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### Applied Surface Science

journal homepage: www.elsevier.com/locate/apsusc

# Secondary ion mass spectrometric signal enhancement of phosphatidylcholine dioleoyl on enlarged nanoparticles surface

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#### ARTICLE INFO

Article history: Received 22 January 2014 Received in revised form 17 July 2014 Accepted 18 July 2014 Available online 3 August 2014

*Keywords:* ToF-SIMS Lipid Signal enhancement Nanoparticles Phosphatidylcholine dioleoyl

#### 1. Introduction

The development of new methods for obtaining information on the organization and dynamics of assemblies is an important frontier in structural biology and mass spectrometry imaging is becoming a significant complementary technique [1,2]. Secondary ion mass spectrometry coupled with a time-of-flight mass spectrometer (ToF-SIMS) imaging technique, using a cluster ion beam as primary bombardment particles, is now an analytical tool of reference for high lateral resolution imaging  $\sim 1 \,\mu m$  of biological surfaces, lateral resolution better than 100 nm is possible with specialized instruments [2]. The majority of SIMS analyses of lipid membranes has been performed using ToF-SIMS. As the large molecular fragments often have distinctive masses, labeling is, in principle, not required for component identification. Winograd and coworkers [3,4] have pioneered the ToF-SIMS analysis of domain formation in Langmuir-Blodgett lipid monolayers deposited onto self-assembled monolayers of alkane thiols on gold.

While ToF-SIMS provides a glimpse of the composition of a surface, it is still a major challenge for ToF-SIMS analysis to overcome the low secondary ion yield in the high-mass region of the spectrum. Low signal intensity of high mass (m/z > 400-500) organic ions is a serious drawback of ToF-SIMS. Discovery cluster

#### ABSTRACT

A silicon wafer surface coverage of nanoparticles (NPs) can enhance the L- $\alpha$ -phosphatidylcholine dioleoyl (DOPC) signal intensity in time-of-flight secondary ion mass spectrometry (TOF-SIMS). A ToF-SIMS mass spectrometer was used with a pulsed primary beam of focused 30 keV Bi<sub>3</sub><sup>+</sup> ions. The signal enhancing effect has been studied for metallic (Ag, Au, Pb), semiconductor (TiO<sub>2</sub>), dielectric (SiO<sub>2</sub>) and hybrid (Au/TiO<sub>2</sub>NPs, core-shell Au/SiO<sub>2</sub>) nanoparticles. Ag NPs can attenuate secondary ions signal, whereas all other studied NPs show the signal enhancement. The emission of DOPC lipid secondary ions immobilized on core-shell Au/SiO<sub>2</sub>NPs was enhanced up to 42 times. This technique is a simple preparatory method enabling an overall increase in molecular lipid ions.

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ion sources (Au<sub>3</sub><sup>+</sup>, Bi<sub>3</sub><sup>+</sup>, SF<sub>5</sub><sup>+</sup>, C<sub>60</sub><sup>+</sup>) allowed to enhance signal intensity [5], because these primary ions open the potential for higher yields of large molecules. Sample modification is an alternative approach. Matrix-enhanced SIMS and metal-assisted SIMS allow to amplify TOF-SIMS signal intensity by organic matrix or metal deposition [6,7]. Metal nanoparticle deposition, which increases TOF-SIMS secondary ion yields of polymer molecules [8], involves exposing the sample surface to a solution of citrate-capped Ag or Au nanoparticles. As a polymer incorporates a submonolayer amount of the particles at the surface, the ToF-SIMS signals become meaningfully enhanced [9–13]. Recently the enhancement of the SIMS signal has been reported for the self-assembled layers of Au nanoparticles (AuNPs) with peptides [14,15] and for *E. coli* bacteria cell with ZnO NPs [16].

The aim of the present work is to compare the effect of different nanoparticles on the SIMS signal from thin lipid layer deposited on the self-assembles layer of nanoparticles at the silicon wafer. We report the enhancement effect of lipid molecular ion yield when particle size and material is varied: (a) metallic Ag, Au and Pb NPs; (b) oxides  $SiO_2$  and  $TiO_2$ ; (c) core–shell Au/SiO<sub>2</sub> NPs and Au nano-clusters photodeposited on the  $TiO_2$  nanoparticles.

