



Synthetic epoxy-mycolic acids



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ABSTRACT

We report the synthesis of single enantiomers of epoxy-mycolic acids containing an α -methyl-*trans*-alkene or a *cis*-cyclopropane with structures that match those of major isomers of such molecules present in complex mixtures in Mycobacteria such as *Mycobacterium fortuitum* or *Mycobacterium smegmatis*

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1. Introduction

Mycolic acids (MAs), **1** (Fig. 1), are major constituents of the cell envelope of *Mycobacterium tuberculosis* and other mycobacteria, some of which are pathogenic to animals and humans.^{1–3} Their structure and biosynthesis has been reviewed,⁴ and a number of structural and stereochemical relationships examined.⁵ Their presence is thought to be linked to the resistance of these organisms to most current antibiotics and other chemotherapeutic agents.⁶

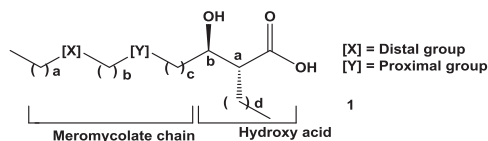
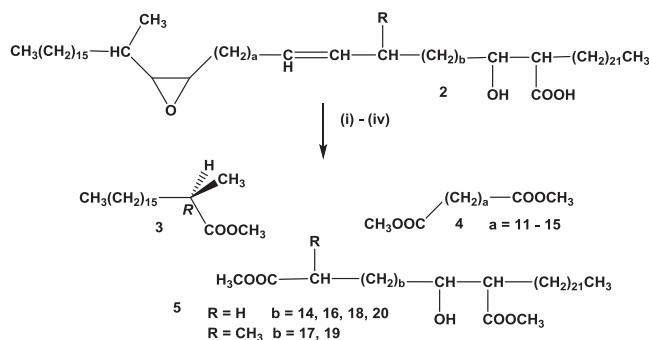


Fig. 1. The typical structure of a mycolic acid.

The two stereocentres in the α and β -positions relative to the carboxylic group have both been found to be in the *R*-configuration for all the mycolic acids examined, irrespective of the other functional groups.^{7,8} In each case 'a–d' represent long alkyl chains, and generally each *Mycobacterium* contains a mixture of several homologues. In the common classes of MA, the proximal group is

often a cyclopropane or an alkene and the distal group is a cyclopropane (α -MA), an α -methoxy- β -methyl fragment (methoxy-MA) or an α -keto- β -methyl fragment (keto-MA).^{2,3} In 1981, Daffé et al. reported the identification of a new kind of mycolic acid in *Mycobacterium fortuitum* containing an α -methyl epoxy-group at the distal position and an alkene at the proximal position (**2**, R=H) (Scheme 1).⁹ Minnikin et al. later described the presence of similar molecules in *M. fortuitum*, *Mycobacterium farcinogenes*, *Mycobacterium senegalense*, '*Mycobacterium peregrinum*', and *Mycobacterium smegmatis*;^{10–12} in these cases the major isomers had a *cis*-alkene at the proximal position, but there was a minor component (ca. 30% by NMR spectroscopy) (**2**, R=Me) containing a proximal α -methyl-*trans*-alkene; the major isomer of the latter contained 78 carbons; the epoxy-MA had carbon skeletons similar to those in methoxy-, keto- or α -MA described earlier.¹⁰

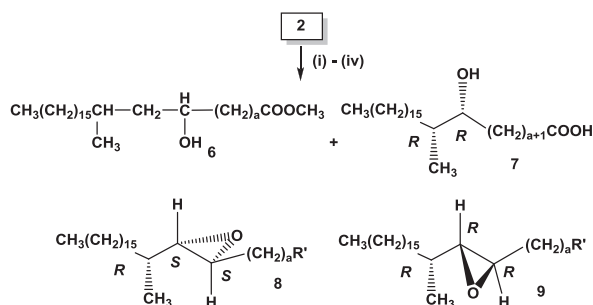


Scheme 1. (i) Acetolysis; (ii) saponification; (iii) oxidative cleavage; (iv) methylation.

