0)	1.13 d (7.1)	1.19 d (7.0)	1.20 d (7.0)	
5''	0.95 t (7.4)		0.93 t (7.4)			

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