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Tetrahedron Letters

Tetrahedron Letters 49 (2008) 44-47

## Highly selective imine-linked fluorescent chemosensor for adenine employing multiple hydrogen bonding

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> Received 12 September 2007; revised 1 November 2007; accepted 5 November 2007 Available online 7 November 2007

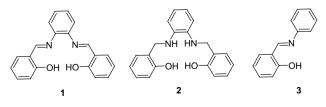
Abstract—We investigated an imine-linked fluorescent receptor bearing both the hydrogen bond donor and the hydrogen bond acceptor motifs as recognition sites in the design of the receptor. The recognition behavior of the receptor toward various nucleobases was evaluated in CH<sub>3</sub>CN/H<sub>2</sub>O (95:5, v/v) solution. The receptor showed significant changes in fluorescent intensity with adenine, but it showed no such changes on the addition of other nucleobases. © 2007 Elsevier Ltd. All rights reserved.

Over the last few years, model studies in molecular recognition have contributed to the development of a wide range of novel molecular devices.<sup>1,2</sup> Within this context, nucleobase recognition is an important challenge in supramolecular chemistry because of its many biological implications.<sup>3–9</sup> To achieve this goal in abiotic receptors, a receptor binding unit for the recognition of a guest and a signaling unit are incorporated into the receptor molecule.<sup>10-16</sup> However, binding of such a receptor in H<sub>2</sub>O finds an important competitor in the solvent itself, which retards the interaction between the host and the guest through forces like H-bonding. Therefore, the alignment of receptor binding sites on a receptor platform must achieve complementarily-binding interactions toward a targeted guest. In other words, the chemical and steric features given by a guest molecule have to be matched by a sufficiently predisposed host.<sup>17</sup>

As part of our ongoing studies on a simple and easyto-make receptor system,  $^{18-21}$  here we present an adenine binding study of a receptor containing imine linkage. To date, there have been no reports in which an iminelinked receptor is used for the recognition of adenine. The strategy for the design of the receptor is based upon the idea that the receptor has both hydrogen bond donor and hydrogen bond acceptor sites to make a complex effectively with adenine.<sup>22</sup> Receptors 1-3 were prepared by following the procedures in the literatures (Scheme 1).<sup>23,24</sup>

Receptor 1 displayed a maximum at 410 nm in its fluorescence spectrum that was recorded with its 10  $\mu$ M concentration in CH<sub>3</sub>CN/H<sub>2</sub>O (95:5, v/v) when excited at 365 nm. Receptor 2 lacks any such type of emission with its 10  $\mu$ M concentration in CH<sub>3</sub>CN/H<sub>2</sub>O (95:5, v/v). Therefore, the imine chromophore of receptor 1 is responsible for the observed emission. The changes in fluorescence intensity of 1 upon the addition of a particular anion are shown in Figure 1, and the fluorescence ratio  $(I_0 - I)/I_0$  is displayed in Figure 2.

As can be seen from Figures 1 and 2, it is clear that there is a marked enhancement in fluorescence intensity of receptor 1 upon the addition of adenine solution. There were no such significant changes in the fluorescence intensity of 1 upon the addition of guanine, thymine, and uracil. This shows that receptor 1 is highly selective



Scheme 1.

*Keywords*: Fluorescent receptor; Molecular recognition; Hydrogen bonding; Nucleobase.

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<sup>0040-4039/\$ -</sup> see front matter © 2007 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2007.11.027

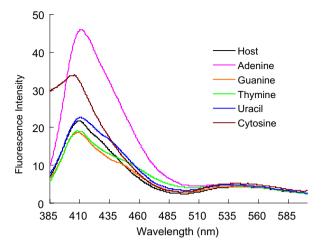
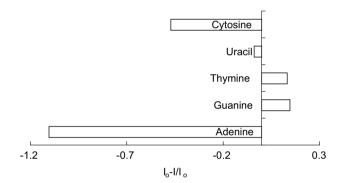


Figure 1. Changes in fluorescence intensity of receptor 1 (10  $\mu$ M) upon the addition of 2.0 equiv of a particular guest in CH<sub>3</sub>CN/H<sub>2</sub>O (95:5, v/v) with excitation at 365 nm.



**Figure 2.** Fluorescence ratio  $(I_0 - I/I_0)$  of receptor **1** (10  $\mu$ M) at 410 nm upon the addition of 2.0 equiv of a particular guest in CH<sub>3</sub>CN/H<sub>2</sub>O (95:5, v/v).

in its response to adenine in comparison to other nucleobases. The receptor- nucleobase binding strength follows the order of adenine > cytosine  $\gg$  guanine  $\approx$ 

thymine  $\approx$  uracil. This implies that sp<sup>2</sup> nitrogens of imine linkage of receptor 1 act as hydrogen bond acceptors to form additional hydrogen bonds with the N–H of adenine and cytosine in addition to the hydrogen bonding between receptor O–H and nitrogens of the nucleobase.<sup>25–28</sup>

To confirm our point, we investigated the recognition properties of another compound 3, which resembles the single pod of receptor 1. We selected a 20  $\mu$ M concentration of compound 3 (a 20  $\mu$ M concentration of 3 has approximately the same number of binding sites as that of a 10  $\mu$ M concentration of 1) to investigate the recognition properties. No significant changes in the fluorescence intensity were observed in the typical experiment. This proved that although 1 and 3 have the same type of binding sites only an appropriate size of the pseudocavity of 1 can bind adenine, and a guest is believed to be bound cooperatively in the cavity of 1.

To learn more about the properties of **1** as a receptor for adenine, fluorescence titrations were carried out. The fluorescence intensity of a 10 µM solution of 1 was enhanced as the concentration of adenine increased as shown in Figure 3. The fluorescence enhancement of receptor 1 may be a consequence of an increase in the conformational restriction of receptor 1 upon the complexation of receptor with adenine. According to this system, as a consequence of the guest coordination, the rigidity of the formed complex increases making the nonradiative decay from the excited state less probable; consequently, the emission intensity increases.<sup>29-31</sup> The association constants  $K_a$  of 1 for adenine and cytosine were calculated on the basis of Benesi-Hildebrand plot,<sup>32</sup> and it was found to be  $(3.8 \pm 0.3) \times 10^4 \text{ M}^{-1}$ and  $(2.0 \pm 0.2) \times 10^3 \text{ M}^{-1}$ , respectively. Thus, receptor 1 can be used for selective recognition of adenine and it can detect adenine as little as a low concentration of 2.1  $\mu M.^{33}$  The stoichiometry of the complex formed was determined by Job's plot,<sup>34</sup> and it turned out to be 1:1.

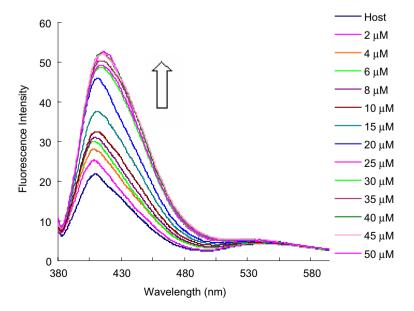


Figure 3. Fluorescence intensity changes of receptor 1 ( $10 \,\mu$ M) upon the addition of adenine ( $0-50 \,\mu$ M) in CH<sub>3</sub>CN/H<sub>2</sub>O (95:5, v/v).

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