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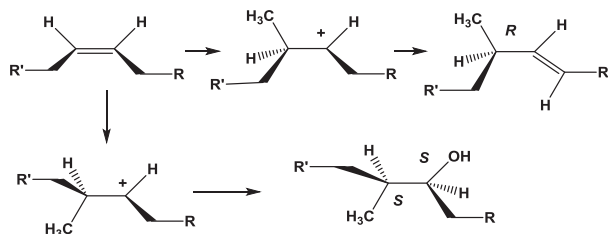
Epoxy-MAs are also present in *Mycobacterium chitae*,^{13,14} *Mycobacterium giae*,¹³ *M. peregrinum*,^{11–13} *Mycobacterium porcinum* and *M. senegalense*,¹⁵ and in a mutant strain of *M. tuberculosis*.¹⁶ In the final case, these included epoxy-MA with a *cis*-cyclopropane at the proximal position. Epoxy-mycolic acids have also been detected directly by MALDI-TOF mass spectrometry; the major isomers containing a *trans*-alkene at the proximal position are reported to be C₇₈ and C₈₀ molecules.^{16–18} Studies using *M. smegmatis* have identified the function of the protein MSMEG0913 in adding the methyl branch adjacent to both an alkene and a cyclopropane at the proximal position of epoxy-mycolates to produce the *trans*-homologues,¹⁸ and examined other aspects of the biochemistry of such species.^{19–22} The relative and absolute stereochemistry of epoxy-mycolates containing a proximal *cis*- or α -methyl-*trans*-alkene has been probed by two methods (Scheme 1).²³ Firstly, opening of the epoxide **2** by acetolysis, then saponification and oxidative cleavage of both the derived 1,2-diol and the alkene leads to three products, including (*R*)-ester **3**.

Secondly, reductive ring-opening of the epoxide **2** followed by oxidative cleavage of the proximal alkene, saponification and methylation led to the two acids **6** and **7** (Scheme 2). The latter was shown to have *R,R*-stereochemistry by comparison to a model compound. On this basis the authors assigned all the stereocentres in the epoxy-mycolic acid as *R*.²³ However, it seems clear in fact that the result actually suggests the epoxy fragment is *R,S,S* as in **8** rather than *R,R,R* as in **9**, the priorities in the epoxide being different from those in the ring-opened alcohol.



Scheme 2. (i) Reduction; (ii) oxidative cleavage; (iii) saponification; (iv) methylation.

What is arguably most interesting is that the methyl group adjacent to the epoxide is in the *R*-configuration, whereas that adjacent to the hydroxyl, methoxy or keto-groups at the distal position of the corresponding mycolic acids is of *S*-configuration. It is attractive to propose that the various types of mycolic acid are formed through a formal intermediate carbocation, formed by methylation of a *cis*-alkene by SAM (Scheme 3). Methylation from the *Re* or *Si* face of the alkene at the alkylated carbon could then lead to a divergence in stereochemistry.



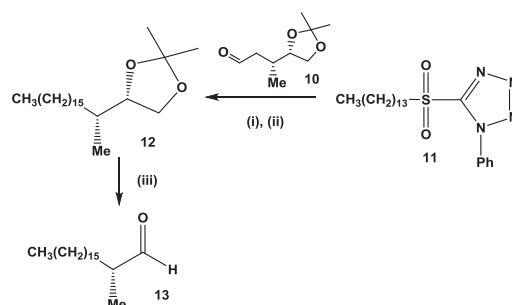
Scheme 3. Possible formal mechanism for production of *S*- and *R*-methyl branches by methylation of a *cis*-alkene by SAM.