#### 2. Materials and methods

#### 2.1. Materials

All descriptions of water below are referred to distilled water. Hydroxylamine hydrochloride NH<sub>2</sub>OH·HCl, sodium hydroxide







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NaOH, silver nitrate AgNO<sub>3</sub>, HAuCl<sub>4</sub>·3H<sub>2</sub>O, lead acetate (II) Pb (Ac)<sub>2</sub>·3H<sub>2</sub>O, sodium citrate Na<sub>3</sub>Cit, sodium borohydride NaBH<sub>4</sub>, octadecanethiol, tetraoctylammonium bromide, tetrapropoxysilane, were purchased from Aldrich (analytical grade). L- $\alpha$ -Phosphatidylcholine dioleoyl (1,2-dioleoyl-sn-glycero-3-phosphocholine, molecular weight – 786.11) was from Sigma–Aldrich. As TiO<sub>2</sub> nanoparticles the Aeroxide P25 were used. K<sub>2</sub>CO<sub>3</sub>, formaldehyde, toluene were purchased from Acros chemicals. Silicon wafers were from J&J Co., South Korea.

#### 2.2. Nanoparticles synthesis

#### 2.2.1. AuNPs type I

AuNPs were synthesized by citrate methods [17]. HAuCl<sub>4</sub> solution of 0.2 M was prepared. 1 mL HAuCl<sub>4</sub> solution was added to 94 mL water with stirring. The solution was heated to boiling with a hemisphere heating mantle under vigorous magnetic stirring. After boiling for 2 min, sodium citrate (353 mg) was rapidly added to the flask. The solution gradually turned gray–blue–crimson within a few minutes, indicating the formation of Au nanoparticles. The solution was kept boiling for additional 23 min. After that, the heating mantle was removed, and the solution was allowed to cool. The average particle size according to scanning electron microscope (SEM) measurements was 30 nm.

#### 2.2.2. AuNPs type II

Gold particles were synthesized by a two-phase synthesis method [18], with some modifications at temperature of 23 °C. Tetraoctylammonium bromide solution in toluene (0.06 M, 35 mL) was added to HAuCl<sub>4</sub> solution (0.03 M, 15 mL), followed by stirring. Noticeable color manifested all HAuCl<sub>4</sub> transition in the organic phase. Octadecanethiol (0.44 mM) was added. It served as the stabilizer of the nanoparticles in the organic phase. A freshly prepared solution of sodium borohydride (0.4 M, 12.5 mL) was slowly added dropwise to the reaction mixture under vigorous stirring continued for other 3 h. The organic phase separated and evaporated on a rotary evaporator to a volume of 10 mL. The octadecanethiol excess was removed by several cycles with toluene–isopropanol sedimentation centrifuge. The resulting precipitate was dissolved in 20 mL of chloroform. The NPs average size was 5 nm.

#### 2.2.3. AgNPs type I

AgNO<sub>3</sub> was preliminarily dissolved in water (0.01 M). NH<sub>2</sub>OH·HCl (12 mg), 1 M NaOH (0.33 mL), AgNO<sub>3</sub> solution (11 mL) were successively added with stirring to 100 mL of water. The solution was stirred for 2 h at a room temperature of 23 °C. As a result, the gray opalescent solution was obtained. The average particle size according to SEM (scanning electron microscope) measurements was 80 nm [19].

#### 2.2.4. AgNPs type II

The kind of sodium citrate reduction method was applied. Briefly,  $AgNO_3$  17 mg was preliminarily dissolved in water 100 mL. The solution was heated to boiling under vigorous magnetic stirring. Solution 4 mL of sodium citrate 1% was rapidly added to the flask. The reaction mixture is reflux during 30 min. The gray turbid solution was obtained. The NPs average size was 23 nm.

#### 2.2.5. AgNPs type III

Method sodium borohydride reduction has been used [20]. Two solutions sodium citrate 0.4 mM in 50 mL water and silver nitrate 0.4 mM in 50 mL water were prepared and cooled in an ice bath. Sodium borohydride solution (25 mM) were prepared in water and kept in ice bath. The silver nitrate and sodium citrate solutions were mixed together in a conical flask and vigorously magnetic stirred. Then, 30  $\mu$ L of the sodium borohydrite solution was added at once. After 10 min of stirring, the solution was heated slowly without boiling and heated at near-boiling temperature of 90 °C for a further 90 min. The solution was cooled to room temperature with stirring. Color was eventually yellow-brown opalescent. The average size of NPs was 5 nm.

