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Lissoclibadins 1–3, three new polysulfur alkaloids, from the ascidian *Lissoclinum* cf. *badium*

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Dedicated to the memory of Dr. Kenneth L. Rinehart and Dr. Katsumi Kakinuma

Abstract—Three new polysulfur alkaloids, lissoclibadins 1 (1)–3 (3), were isolated from the ascidian *Lissoclinum* sp. (cf. *L. badium* Monniot, F. and Monniot, C., 1996). The structures of 1–3 were assigned on the basis of their spectral data, and the computational modeling study was utilized for 1. Compound 1 had a trimeric structure of three identical aromatic anime moieties connected through two sulfide and a disulfide bonds. Compounds 2 and 3 were dimeric structures of the same unit as that of 1 connected through a sulfide and disulfide bonds (2) and two sulfide bonds (3). Compounds 1 and 2 inhibited the growth of the marine bacterium *Ruegeria atlantica* (15.2 mm at 20 μg/disk and 12.2 mm at 5 μg/disk, respectively) and 2 showed antifungal activity to *Mucor hiemalis* (13.8 mm at 50 μg/disk). Compounds 1–3 were cytotoxic against HL-60 (IC₅₀=0.37, 0.21, and 5.5 μM, respectively).

1. Introduction

Many interesting bioactive compounds have been obtained from ascidians (tunicates). Aromatic alkaloids possessing polysulfide structures have been isolated from ascidians of the genera Lissoclinum, $^{1-6}$ Eudistoma, 4 and Polycitor. More than 10 monomeric cyclic polysulfides $^{1-4,6,7}$ and four dimeric polysulfides 2,5,8 have been reported. These compounds presented various biological activities, for example, antifungal activity, $^{1-3,7}$ antibacterial activity, 3,7 cytotoxicity, 1,3,8 antimalarial (Plasmodium falciparum) activity, 3 inhibition of protein kinase C, 4,5 and inhibition of IL-8 R α and R β receptors. 5

In the course of our study on the bioactive metabolites from marine organisms, we found that the ethanol extract of the ascidian Lissoclinum sp. (cf. L. badium Monniot, F. and Monniot, C., 1996)⁹ collected at Manado, Indonesia, showed strong antimicrobial activity against the fungus Mucor hiemalis and the marine bacterium Ruegeria atlantica. Bioassay-guided separation gave three new polysulfide aromatic alkaloids, named lissoclibadins 1 (1), 10 2 (2), and 3 (3), together with two known dimeric alkaloids, lissoclinotoxins E (4) and F (5), and two known monomeric 3,4-dimethoxy-6-(2'-N,Ncompounds, dimethylaminoethyl)-5-(methylthio)benzotrithiane (6) and N,N-dimethyl-5-(methylthio)varacin (7). We have reported the structure of 1 in the previous communication fo and describe here the isolation, structure elucidation, and biological activity of three new lissoclibadins 1-3 (1-3) and compounds 4–7.

2. Results and discussion

2.1. Ascidian and isolation of alkaloids

Lissoclinum cf. badium was collected at Manado, Indonesia

Keywords: Tunicate; Lissoclinum sp.; Lissoclinotoxin; Polysulfur compound; Structure assignment.

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in 2003 and 2004 and extracted with ethanol. The ethanol extract was redissolved in MeOH–H₂O (9/1) and extracted with n-hexane. The aqueous MeOH layer was diluted with water and extracted with n-BuOH. The BuOH extract showed moderate antimicrobial activity and gave 1, 2, 3, and 5 by ODS and SiO₂ column chromatographies followed by HPLC. The hexane extract revealed strong antifungal and antibacterial activities and was separated by SiO₂ column chromatography followed by HPLC to yield 6 and 7. Compound 4 was isolated from the ascidian collected in 2004 together with 1–3 and 5–7.

Structures of three known compounds **5**–**7** were assigned on the basis of their spectral data and comparison with those for the reported values.^{4,8}

2.2. Structure of lissoclibadin 1

Lissoclibadin 1 (1) was isolated as a Tris–TFA salt. The molecular weight (887) and formula ($C_{39}H_{57}N_3O_6S_7$) were deduced from HRFABMS and NMR data (Table 1). Three sets of 1H and ^{13}C NMR signals were observed in the NMR

spectra of **1** and assigned to three identical aromatic amine moieties by the analysis of ¹H–¹H COSY, HMQC, HMBC, ROESY, and NOESY spectra.

The geminal couplings and connectivity of two methylene groups at the 7 and 8 positions were revealed by ¹H-¹H COSY spectrum. HMBC correlations were detected from H₂-7 to three aromatic carbon signals (C-1, 5, and 6) and C-8. The ¹³C signal assigned as C-5 showed an HMBC correlation from a methyl singlet of SMe (H₃-11). This SMe revealed an NOE with one of two methoxy methyl singlets due to H₃-10 in the NOESY spectrum. The ¹H signal of H₃-10 showed an HMBC correlation to an aromatic carbon signal (C-4) and an NOE with the H₃-9 (OMe) singlet, which had an HMBC correlation to an aromatic carbon signal (C-3). HMBC correlations were observed from H₂-8 to NMe₂ (C-12) and vice versa. Therefore, three sets of ¹H and ¹³C NMR signals were assigned except each one aromatic carbon signal, which had no cross peak in any 2D NMR spectra and was deduced as C-2.

¹H and ¹³C NMR data for three identical aromatic units

Table 1. ¹³C (150 MHz) and ¹H NMR (600 MHz) data for the three units (TFA salts) in 1 (CD₃OD)

C#	Unit 1		Unit 2		Unit 3		HMBC
	¹³ C (δ)	¹ H (δ, m)	¹³ C (δ)	¹ H (δ, m)	¹³ C (δ)	¹ H (δ, m)	_
1	142.2ª		140.3 ^b		140.3°		
	142.3 ^d		141.2 ^d		137.9 ^d		
	156.9		158.8		159.0		
	153.6		153.5		158.2		
	135.1		134.4		133.6		
	136.9 ^a		134.9 ^b		135.0°		
7	31.3	3.80, m	30.7	3.71, m	30.6	3.68, m	1, 5, 6, 8
		4.01, m		3.92, m		3.85, m	1, 5, 6, 8
8	58.3	3.00, m	58.3	3.25, m	58.2	3.23, m	7, 12
		3.27, m		3.36, m		3.32, m	7, 12
	61.1	3.99, s	60.5	3.70, s	60.8	3.67, s	3
0	60.8	3.34, s	60.4	3.15, s	62.3	3.88, s	4
1	19.3	2.53, s	19.1	2.46, s	19.1	2.44, s	5
2	43.4	3.06, br s	43.4	3.06, br s	43.4	2.96, br s	8, 12

^{a,b,c,d} Signals are interchangeable within the same letters.

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