We have reported the synthesis of an α -mycolic acid,^{24,25} of methoxymycolic acids with either absolute stereochemistry at the *cis*-cyclopropane or α -methyl- β -methoxy fragment,²⁶ and of keto mycolic acids.^{27,28} We have also reported the synthesis of methoxy

and keto mycolic acids containing an α -methyl-*trans*-cyclopropane unit,^{29,30} of *cis*-alkene mycolic acids,³¹ and of hydroxy- and keto mycolic acids containing an (*R*)- α -methyl-*trans*-alkene unit.³² We now report in full the synthesis of two stereoisomeric epoxy-mycolic acids containing an (*R*)- α -methyl-*trans*-alkene at the proximal position; these have the chain lengths reported for one mycolic acid in *M. fortuitum*,^{4,5} and the total carbon number is consistent with those observed for one component of a mixture from *M. smegmatis* by MALDI mass spectrometry.^{17,18} This has been reported briefly earlier.³³ We also report the synthesis of an epoxy-mycolic acid containing a *cis*-cyclopropane.

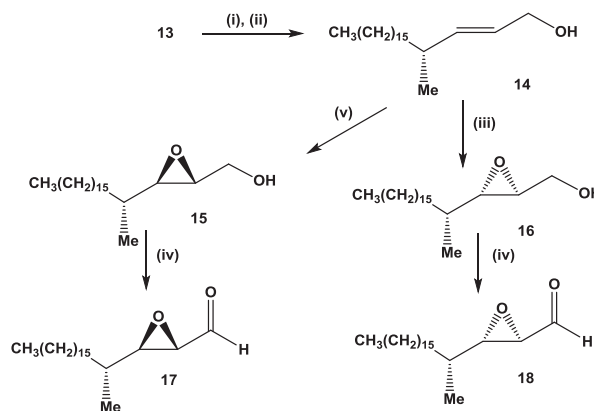
2. Results and discussion

The aldehyde **13** was first prepared from the known aldehyde **10**,^{28,34,35} with sulfone **11** (see Supplementary data) and base in a modified Julia-Kocienski reaction, followed by hydrogenation to give **12**. Deprotection and oxidative cleavage of the acetal **12** led to **13** (Scheme 4).



Scheme 4. (i) **10**, LiN(SiMe₃)₂ (LiBSA), THF, –10 °C (67%); (ii) H₂, Pd/C, ethanol, (92%); (iii) periodic acid, dry ether (77%).

A Wittig reaction and then reduction with DIBAL led to the *trans*-alcohol **14** (Scheme 5). Asymmetric epoxidation using Sharpless conditions provided the two diastereoisomers **15** and **16**.^{36,37} The two epoxides showed $[\alpha]_D^{22}$ of +18.1 and –21.2, respectively. The literature value for (*2R,3R*)-dodecyloxiranyl-methanol is –25.5.³⁸



Scheme 5. (i) Ph₃P=CHCOOMe, toluene (65%); (ii) DIBAL, CH₂Cl₂ (95%); (iii) *l*-(+)-diethyl tartrate, Ti(Oi-Pr)₄, *t*-BuOOH, CH₂Cl₂, –20 °C (75%); (iv) PCC, CH₂Cl₂ (59% **18**), (60% **17**); (v) *D*-(-)-diethyl tartrate, Ti(Oi-Pr)₄, *t*-BuOOH, CH₂Cl₂, –20 °C (67%).

Oxidation of each alcohol led to the corresponding aldehyde. The aldehyde **18** was chain extended by reaction with the sulfone **19** (see Supplementary data) and base, followed by saturation of the derived alkene using di-imide. The derived bromide **20**, $[\alpha]_D^{22}$ –13.1 (*c* 1.2, CHCl₃) ($[M]_D$ –76) {molecular rotation, $[M]_D$ = (molecular weight × specific rotation)/100}, was then converted into the corresponding sulfone **21**. In the same manner, diastereoisomer **17** was converted into **22** (Scheme 6). The molecular rotations of the two

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