#### 2.2.6. PbNPs

To 10 mL of water was added with stirring 1.8 mg of Pb(Ac)<sub>2</sub>·3H<sub>2</sub>O, Na<sub>3</sub>Cit<sub>3</sub>,1 mg and freshly prepared 0.1 M NaBH<sub>4</sub> solution of 4 mL. The solution was stirred under heating for 2 h. The color of the solution was white opalescent. The average particle size according to SEM was 30 nm.

#### 2.2.7. Au/TiO<sub>2</sub>

The deposition of Au nanoparticles on TiO<sub>2</sub> nanoparticles was realized by photocatalytic reduction of Au<sup>3+</sup> ions in aqueous TiO<sub>2</sub> suspension with methanol addition at 23 °C [21]. UV-A (Philips) operating at  $362 \pm 10$  nm with power of 8W was used for TiO<sub>2</sub> nanoparticles excitation and Au<sup>3+</sup> ions reduction. The light flux was  $6.9 \text{ mW/cm}^2$  and the exposition was 20 min. Methanol was used as an effective hole scavenger. The morphology of the particles was a TiO<sub>2</sub> core with a diameter of 20 nm with adjacent gold particles sized 5 nm according to the TEM.

#### 2.2.8. SiO<sub>2</sub>NPs and core-shell Au/SiO<sub>2</sub>NPs

Synthesis includes a sequence of several stages: (1) preparation of spherical SiO<sub>2</sub> particles with a narrow size distribution; (2) chemical modification of the  $SiO_2$  surface by amino groups; (3) synthesis seed gold nanoparticles; (4) binding seed gold nanoparticles to the surface of amino groups; (5) growth of continuous gold shell on the particle surface. Synthesis temperature was 23 °C. SiO<sub>2</sub> core synthesis: therefore, into 6 mL of isopropanol containing 0.77 mL aqueous ammonia solution (25%) there was added dropwise 0.42 mL tetrapropoxysilane (C<sub>3</sub>H<sub>7</sub>O)<sub>4</sub>Si, magnetically stirred for 2h and standing overnight. The product was washed twice by centrifuging in ethanol at 3000 min<sup>-1</sup> and was reslurried in 6 mL of isopropanol. Spherical silica particles of 460 nm were formed. The SiO<sub>2</sub> nanoparticles of 250 nm were synthesized in similar manner with different precursor concentrations. This technique gives a fairly narrow particle size distribution. These silica particles were functionalized by 3-aminopropyl-triethoxysilane (APTES). Tetra-hydroxymethyl phosphonium chloride (THPC) was used as a reducer to prepare 2-3 nm Au seed nanoparticles. 0.1 mL of SiO<sub>2</sub>/APTES suspension was mixed with 20 mL of THPC gold colloid, followed by 4h stirring and held for days. The precipitate was washed twice with water and redispersed in 5 mL of water. To initiate the growth of the gold shell, the precursor silica particles covered with the small gold clusters were added to the aged  $HAuCl_4/K_2CO_3$  solution. The ratio of the amount of the precursor particles added to the volume of the gold salt solution depended on the intended thickness of the gold shells. It was calculated assuming all added HAuCl<sub>4</sub> would reduce to gold shells around the silica. 37% formaldehyde was used as a reducer [22,23]. In the present work SiO<sub>2</sub> and Au/SiO<sub>2</sub> NPs were sensitized with diameter of SiO<sub>2</sub> core D = 250 nm and D = 518 nm, the Au shell layer was about 20 nm.

#### 2.3. Sample preparation

 $L-\alpha$ -Phosphatidylcholine dioleoyl (DOPC) solution in ethanol (600 mM) was used for the preparation of lipid films on a silicon wafer surface. To prepare control samples 25  $\mu$ L of lipid solution was deposited on a silicon wafer and then dried under argon. For sample preparation a 50  $\mu$ L NPs suspension (~100 ppm) was dropped on a wafer and dried under argon. After that 25  $\mu$ L of lipid

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